Bulletin of Environment, Pharmacology and Life Sciences Bull. Env. Pharmacol. Life Sci., Special Issue [1]2022 : 339-345 ©2022 Academy for Environment and Life Sciences, India Online ISSN 2277-1808 Journal's URL:http://www.bepls.com CODEN: BEPLAD ORIGINAL ARTICLE



## Antioxidant Potential of water soluble crude sulfated polysaccharide of *Sargassum polycystum*

#### Pratapavarma Digala1, Manoj Dhanraj2, Vincent Savariar1\*

1- Research scholar, Advanced zoology and Biotechnology, Loyola College, Nungambakkam, Chennai
2 - Assistant Professor, Department of Zoology, Madras Christian College, Tambaram, Chennai
1\* - Associate Professor, Advanced zoology and Biotechnology, Loyola College, Nungambakkam, Chennai
<u>Email: kmdmanoj@gmail.com</u>

#### ABSTRACT

The consumption of sea algae is widespread around the world. Brown algae are one of the seaweeds that have been approved for human consumption. These plants have a wide range of possible biological actions and contain essential phytochemical elements. Bioactive compounds from marine alage have been studied in a variety of locations across the world. Polysaccharides and their sulfated derivatives known as carrageenan, which are utilised as antigens and anticoagulants, were among the first bioactive ingredients discovered in marine algae. Sargassum polycystum is one of the important species belonging to the genus Sargassum and a wide range of bioactive properties have been reported. The present study investigated the presence of phytochemical constituents and antioxidant activity of Sargassum polycystum on the DPPH, FRAP and ABTS Assay. The extraction of the water soluble crude Sulfated polysaccharide (WSCSPs) from Sargassum polycystum was performed as described by with minor modifications. Phytochemical screening of the extracts was carried out according to the standard methods. WSCSPs from Sargassum polycystum were investigated for their DPPH radical scavenging activity, FRAP reducing power activity and ABTS radical scavenging activity was performed at the concentration ranging from  $50 - 250 \mu g/ml$ . From the phytochemical screening, it was observed that the WSCSPs from Sargassum polycystum revealed the presence of alkaloids, flavonoids, steroids, tannins, phenols, terpenoids and saponins. WSCSPs from Sargassum polycystum exhibit remarkable antioxidant potential through DPPH. FRAP and ABTS Assay. These findings show that the phytoconstituents in the extracts are responsible for free radical scavenging capacity. Seaweeds have gotten a lot of attention as a source of natural antioxidants, and they've been found to be a good source. The results of the present study confirmed that Sargassum polycystum may be rich sources of phytoconstituents which can be isolated and further screened for various biological activities. Keywords: Phytochemical screening, Antioxidant activity, DPPH, FRAP and ABTS

Received 19.02.2022

#### Revised 21.03.2022

Accepted 02.04.2022

## INTRODUCTION

Herbal medicines have been utilized to treat sickness symptoms since ancient times. Despite the significant improvements in modern medicine over the last few decades, plants continue to play a vital role in health care. However, medicinal plants and seaweed have sparked a lot of attention due to their long history of usage in folk medicine along with their preventive effects, particularly in developing nations [1]. Antioxidant capabilities have been studied in a large range of seaweed. Natural antioxidants, whether in the form of raw extracts or chemical components, are extremely effective in preventing oxidative stress-related damage. Exogenous substances and endogenous metabolic activities in the human body produce free radicals, or highly reactive oxygen species. These are capable of oxidizing biomolecules such as nucleic acids, proteins, lipids, and DNA, and can cause neurological problems, cancer, emphysema, cirrhosis, atherosclerosis, arthritis, and other degenerative diseases [2]. Antioxidants are substances that stop free radicals from attacking cells and thereby lessen the chance of disease. Almost all organisms are protected from free radical damage to some extent by antioxidant substances such as ascorbic acid, tocopherol, phenolic acids, polyphenols, flavonoids, and glutathione, as well as enzymes such as superoxide dismutase and catalase. Natural antioxidants are currently receiving a lot of interest as a way to protect the human body, particularly brain regions, from oxidative damage caused by free radicals [3]. Seaweed extract exhibits various biological activities as a result of its phytochemical content of bioactive compounds like antioxidants, anti-cancer agents, antiproliferative, antimicrobial properties, and treatments for cardiovascular diseases and ailments like atherosclerosis, hyperlipidemia, hypertension, and thyroid diseases [4-5]. The present study has been conducted to evaluate the antioxidant activity of *Sargassum polycystum*.

*Sargassum* species can be found in tropical and subtropical areas of the world, and they are known to produce metabolites of various structural classes, including terpenoids, polysaccharides, polyphenols, sargaquinoic acids, sargachromenol, plastoquinones, steroids, glycerides, and others, which have a variety of therapeutic properties. The metabolites meroterpenoids, phlorotanins and fucoidans are isolated from *Sargassum* sp. *Sargassum polycystum* is a marine alga with powerful antioxidant and anticancer properties. Chloroform extract of *Sargassum polycystum* demonstrated the strongest cytotoxicity against cervical HeLa cells which is potential to be developed as a candidate for new anticervical cancer agents [6]. Therefore, the present investigation was undertaken to elucidate the antioxidant activity of water soluble crude sulfated polysaccharide of *Sargassum polycystum* on the DPPH, FRAP and ABTS Assay.

## MATERIAL AND METHODS

#### Seaweed sample collection

The brown seaweed *surgassum polycystum* was collected from the Mandapam, Gulf of Mannar (Lat.09° 17'N; Long.79° 08'E), Rameshwaram, Tamil Nadu, India. The collected seaweed sample was washed first in seawater and then followed by distilled water to remove the unwanted debris present over the seaweed sample. Then washed seaweeds were shad dried at room temperature. The dried seaweeds were cut into small pieces and make into powdered in a mixer grinder for further extraction process.

#### Extraction of Fucoidan

The extraction of the water soluble crude Sulfated polysaccharide (WSCSPs) was performed as described by with minor modifications [7]. 100 grams of *Sargassum polycystum* sample was treated with a liter of ethanol and stirred with a mechanical stirrer for about 12 h at room temperature in order to remove proteins and pigments, centrifuged at 3000 rpm for 10 min. Then the residue was left to dry at room temperature. Five grams of the biomass was taken and extracted with 100 ml of acidic water pH 2.0 at 90°C with stirring for a 3 h. The extraction was done twice and the extracts were pooled. The combined extracts were centrifuged at 15,000 rpm for 10 min and the supernatant was collected. Then the supernatant was mixed well with 1% of CaCl<sub>2</sub> and the solution was kept at 4°C overnight to precipitate alginic acid. The solution was then centrifuged at 15,000 rpm for 10 min and the supernatant was collected. Ethanol (99%) was added into the supernatant in order to arrive upon the final ethanol concentration of 30% and the solution was positioned at 4°C for 4h. The obtained solution was centrifuged at 15,000 rpm for 10 min and the pellet was collected. Ethanol (99%) was added into the supernatant in order to reach the final ethanol concentration of 70% and the solution was kept at 4°C overnight.

## Qualitative analysis of the water soluble crude sulfated polysaccharide of *Sargassum polycystum* [8-10]

**Phenols**2 ml of plant extract was taken in a test tube and add 1% lead acetate solution. Formation of white precipitate indicates the presence of phenolic compounds

## Tannins

Take 2 ml of plant extract and add few drops of 0.1% ferric chloride solution in it, formation of brownish green color indicates the presence of tannins

#### Flavanoids

2ml of plant extract was treated with 2ml of 10% Lead acetate solution. Appearance of yellowish green color indicated the presence of flavanoid

## Saponins

To about 1ml of each extract was added to 2ml of distilled water in a test tube and shaken vigorously with few drops of olive oil. Foam which persisted was taken as an evidence for the presence of saponins

## Terpenoids

2ml of each extract of plant samples was mixed with 2ml of chloroform. Then allow evaporating and adding 2ml of concentrated sulfuric acid, then heat for 2 minutes. Greyish color indicates the presence of terpenoids

#### Alkaloids

Take 2 ml of plant extract and add 2ml Wagner's reagent. Test tubes were observed for the appearance of reddish brown precipitate

#### Glycosides

To 100  $\mu l$  of plant extract 2ml of concentrated H2SO4 was added. Formation of reddish brown color indicates the presence of glycosides

#### Quinones

2ml of plant extract was mixed with 3 or 4 drops of concentrated HCl. Formation of yellow color indicated the presence of quinones

## Fatty Acids

0.5 ml of extract was added to 5ml of ether and allowed it to evaporate on filter paper. Then the filter paper was dried and the appearance of transparency on filter paper is the indication of presence of fatty acids

## Phytosterols

To 2ml of plant extract, 2ml of chloroform and 2ml of concentrated H2SO4 was added and shaken well. If the chloroform layer appeared red and acid layer appeared greenish yellow fluorescent indicates the presence of sterols

# Antioxidant activity of water soluble crude sulfated polysaccharide of *Sargassum polycystum* DPPH Radical Scavenging Assay

The WSCSPs of *S. Polycystm* was tested against 2,2-diphenyl-1-picrylhydrazyl (DPPH) at varying concentrations such as 50, 100, 150, 200, and 250µg/ml and phenytoin various concentrations such as 6.25, 12.5, 25, 50, and 100 and 250µg/ml to check their donor capacity at dark conditions. Ascorbic acid was used as standard. The readings were taken in UV-Spectrometer at 517nm. The experiments were performed in triplicates [11].

## Ferric Reducing Antioxidant Power Assay

The WSCSPs of *S. Polycystm* and phenytoin was tested. The FRAP assay was performed according to the references with minor modifications (Nishaa et al., 2012). Various concentration WSCSPs and Phenytoin such as 50, 100, 150, 200, and 250 $\mu$ g/ml and phenytoin various concentrations such as 6.25, 12.5, 25, 50, and 100 250  $\mu$ g/ml mixed with 2mL of the FRAP reagent and added with distilled water up to 1 mL. After 30 minutes of incubation. Absorbance was measured at 593 nm against blank [12].

## ABTS Assay

ABTS [2, 2'-azinobis-(3-ethylbenzothiazoline-6-sulfonic acid)] Radical Cation Scavenging Assay. Free radical scavenging activity of the extract was also determined by ABTS (Sigma-Aldrich) radical cation decolorization assay [13]. ABTS radical cation was generated by mixing 20 mM ABTS solution with 70 mM potassium peroxodisulphate and allowing it to stand in dark at room temperature for 24 hours before use. 0.6 ml of extract (0.25 mg) was mixed with 0.45 ml of ABTS reagent and absorbance of these solutions was measured at 734 nm after 10 min.

#### **Statistical Analysis**

Statistical analysis of was done by one-way ANOVA followed by Student's t test. P<0.05 was considered as significant.

## **RESULTS AND DISCUSSION**

The present study exemplified the antioxidant potential of water soluble crude sulfated polysaccharide of Sargassum polycystum. Seaweeds are known as medicinal plants, rich in metabolites and have been extensively studied and used in the pharmaceutical industry. Phytochemical analysis of water soluble crude sulfated polysaccharide of *Sargassum polycystum* exhibits various phytoconstituents viz., phenols, tannins, flavonoids, saponins, terpenoids, alkaloids and phytosterols (Table 1). Plants, especially seaweeds, contain phenolic compounds, which have been shown to have a wide range of biological activities, including antioxidant properties [14 -15]. According to the reports, phenolic compounds are one of the most powerful antioxidants found in brown algae. Our results were akin with Nazarudin et al. [16] reported that the *S. polycystum* exhibits various secondary metabolites Tannins, which may bind to adhesives and play roles in enzyme inhibition, substrate deprivation, and membrane disruption, have been discovered to have antibacterial effects [17]. Saponins have specific biological activities such as anticancer, anti inflammatory, antimicrobial and antioxidant properties [18]. Flavonoids are phenolic compounds that have been hydroxylated and have been studied for their antioxidant properties [19]. The ability of flavonoids to scavenge hydroxyl radicals, superoxide anion radicals, and lipids is thought to be the reason for their action. Peroxy radicals are essential for preventing illnesses caused by oxidative cell, membrane, and protein damage. Yadav and Agarwala [20] reported that the steroids have been possess antibacterial properties and they are very important compounds due to their relationship with compounds like sex hormones. Terpenoids have been reported to possess cytotoxicity against a variety of cancer cells and cancer prophylactic [20]. Hence the presence of secondary metabolites from S. *polycystum* suggests that, it can be used as various biological properties that have great medicinal values and extensively used in the drug and pharmaceutical industry.

S. No	Phytochemical test	WSCSP
1	Phenols	Present
2	Tannins	Present
3	Flavonoids	Present
4	Saponins	Present
5	Terpenoids	Present
6	Alkaloids	Present
7	Glycosides	Absent
8	Quinones	Absent
9	Fatty acids	Absent
10	Phytosterols	Present

Table 1 Qualitative analysis of the water soluble crude sulfated polysaccharide of
Sargassum polycystum

Table 2 DPPH Radical Scavenging Assay of water soluble crude sulfated polysaccharide of
Sargassum polycystum

S. No	Concentrations µg/ml	WSCSPs
1	50	32.10 ± 0.09
2	100	$43.16 \pm 0.04$
3	150	$51.08 \pm 0.06$
4	200	76.13 ± 0.06
5	250	92.10 ± 0.09

The results of the antioxidant characteristics of water soluble crude sulfated polysaccharide of Sargassum polycystum, estimated by the DPPH scavenging activity, Ferric reducing antioxidant power assay and ABTS assay. The free radical scavenging antioxidant assay 2,2-diphenyl-1-picrylhydrazyl (DPPH) has been widely used to assess the antioxidant capacity of extracts. Sachindra et al. [21] opined that the radical molecule is stable and does not need to be manufactured, the DPPH test is regarded a valid, accurate, simple, and cost-effective approach for evaluating antioxidant radical scavenging activity. A novel methodology for assessing antiradical efficiency against DPPH has been developed, which has several advantages over existing methods. In the present study, water soluble crude sulfated polysaccharide of Sargassum polycystum depicted higher tendency to scavenge the DPPH radicals in a dose dependent manner and steadily increased with increase in extract concentrations. At 50 µg/mL of water soluble crude sulfated polysaccharide of Sargassum polycystum possessed 32.10 % and the maximum level of DPPH reduction was observed at 250  $\mu$ g/mL and it was found to be 92.10% which suggested that the polysaccharide of Sargassum polycystum possess more potential to scavenge DPPH free radicals due to the higher concentration of phenolic content (Table 2 and Fig.1). Previous studies reported that ethanolic and aqueous extracts of Sargassum spp. including S. horneri, S. macrocarpum and *S. siliquastrum* produced more than 60% of DPPH radical scavenging activity [22-23] Seaweed containing antioxidant molecules can diminish DPPH free radicals by attacking them and adding hydrogen atoms or electrons to them, changing the colour of the DPPH solution from purple to yellow [24]. In compared to red and green seaweeds, brown seaweed rich in natural bioactive components such as carotenoid, fucoxanthin, phenolic, and flavonoids has been shown to have better antioxidant capacity [25].

Antioxidants in the sample reduce the ferric (III) to ferrous (II) in a redox-linked colourimetric reaction involving single electron transfer in the FRAP assay. The reducing power of antioxidant chemicals shows that they are electron donors that can reduce the oxidised intermediate of the lipid peroxidation process, allowing them to function as primary and secondary antioxidants [26]. The reducing capacity of the samples were analyzed by the FRAP method measuring the absorbance at 593 nm and antioxidant power was calculated. The highest FRAP values was found to be 89.12 % at 250  $\mu$ g/mL of polysaccharide of *Sargassum polycystum*. Similarly, Giriwono et al. [27] reported that the lipid-soluble fraction of *Sargassum cristaefolium* possessed free radical scavenging activity. The antioxidant compounds reducing power suggests that they are electron donors and can reduce the oxidised intermediates of the lipid per oxidation process, allowing them to function as primary and secondary antioxidants. Using the FRAP assay, we found that the brown seaweed *Sargassum polycystum* exhibits better FRAP activity.

The ABTS assay is a simple indirect approach for evaluating natural antioxidant activity. ABTS radical is rather stable in the absence of phenolics, but it interacts energetically with an H-atom donor such as phenolics, resulting in a non-colored type of ABTS [28]. The antioxidant activity rises with increasing

concentrations, according to the ABTS radical cation-scavenging assay. The maximum scavenging effect was shown by the polysaccharide of Sargassum polycystum at 250 µg/ml and it was found to be 91.06 % and it was statistically significant (p<0.05) (Table 4 and Fig.3). The lower concentration of extract was sufficient to nullify free radical mediated chain reactions in this assay. Parthiban et al. [29] reported that the strongest anti-oxidant activity in the ABTS method was achieved by the compound phenols. The results of the present study correlated with the result of Sroka and Cisowski [30] that they were confirmed the strong quench ABTS radical is related with the more phenol contents of the experimental alga. The capacity of seaweed extracts to scavenge free radicals may account for their antioxidant action. Seaweeds, with their multiple polyphenols and polysaccharides enriched in a hydroxyl group and carbonyl group on ring C, have several sites for metal complexation able to chelate metal ions. Furthermore, the antioxidant activity of seaweed extracts appears to be mediated by phenolic substances. Sargassum polycystum can be employed for a variety of beneficial chemo-preventive effects, according to the findings. As there is a growing trend of disease and an increased requirement for medicine or drugs, this study recommends that macroalgae being an underutilized bioresource could be exploited in a sustainable manner for the welfare of mankind. The findings of this study support the fact that some seaweed commonly consumed in India are promising sources of potential antioxidants.

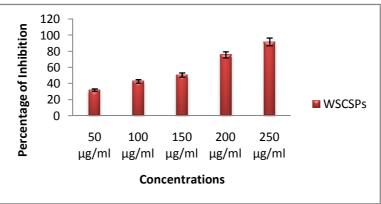
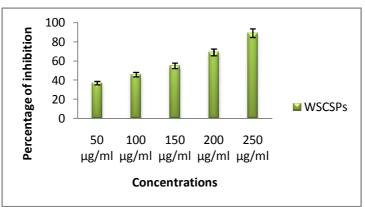
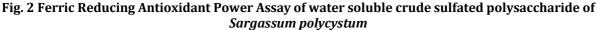


Fig.1 DPPH Radical Scavenging Assay of water soluble crude sulfated polysaccharide of Sargassum polycystum

۲able 3 Ferric Reducing Antioxidant Power Assay of water soluble crude sulfated polysaccharid		
of Sargassum polycystum		

S. No	Concentrations µg/ml	WSCSPs
1	50	$37.03 \pm 0.12$
2	100	$46.08 \pm 0.1$
3	150	55.21 ± 0.21
4	200	$69.08 \pm 0.28$
5	250	89.12 ± 0.32





S. No	Concentrations µg/ml	WSCSPs
1	50	33.11 ± 0.11
2	100	44.05 ± 0.17
3	150	58.98 ± 0.26
4	200	$74.19 \pm 0.32$
5	250	91.06 ± 0.40

Table 4 ABTS Assay of water soluble crude sulfated polysaccharide of Sargassum polycystum

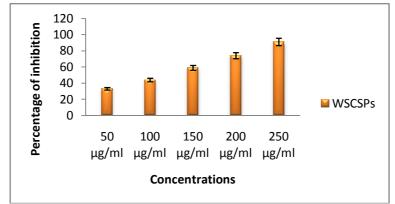


Fig.3 ABTS Assay of water soluble crude sulfated polysaccharide of Sargassum polycystum

#### **CONFLICT OF INTEREST**

The authors declare that they have no conflict of interest.

#### REFERENCES

- 1. Saeed, N., Khan, M. R., & Shabbir, M. (2012). Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts Torilis leptophylla L. BMC Complementary and alternative medicine, 12(1), 1-12.
- 2. Tziveleka, L. A., Tammam, M. A., Tzakou, O., Roussis, V., & Ioannou, E. (2021). Metabolites with Antioxidant Activity from Marine Macroalgae. Antioxidants, 10(9), 1431.
- 3. Rekha, C., Poornima, G., Manasa, M., Abhipsa, V., Devi, J. P., Kumar, H. T. V., & Kekuda, T. R. P. (2012). Ascorbic acid, total phenol content and antioxidant activity of fresh juices of four ripe and unripe citrus fruits. Chemical Science Transactions, 1(2), 303-310.
- 4. Kuda, T., Nemoto, M., Kawahara, M., Oshio, S., Takahashi, H., & Kimura, B. (2015). Induction of the superoxide anion radical scavenging capacity of dried 'funori'Gloiopeltis furcata by Lactobacillus plantarum S-SU1 fermentation. Food & function, 6(8), 2535-2541.
- 5. Ismail, G. A. (2017). Biochemical composition of some Egyptian seaweeds with potent nutritive and antioxidant properties. Food Science and Technology, 37, 294-302.
- 6. Arsianti, A., Bahtiar, A., Wangsaputra, V. K., Azizah, N. N., Fachri, W., Nadapdap, L. D., ... & Kakiuchi, K. (2020). Phytochemical Composition and Evaluation of Marine Algal Sargassum polycystum for Antioxidant Activity and In Vitro Cytotoxicity on Hela Cells. Pharmacognosy Journal, 12(1).
- Tabarsa, M., You, S., Dabaghian, E. H., & Surayot, U. (2018). Water-soluble polysaccharides from Ulva intestinalis: Molecular properties, structural elucidation and immunomodulatory activities. Journal of food and drug analysis, 26(2), 599-608.
- 8. Kancherla, N., Dhakshinamoothi, A., Chitra, K., & Komaram, R. B. (2019). Preliminary Analysis of Phytoconstituents and Evaluation of Anthelminthic Property of Cayratia auriculata (In Vitro). Maedica, 14(4), 350.
- 9. Saxena, M., Saxena, J., Nema, R., Singh, D., & Gupta, A. (2013). Phytochemistry of medicinal plants. Journal of pharmacognosy and phytochemistry, 1(6).168-182
- 10. Harborne, A. J. (1998). Phytochemical methods a guide to modern techniques of plant analysis. springer science & business media. ISBN: 978-94-009-5570-7
- 11. Sharma, O. P., & Bhat, T. K. (2009). DPPH antioxidant assay revisited. Food chemistry, 113(4), 1202-1205.
- 12. Guo, C., Yang, J., Wei, J., Li, Y., Xu, J., & Jiang, Y. (2003). Antioxidant activities of peel, pulp and seed fractions of common fruits as determined by FRAP assay. Nutrition research, 23(12), 1719-1726.
- 13. Re, R., Pellegrini, N., Proteggente, A., Pannala, A., Yang, M., & Rice-Evans, C. (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free radical biology and medicine, 26(9-10), 1231-1237.
- 14. Athukorala, Y., Kim, K. N., & Jeon, Y. J. (2006). Antiproliferative and antioxidant properties of an enzymatic hydrolysate from brown alga, Ecklonia cava. *Food and chemical toxicology*, *44*(7), 1065-1074.

- 15. Kalasariya, H. S., Yadav, V. K., Yadav, K. K., Tirth, V., Algahtani, A., Islam, S., ... & Jeon, B. H. (2021). Seaweed-Based Molecules and Their Potential Biological Activities: An Eco-Sustainable Cosmetics. *Molecules*, *26*(17), 5313.
- Nazarudin, M. F., Paramisparam, A., Khalid, N. A., Albaz, M. N., Shahidan, M. S., Yasin, I. S. M., ... & Aliyu-Paiko, M. (2020). Metabolic variations in seaweed, Sargassum polycystum samples subjected to different drying methods Via 1H NMR-based metabolomics and their bioactivity in diverse solvent extracts. Arabian Journal of Chemistry, 13(11), 7652-7664.
- 17. Tiwari, P., Kumar, B., Kaur, M., Kaur, G., & Kaur, H. (2011). Phytochemical screening and extraction: a review. *Internationale pharmaceutica sciencia*, 1(1), 98-106.
- 18. Sidana, J., Singh, B., & Sharma, O. P. (2016). Saponins of Agave: Chemistry and bioactivity. Phytochemistry, 130, 22-46.
- 19. Sytar, O., Hemmerich, I., Zivcak, M., Rauh, C., & Brestic, M. (2018). Comparative analysis of bioactive phenolic compounds composition from 26 medicinal plants. Saudi journal of biological sciences, 25(4), 631-641.
- 20. Yadav, R. N. S., & Agarwala, M. (2011). Phytochemical analysis of some medicinal plants. Journal of phytology, 3(12).
- 21. Sachindra, N. M., Airanthi, M. K. W. A., Hosokawa, M., & Miyashita, K. (2010). Radical scavenging and singlet oxygen quenching activity of extracts from Indian seaweeds. Journal of food science and technology, 47(1), 94-99.
- 22. Boonchum, W., Peerapornpisal, Y., Kanjanapothi, D., Pekkoh, J., Pumas, C., Jamjai, U., ... & Vacharapiyasophon, P. (2011). Antioxidant activity of some seaweed from the Gulf of Thailand. International Journal of Agriculture and Biology, 13(1).
- 23. Matsukawa, R., Dubinsky, Z., Kishimoto, E., Masaki, K., Masuda, Y., Takeuchi, T., ... & Karube, I. (1997). A comparison of screening methods for antioxidant activity in seaweeds. Journal of Applied Phycology, 9(1), 29-35.
- 24. Indu, H., & Seenivasan, R. (2013). In vitro antioxidant activity of selected seaweeds from southeast coast of India. International Journal of Pharmacy and Pharmaceutical Sciences, 5(2), 474-484.
- 25. Leelavathi, M. S., & Prasad, M. P. (2014). Evaluation of antioxidant properties of marine seaweed samples by DPPH method. Int. J. Pure App. Biosci, 2(6), 132-137.
- 26. Meenakshi, S., Umayaparvathi, S., Arumugam, M., & Balasubramanian, T. (2011). In vitro antioxidant properties and FTIR analysis of two seaweeds of Gulf of Mannar. Asian Pacific Journal of Tropical Biomedicine, 1(1), S66-S70.
- 27. Giriwono, P. E., Iskandriati, D., Tan, C. P., & Andarwulan, N. (2020). In-vitro anti-inflammatory activity, free radical (DPPH) scavenging, and ferric reducing ability (FRAP) of Sargassum cristaefolium lipid-soluble fraction and putative identification of bioactive compounds using UHPLC-ESI-ORBITRAP-MS/MS. Food Research International, 137, 109702.
- 28. Roginsky, V., & Lissi, E. A. (2005). Review of methods to determine chain-breaking antioxidant activity in food. Food chemistry, 92(2), 235-254.
- 29. Parthiban, C., Saranya, C., Girija, K., Hemalatha, A., Suresh, M., & Anantharaman, P. (2013). Evaluation of in vitro antioxidant properties of some selected seaweeds from Tuticorin coast. Int J Curr Microbiol App Sci, 2(9), 64-73.
- 30. Sroka, Z., & Cisowski, W. (2003). Hydrogen peroxide scavenging, antioxidant and anti-radical activity of some phenolic acids. Food and Chemical Toxicology, 41(6), 753-758.

#### CITATION OF THIS ARTICLE

P Digala, M Dhanraj, Vi Savariar. Antioxidant Potential of water soluble crude sulfated polysaccharide of *Sargassum polycystum*. Bull. Env.Pharmacol. Life Sci., Spl Issue [1] 2022 : 339-345