



The Efficacy of Nutritional Supplementation in The Therapy of Metabolic Dysfunctions Associated with Polycystic Ovarian Syndrome: A Critical Review

^{*,1}Shahid khan, ² Akhand Pratap Singh, ³Mohd Ashraf Gaine

¹Research Scholar, Maharishi University of information technology, Lucknow, India

²Professor, Maharishi University of information technology, Lucknow, India

³Professor, SKIMS, Jammu&Kashmir, India

*Corresponding author

ABSTRACT

PCOS is a complicated heterogeneous condition with multiple pathological manifestations, including metabolic, endocrine, reproductive, and psychological. However, the aetiology of PCOS is yet unknown. Numerous studies indicate that insulin resistance and hyperandrogenism are critical factors in the pathogenesis of PCOS. As a result, the majority of PCOS treatment techniques focus on lifestyle improvement, including exercise, nutrition, and nutrient supplementation therapy. Recent studies have indicated some nutrients for the treatment of PCOS, including vitamins, minerals, and vitamin-like substances, because each has at least one functional feature in PCOS-induced pathways. As a result, vitamin or mineral deficiency is suggested as a possible cause of PCOS. The purpose of this review is to conduct a comprehensive literature search on nutritional supplements for the treatment of PCOS-related endocrine and metabolic dysfunctions and to examine the role of nutrients in PCOS management in light of clinical trials and experimental investigations. (*Turkish Journal of Gynecology and Obstetrics* 2018; 19: 220-32).

Keywords: PCOS, insulin sensitivity, hyperandrogenism, metabolism disorder, supplements

Received 11.02.2022

Revised 08.03.2022

Accepted 28.03.2022

INTRODUCTION

PCOS is one of the most prevalent endocrine disorders, affecting between 5% and 10% of women of adolescent and reproductive age[1,2]. Stein and Leventhal characterised PCOS for the first time in 1935. PCOS is defined by hyperandrogenism, which occurs as a result of increased androgen secretion or activity[3]. However, hyperandrogenism alone is not sufficient to diagnose PCOS. PCOS is described by the Rotterdam criteria as the presence of at least two of the following three criteria: hyperandrogenism, chronic anovulation, and polycystic ovaries on ultrasound findings[4]. Later, the Androgen Excess and PCOS Society reported that hyperandrogenism and ovarian dysfunction (anovulation and polycystic ovaries) to be present in order to diagnose PCOS[5]. PCOS patients present with a variety of symptoms, including menstrual dysfunction, hyperinsulinemia, infertility, glucose intolerance, type 2 diabetes, hirsutism, obesity, acne, metabolic syndrome, an increased risk of developing cardiovascular disease, endometrial cancer, anxiety, obstructive sleep apnea, and abnormal lipid profiles[6,7]. Despite substantial research, the origin of PCOS remains unknown because to the poorly understood interplay of genetic and environmental factors[8]. PCOS can be caused by a variety of neuroendocrine abnormalities, including decreased ovarian steroidogenesis, insulin resistance (IR), and elevated cortisol metabolism-related adrenal hyperandrogenism[9-11]. Recent research indicates that IR may lead to metabolic and reproductive problems. As a result, IR is critical in the pathophysiology of PCOS[12].

In a nutshell, insulin is considered a critical hormone for hyperandrogenism in the pathogenesis of PCOS via two distinct pathways:

(1) Insulin stimulates theca cells to produce androgen via luteinizing hormone (LH), and increased androgen production results in hirsutism, acne, and anovulatory infertility.

(2) Insulin's hyperandrogenism-associated function is to limit the synthesis of sex hormone-binding globulin (SHBG) in the liver [13].

SHBG is a plasma protein that binds androgens and estrogens, and so low SHBG levels in PCOS can result in hyperandrogenism. Insulin is critical in controlling glucose metabolism, inhibiting lipolysis, and

activating amino acid transport[14]. Numerous nutrients are involved in the regulation of the insulin signalling pathway and androgen production.

Sufficient nutrients and energy for growth and reproduction are contingent upon the ideal nutritional composition being defined. It is well established that nutrition-related signaling pathways are critical for the regulation of ovarian follicle growth and ovulation rates[15]. Deficiencies in myo-inositol and vitamin D, in particular, can result in difficulties associated with PCOS pathogenesis [16-18]. Thus, nutritional supplementation may aid in the resolution of PCOS issues such as immature oocytes, infertility, hyperandrogenism, and oxidative stress. The current state of knowledge regarding the efficacy of nutrients in the treatment of PCOS is discussed in light of experimental and clinical studies.

SUPPLEMENTS WITH VITAMINS

Vitamin A, commonly known as retinol, is a fat-soluble vitamin. Vitamin A metabolites such as retinoids, retinoic acid, and retinol contribute to antioxidant activity, steroid metabolism, nuclear maturation of oocytes, and prevention of cumulus cell apoptosis[19,20]. It is well established that genes involved in retinoic acid production are differently expressed in theca internal cells isolated from patients with PCOS[21]. To investigate the effects of retinol and retinoids, retinol derivatives were added to theca internal cell culture derived from PCOS and healthy women. Trans retinol treatment of all theca internal cells enhanced dehydroepiandrosterone levels and mRNA accumulation of the enzyme cytochrome P450 17 hydroxylase (CYP17), which is implicated in androgen production and retinol biosynthesis[22]. In overweight women with PCOS, obesity and impaired glucose metabolism are related with higher retinol-binding protein 4 (RBP4) levels [23]. Another study examining RBP4 expression in isolated subcutaneous and omental adipose tissue from women with PCOS was published. The authors then proposed that higher 17 estradiol could contribute to the altered gonadal and adrenal steroid profiles by upregulating the RBP4 gene[24].

B group vitamins; the majority of research on this group focuses on B6, B12, and folic acid due to the growing significance of homocysteine (Hcy) in PCOS. Hcy is an important amino acid produced from dietary methionine in this pathway, and increasing total plasma Hcy levels enhance the risk of cardiovascular and reproductive symptoms in PCOS[25]. Additionally, other metabolic pathways required for cell and tissue growth are intimately linked to Hcy[26]. Folic acid, vitamin B6, and vitamin B12 all play a critical role in regulating Hcy. A favourable association between IR and high Hcy levels has been observed in the physiopathology of PCOS[27,28]. Kaya et al. [29] revealed that in women with PCOS, IR, obesity, and elevated Hcy levels were all associated with low serum insulin B12 concentrations. Folic acid supplementation for three months was helpful in lowering serum Hcy levels, particularly in women without IR. However, it is unknown if folic acid supplementation has a dose-dependent effect[30]. Regular exercise has also been associated with a reduction in plasma Hcy levels in the pathophysiology of PCOS. According to Randeve et al. [31], six months of regular exercise results in considerably decreased plasma Hcy levels in young obese and overweight women with PCOS.

Numerous women with PCOS are need to utilise insulin-sensitizing medications such as metformin to improve insulin sensitivity. Metformin inhibits the intrinsic factor-B complex and its receptor, and also causes a drop in serum vitamin B12 and folic acid levels during metformin therapy[32]. Additionally, metformin elevates Hcy levels, increasing the risk of cardiovascular disease in women with PCOS over the long term [33]. Two studies demonstrated an interaction between metformin and B group vitamins: the first demonstrated that daily treatment of folic acid or B group vitamins may be useful in lowering increased Hcy levels in women with PCOS receiving short-term metformin therapy. The investigators did, however, suggest that vitamin supplementation had no influence on androgen and lipid levels in the pathophysiology of PCOS [34]. The second investigation demonstrated that six months of metformin combined with folate supplementation had favourable effects on the vascular endothelium. Because this medication results in decreased Hcy levels, it may be useful in managing long-term consequences of PCOS, such as cardiovascular disease [35].

Inositol and its metabolites are classified as sugar alcohols and are also vitamins of the B complex. Additionally, inositol exists in nine stereoisomers: myo-, cis-, allo-, epi-, muco-, neo-, scyllo-, D-chiro, and L-chiro. Insulin-sensitive metabolites generated from inositol play critical roles in lipid synthesis, signal transduction, oocyte maturation, oogenesis, cell morphogenesis, and cytoskeleton organisation [37]. According to randomised controlled trials including inositol supplementation in women with PCOS, inositol improves nearly all pathologic symptoms associated with the disorder, including the recovery of reproductive anomalies, lowered androgen levels, and improved insulin levels[38].

Interestingly, a combination of inositol isomers such as myo-inositol (MI) and D-chiro inositol (DCI) should be used in a specific ratio, referred to as the plasma physiologic ratio (MI/DCI: 40/1)[39]. Otherwise, immature oocytes may emerge, and inositol's efficacy may be compromised in the

pathophysiology of PCOS[40]. According to several investigations, these pathologic symptoms may be explained by the 'DCI paradox' (41). MI is involved in the follicle-stimulating hormone (FSH) signalling system and glucose uptake homeostasis, whereas DCI stimulates insulin-associated androgen production. Except when the physiologic ratio is exceeded, epimerase performs a functional role in the conversion of MI to CDI, dependent on insulin levels and also on inositol isomerase intake. When hyperinsulinemia is present during the pathogenesis of PCOS, increased epimerase activity might result in anomalies in the FSH signalling pathway, resulting in the development of immature oocytes and hyperandrogenism[42]. Table 1 summarises the contribution of MI to treatment in women with PCOS[43-50]. According to the available literature, therapy of MI improves hyperandrogenism and IR-related indicators, as well as the lipid profile.

Vitamin D is critical for skeletal growth, serotonin synthesis regulation, bone mineral density, dental health, lower extremity function, and calcium (Ca) and phosphorus metabolism modulation. Additionally, prior research has indicated that vitamin D may be a substantial and independent predictor of IR[50]. Vitamin D levels drop in obese patients compared to non-obese individuals as a result of IR. In terms of PCOS, a recent review by Krul-Poel et al. [52] on the role of vitamin D in metabolic disturbances associated with the condition confirmed a link between vitamin D and metabolic disturbances. Thus, it was discovered that obese women with PCOS had considerably lower 25-dehydroxy vitamin D levels[53]. Additionally, a cross-sectional study indicated that low D vitamin levels were associated with IR as a result of PCOS's pathophysiology[54].

References	Patients	Treatment	Outcomes
Nestler et al. (43)	44 obese women with PCOS (placebo group n=22, inositol group n=22)	Oral administration of 1200 mg of D-chiro inositol per day for 7-8 weeks	<ul style="list-style-type: none"> Plasma triglyceride ↓ Diastolic and systolic pressure ↓ DHEA-S ↓ SHBG ↑
Baillargeon et al. (44)	19 obese women with PCOS	For 4-8 weeks, oral administration of metformin therapy (n=10) (500 mg orally thrice daily) and placebo group (n=9)	<ul style="list-style-type: none"> Improvement of insulin mediated release of DCI-IPG
Gerfi et al. (45)	92 women with oligomenorrhea and polycystic ovaries	For 12-16 weeks, 400 mcg folic acid intake in placebo group (n=47) and 400 mcg folic acid + 4 g inositol intake in treatment group (n=45)	<ul style="list-style-type: none"> Higher ovulation rate Weight loss Follicular maturation Circulating HDL ↑
Papaleo et al. (46)	25 women with PCOS who have oligo or amenorrhea since childbearing age	Myo-inositol + folic acid (inofolic) (2 g twice a day) for 6 months	<ul style="list-style-type: none"> Improvement in menstrual cyclicity Replacement of healthy ovarian activity Serum free testosterone ↓
Genazzani et al. (47)	20 overweight women with PCOS	Group A (n=10); 2 g myo-inositol + 200 µg folic acid per day Group B (n=10); 200 µg folic acid per day for 12 weeks	<ul style="list-style-type: none"> Circulating LH, T, PRL and insulin level ↓ Ratio of LH/FSH ↓ Restoration of menstrual cyclicity
Costantino et al. (48)	42 women with PCOS from reproductive age (18-40 years)	Placebo group (n=19): 400 mcg folic acid alone; experiment group (n=23): 4 g myo-inositol + 400 mcg folic acid for 12-16 weeks	<ul style="list-style-type: none"> Insulin and androgen level ↓ Improved glucose tolerance
Minozzi et al. (49)	155 women with PCOS	12 weeks' treatment: placebo group (n=75) oral contraceptive pills (OCP) intake, and the treatment group OCP + myo-inositol (4 g/day) intake	<ul style="list-style-type: none"> Insulin sensitivity ↑ Recovery of hirsutism Androgen synthesis ↓ HDL cholesterol level ↑ LDL cholesterol level ↓
Morgante et al. (50)	Insulin resistant women with PCOS (n=15)	Low dose step-down gonadotropin regimen + Redestop (1500 mg inositol, 100 mg lactoferrin)	<ul style="list-style-type: none"> Improved clinical outcomes Pregnancy rate ↑ Number of follicles >15 mm in diameter ↓ Cancellation rate ↓

DHEA-S: Dehydroepiandrosterone sulfate; PCOS: Polycystic ovary syndrome; SHBG: Sex hormone-binding globulin; HDL: High-density lipoprotein; LDL: Low-density lipoprotein; T: Testosterone; LH: Luteinizing hormone; PRL: Prolactin; DCI-IPG: D-chiro-inositol-containing-inositolphosphoglycan; FSH: Follicle-stimulating hormone; ↓ decreasing; ↑ increasing

Table 1 The effects of myo-inositol compounds in women with polycystic ovary syndrome

The researchers concentrate on vitamin D supplementation as a means of treating women with PCOS in order to demonstrate a link between vitamin D insufficiency and PCOS. A recent investigation on vitamin D replacement therapy in 11 people with PCOS found some favourable benefits on IR but no changes in androgen levels[55]. Additionally, Kotsa et al.[56] examined the effect of vitamin D on PCOS using a vitamin D3 analogue (alphacalcidol). Their findings indicated an increase in insulin secretion during the first phase, a drop in serum triglyceride (TG) levels, and an increase in serum high-density lipoprotein (HDL) cholesterol levels.

The biochemical mechanism through which vitamin D supplementation improves PCOS is unknown at the moment. However, a recent investigation reported that supplementing with vitamin D3 improved several biochemical indicators in women with PCOS by increasing the amount of soluble receptor for Advanced

Glycosylated Ends (AGEs). As a result, vitamin D3 reduces the progression of inflammation in the pathogenesis of PCOS. Additionally, vitamin D3 therapy promotes folliculogenesis by lowering increased anti-mullerian hormone levels[57]. Interestingly, Jafari-Sfidvajani et al.[58] demonstrated that when vitamin D supplementation was combined with a low-calorie diet, no statistically significant changes in the androgen profile occurred; however, menstrual frequency improved.

Vitamin E is a lipid-soluble antioxidant and free radical scavenger that helps maintain a healthy balance of antioxidant and oxidant systems[59]. Additionally, fresh studies indicated that vitamin E can increase endometrial thickness in women with unexplained infertility, with the effects linked to the vitamin's anticoagulant and antioxidant properties[60]. Additionally, coenzyme q10 and vitamin E cotreatment for eight weeks improved SHBG values in patients with PCOS[61]. Another study found that co-supplementing vitamin E (400 IU) and omega-3 fatty acids (1000 mg) for 12 weeks significantly improved IR and androgen levels in women with PCOS[62]. Vitamin E is a lipid-soluble antioxidant and free radical scavenger that helps maintain a healthy balance of antioxidant and oxidant systems[59]. Additionally, fresh studies indicated that vitamin E can increase endometrial thickness in women with unexplained infertility, with the effects linked to the vitamin's anticoagulant and antioxidant properties[60]. Additionally, coenzyme q10 and vitamin E cotreatment for eight weeks improved SHBG values in patients with PCOS[61]. Another study found that co-supplementing vitamin E (400 IU) and omega-3 fatty acids (1000 mg) for 12 weeks significantly improved IR and androgen levels in women with PCOS[62].

VITAMIN-LIKE NUTRIENT SUPPLEMENTATION IN PCOS:

Alpha-Lipoic Acid (α-LA) is a powerful antioxidant, an essential cofactor in the citric acid cycle, and a weight-regulating agent (63,64). Interestingly, Masharani et al. [65] discovered that regulated release of -LA was not associated with an increase in plasma antioxidant capacity or a decrease in plasma oxidation metabolites in six nondiabetic women with PCOS. To examine the effect of -LA and DCI (DCA) in the short-term therapy of PCOS, both metabolites were administered for 180 days to 46 women (26 with PCOS and 20 female controls). They suggested that certain reproductive features such as menstrual periods, ovarian cysts, and progesterone levels improved. From a metabolic standpoint, IR improved dramatically in participants with PCOS, while poor lipid metabolism was significantly improved[66].

OTHER SUPPLEMENTS:

Melatonin (MT) is a pineal gland-secreted neuroendocrine hormone. It is critical for circadian rhythm regulation. Due to its potent free radical scavenger activity, high amounts of MT have been detected in follicular fluid, affecting physiologic processes in the ovaries such as folliculogenesis, follicular atresia, ovulation, steroidogenesis in theca cells, and corpus luteum development [67-69]. Additionally, Wei et al. [70] found that low-dose MT supplementation promotes nuclear maturation of oocytes in vitro. As a result, MT may enhance oocyte quality and boost conception rates[71]. In women with PCOS, the concentration of MT in pre-ovulatory follicular fluid is decreased. Kim et al. [72] reported that MT treatment may be beneficial in enhancing the success of in vitro fertilisation and improving the clinical results of PCOS.

LIMITATIONS:

Our study was limited by the large number of related publications published: the doses, types, and combinations of supplemented nutrients are significantly varied, depending on the investigated group, making it difficult to evaluate the results. Another problem is the low nutritional doses employed in the trials, as well as the inadequacy of the PCOS diagnosis criteria. Additionally, each woman with PCOS requires a unique supplements regimen based on her signs and symptoms and physiologic anomalies. For example, some people experience infertility as a result of PCOS, while others suffer from endocrine and metabolic disorders. However, the majority of studies on nutrient supplementation is focused on the metabolic components of PCOS. As a result, this review focused mostly on the treatment effects on metabolic and endocrine dysfunctions rather than on infertility, which was also a constraint in this study. Large-scale molecular investigations might be organised to shed light on the altered signalling pathways associated with PCOS. By employing a molecular targeting technique, nutrients can be employed effectively to control all elements of PCOS.

CONCLUSION

In conclusion, vitamin and mineral supplements may help alleviate symptoms associated with PCOS, such as immature oocytes, hyperinsulinemia, hyperandrogenism, higher BMI, cardiovascular illnesses, and mental and psychological issues.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

REFERENCES

1. Knochenhauer ES, Key TJ, Kahsar-Miller M, Waggoner W, Boots LR, Azziz R. (1998). Prevalence of the polycystic ovary syndrome in unselected black and white women of the southeastern United States: a prospective study. *J Clin Endocrinol Metab* 83:3078-82.
2. Carreau AM, Baillargeon JP. (2015). PCOS in adolescence and type 2 diabetes. *Curr Diab Rep*; 15: 564.
3. Stein JD, Andrews C, Musch DC, Green C, Lee PP. Sight-Threatening Ocular Diseases Remain Under diagnosed Among Children Of Less Affluent Families. *Health Aff (Millwood)* 2016; 35: 1359-66.
4. Rotterdam ESHRE/ASRM-Sponsored PCOS consensus workshop group. (200). Revised 2003 consensus on diagnostic criteria and longterm health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod*; 19: 41-7.
5. Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, EscobarMorreale HF, Futterweit W, et al. (2014). The Androgen Excess and PCOS Society criteria for the polycystic ovary syndrome: the complete task force report. *Fertil Steril*; 91: 456-88.
6. Sheehan MT. (2013). Polycystic ovarian syndrome: diagnosis and management. *Clin Med Res*; 2: 13-27.
7. Sirmans SM, Pate KA. (2013). Epidemiology, diagnosis, and management of polycystic ovary syndrome. *Clin Epidemiol*; 6: 1-13.
8. Carmina E. (2013). Genetic and environmental aspect of polycystic ovary syndrome. *J Endocrinol Invest*; 26: 1151-9.
9. Doi SA. (2017). Neuroendocrine dysfunction in PCOS: a critique of recent reviews. *Clin Med Res* 2017; 6: 47-53.
10. Akayama K, Fukaya T, Sasano H, Funayama Y, Suzuki T, Takaya R, et al. (2006). Immunohistochemical study of steroidogenesis and cell proliferation in polycystic ovarian syndrome. *Hum Reprod*; 11: 1387-92.
11. Yaba A, Demir N. (2016). The mechanism of mTOR (mammalian target of rapamycin) in a mouse model of polycystic ovary syndrome (PCOS). *J Ovarian Res*; 5: 38.
12. Diamanti-Kandarakis E, Dunaif A. (2016). Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. *Endocr Rev*; 33: 981-1030.
13. Ehrmann DA. (2005). Polycystic ovary syndrome. *N Engl J Med*; 352: 1223-36.
14. Kahn CR. Banting Lecture. (1994). Insulin action, diabetogenesis, and the cause of type II diabetes. *Diabetes*; 43: 1066-84.
15. Yu J, Yaba A, Kasiman C, Thomson T, Johnson J. (2011). mTOR controls ovarian follicle growth by regulating granulosa cell proliferation. *PLoS One*; 6: 21415.
16. Muscogiuri G, Policola C, Prioletta A, Sorice G, Mezza T, Lassandro A, et al. (2012). Low levels of 25(OH)D and insulin-resistance: 2 unrelated features or a cause-effect in PCOS? *Clin Nutr*; 31: 476-80.
17. Thomson RL, Spedding S, Buckley JD. (2012). Vitamin D in the aetiology and management of polycystic ovary syndrome. *ClinEndocrinol (Oxf)*; 77: 343-50.
18. Lakimiuk AJ, Szamatowicz J. (2014). The role of inositol deficiency in the etiology of polycystic ovary syndrome disorders. *Ginekol Pol*; 85: 54-7.
19. Pu Y, Wang Z, Bian Y, Zhang F, Yang P, Li Y, et al. (2014). All-trans retinoic acid improves goat oocyte nuclear maturation and reduces apoptotic cumulus cells during in vitro maturation. *Anim Sci J* 2014; 85: 833-9.
20. Deb GK, Dey SR, Bang JI, Lee JG, Kong IK. (2012). 9-cis Retinoic acid inhibits cumulus cell apoptosis during the maturation of bovine cumulus-oocyte-complexes. *J Anim Sci*; 90: 1798-806.
21. Wood JR, Nelson VL, Ho C, Jansen E, Wang CY, Urbanek M, et al. (2013). The molecular phenotype of polycystic ovary syndrome (PCOS) theca cells and new candidate PCOS genes defined by microarray analysis. *J Biol Chem*; 278: 26380-90.
22. Wickenheisser JK, Nelson-DeGrave VL, Hendricks KL, Legro RS, Strauss JF, 3rd, McAllister JM. (2015). Retinoids and retinol differentially regulate steroid biosynthesis in ovarian theca cells isolated from normal cycling women and women with polycystic ovary syndrome. *J Clin Endocrinol Metab*; 90: 4858-65.
23. Hahn S, Backhaus M, Broecker-Preuss M, Tan S, Dietz T, Kimmig R, et al. (2007). Retinol-binding protein 4 levels are elevated in polycystic ovary syndrome women with obesity and impaired glucose metabolism. *Eur J Endocrinol*; 157: 201-7.
24. Tan BK, Chen J, Lehnert H, Kennedy R, Randeve HS. (2017). Raised serum, adipocyte, and adipose tissue retinol-binding protein 4 in overweight women with polycystic ovary syndrome: effects of gonadal and adrenal steroids. *J Clin Endocrinol Metab*; 92: 2764-72.
25. Yarali H, Yildirim A, Aybar F, Kabakci G, Bükülmez O, Akgül E, et al. (2011). Diastolic dysfunction and increased serum homocysteine concentrations may contribute to increased cardiovascular risk in patients with polycystic ovary syndrome. *Fertil Steril*; 76: 511-6.
26. De la Calle M, Usandizaga R, Sancha M, Magdaleno F, Herranz A, Cabrillo E. (2013). Homocysteine, folic acid and B-group vitamins in obstetrics and gynaecology. *Eur J Obstet Gynecol Reprod Biol*; 107: 125-34.
27. Loverro G, Lorusso F, Mei L, Depalo R, Cormio G, Selvaggi L. (2012). The plasma homocysteine levels are increased in polycystic ovary syndrome. *Gynecol Obstet Invest*; 53: 157-62.
28. Badawy A, State O, El Gawad S, El Aziz OA. (2017). Plasma homocysteine and polycystic ovary syndrome: the missed link. *Eur J Obstet Gynecol Reprod Biol*; 131: 68-72.

29. Kaya C, Cengiz SD, Satiroglu H. (2017). Obesity and insulin resistance associated with lower plasma vitamin B12 in PCOS. *Reprod Biomed Online* ; 19: 721-6.
30. Kazerooni T, Asadi N, Dehbashi S, Zolghadri J. (2008). Effect of folic acid in women with and without insulin resistance who have hyperhomocysteinemic polycystic ovary syndrome. *Int J Gynaecol Obstet* ; 101: 156-60.
31. Randeve HS, Lewandowski KC, Drzewoski J, Brooke-Wavell K, O'Callaghan C, Czupryniak L, et al. (2015). Exercise decreases plasma total homocysteine in overweight young women with polycystic ovary syndrome. *J Clin Endocrinol Metab*; 87: 4496-501.
32. De Jager J, Kooy A, Lehert P, Wulffele MG, van der Kolk J, Bets D, et al. (2011). Long term treatment with metformin in patients with type 2 diabetes and risk of vitamin B-12 deficiency: randomised placebo controlled trial. *BMJ* ; 340: 2181.
33. Kilicdag EB, Bagis T, Zeyneloglu HB, Tarim E, Aslan E, Haydardedeoglu B, et al. (2015). Homocysteine levels in women with polycystic ovary syndrome treated with metformin versus rosiglitazone: a randomized study. *Hum Reprod* ; 20: 894-9.
34. Kilicdag EB, Bagis T, Tarim E, Aslan E, Erkanli S, Simsek E, et al. (2015). Administration of B-group vitamins reduces circulating homocysteine in polycystic ovarian syndrome patients treated with metformin: a randomized trial. *Hum Reprod*; 20: 1521-8.
35. Palomba S, Falbo A, Giallauria F, Russo T, Tolino A, Zullo F, et al. (2018). Effects of metformin with or without supplementation with folate on homocysteine levels and vascular endothelium of women with polycystic ovary syndrome. *Diabetes Care* ; 33: 246-51.
36. Daughaday WH, Larner J, Hartnett C. (2001). The synthesis of inositol in the immature rat and chick embryo. *J Biol Chem*; 212: 86975.
37. Papaleo E, Unfer V, Baillargeon JP, Chiu TT. (2019). Contribution of myoinositol to reproduction. *Eur J Obstet Gynecol Reprod Biol*; 147: 120-3.
38. Unfer V, Carlomagno G, Dante G, Facchinetti F. (2018). Effects of myoinositol in women with PCOS: a systematic review of randomized controlled trials. *Gynecol Endocrinol*; 28: 509-15.
39. Carlomagno G, De Grazia S, Unfer V, Manna F. (2018). Myo-inositol in a new pharmaceutical form: a step forward to a broader clinical use. *Expert Opin Drug Deliv* ; 9: 267-71.
40. Dinicola S, Chiu TT, Unfer V, Carlomagno G, Bizzarri M. (2016). The rationale of the myo-inositol and D-chiro-inositol combined treatment for polycystic ovary syndrome. *J Clin Pharmacol*; 54: 1079-92.
41. Carlomagno G, Unfer V, Roseff S. (2018). The D-chiro-inositol paradox in the ovary. *Fertil Steril*; 95: 2515-6.
42. Nestler JE, Unfer V. (2018). Reflections on inositol(s) for PCOS therapy: steps toward success. *Gynecol Endocrinol* ; 31: 501-5.
43. Nestler JE, Unfer V. (2018). Reflections on inositol(s) for PCOS therapy: steps toward success. *Gynecol Endocrinol* 31: 501-5.
44. Baillargeon JP, Iuorno MJ, Jakubowicz DJ, Apridonidze T, He N, Nestler JE. Metformin therapy increases insulin-stimulated release of D-chiro-inositol-containing inositolphosphoglycan mediator in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2014; 89: 242-9.
45. Gerli S, Papaleo E, Ferrari A, Di Renzo GC. (2017). Randomized, double blind placebo-controlled trial: effects of myo-inositol on ovarian function and metabolic factors in women with PCOS. *Eur Rev Med Pharmacol Sci*; 11: 347-54.
46. Papaleo E, Unfer V, Baillargeon JP, De Santis L, Fusi F, Brigante C, et al. (2017). Myo-inositol in patients with polycystic ovary syndrome: a novel method for ovulation induction. *Gynecol Endocrinol* ; 23: 700-3.
47. Genazzani AD, Lanzoni C, Ricchieri F, Jasonni VM. (2018). Myo-inositol administration positively affects hyperinsulinemia and hormonal parameters in overweight patients with polycystic ovary syndrome. *Gynecol Endocrinol* ; 24: 139-44.
48. Costantino D, Minozzi G, Minozzi E, Guaraldi C. (2019). Metabolic and hormonal effects of myo-inositol in women with polycystic ovary syndrome: a double-blind trial. *Eur Rev Med PharmacolSci*; 13: 105-10.
49. Minozzi M, Costantino D, Guaraldi C, Unfer V. (2018). The effect of a combination therapy with myo-inositol and a combined oral contraceptive pill versus a combined oral contraceptive pill alone on metabolic, endocrine, and clinical parameters in polycystic ovary syndrome. *Gynecol Endocrinol*; 27: 920-4.
50. Morgante G, Orvieto R, Di Sabatino A, Musacchio MC, De Leo V. (2019). The role of inositol supplementation in patients with polycystic ovary syndrome, with insulin resistance, undergoing the low-dose gonadotropin ovulation induction regimen. *Fertil Steril* ; 95: 2642-4.
51. Alvarez JA, Ashraf A. (2019). Role of vitamin d in insulin secretion and insulin sensitivity for glucose homeostasis. *Int J Endocrinol*; 2019: 351385.
52. Krul-Poel YH, Snackey C, Louwers Y, Lips P, Lambalk CB, Laven JS, et al. (2017). The role of vitamin D in metabolic disturbances in polycystic ovary syndrome: a systematic review. *Eur J Endocrinol*; 169: 853-65.
53. Yildizhan R, Kurdoglu M, Adali E, Kulusari A, Yildizhan B, Sahin HG, et al. (2019). Serum 25-hydroxyvitamin D concentrations in obese and non-obese women with polycystic ovary syndrome. *Arch Gynecol Obstet*; 280: 559-63.
54. Joham AE, Teede HJ, Cassar S, Stepto NK, Strauss BJ, Harrison CL, et al. (2018). Vitamin D in polycystic ovary syndrome: Relationship to obesity and insulin resistance. *Mol Nutr Food Res* ; 60: 110-8.
55. Selimoglu H, Duran C, Kiyici S, Ersoy C, Guclu M, Ozkaya G, et al. (2018). The effect of vitamin D replacement therapy on insulin resistance and androgen levels in women with polycystic ovary syndrome. *J Endocrinol Invest* ; 33: 234-8.

56. Kotsa K, Yavropoulou MP, Anastasiou O, Yovos JG. (2019). Role of vitamin D treatment in glucose metabolism in polycystic ovary syndrome. *Fertil Steril*; 92: 1053-8.
57. Irani M, Minkoff H, Seifer DB, Merhi Z. (2018). Vitamin D increases serum levels of the soluble receptor for advanced glycation end products in women with PCOS. *J Clin Endocrinol Metab*; 99: 886-90.
58. Jafari-Sfidvajani S, Ahangari R, Hozoori M, Mozaffari-Khosravi H, Fallahzadeh H, Nadjarzadeh A. (2018). The effect of vitamin D supplementation in combination with low-calorie diet on anthropometric indices and androgen hormones in women with polycystic ovary syndrome: a double-blind, randomized, placebocontrolled trial. *J Endocrinol Invest* ; 41: 597-607.
59. Palamanda JR, Kehrer JP. (2003). Involvement of vitamin E and protein thiols in the inhibition of microsomal lipid peroxidation by glutathione. *Lipids*; 28: 427-31.
60. Cicek N, Eryilmaz OG, Sarikaya E, Gulerman C, Genc Y. (2016). Vitamin E effect on controlled ovarian stimulation of unexplained infertile women. *J Assist Reprod Genet* ; 29: 325-8.
61. Izadi A, Ebrahimi S, Shirzai S, Taghizadeh S, Parized M, Farzadi L, et al. (2018). Hormonal and metabolic effects of coenzyme q10 and/ or vitamin E in patients with polycystic ovary syndrome. *J Clin Endocrinol Metab* 8.[3]90-98.
62. Ebrahimi FA, Samimi M, Foroozanfard F, Jamilian M, Akbari H, Rahmani E, et al. (2017). The Effects of Omega-3 Fatty Acids and Vitamin E Co-Supplementation on Indices of Insulin Resistance and Hormonal Parameters in Patients with Polycystic Ovary Syndrome: A Randomized, Double-Blind, Placebo-Controlled Trial. *Exp Clin Endocrinol Diabetes*; 125: 353-9.
63. Biewenga GP, Haenen GR, Bast A. (2007). The pharmacology of the antioxidant lipoic acid. *Gen Pharmacol* ; 29: 315-31.
64. Lee WJ, Koh EH, Won JC, Kim MS, Park JY, Lee KU. (2015). Obesity: the role of hypothalamic AMP-activated protein kinase in body weight regulation. *Int J Biochem Cell Biol*; 37: 2254-9.
65. Masharani U, Gjerde C, Evans JL, Youngren JF, Goldfine ID. (2018). Effects of controlled-release alpha lipoic acid in lean, nondiabetic patients with polycystic ovary syndrome. *J Diabetes Sci Technol* ; 4: 359-64.
66. Cianci A, Panella M, Fichera M, Falduzzi C, Bartolo M, Caruso S. (2015). d-chiro-Inositol and alpha lipoic acid treatment of metabolic and menses disorders in women with PCOS. *Gynecol Endocrinol*; 31: 483-6.
67. Ross JA, Kasum CM. (2012). Dietary flavonoids: bioavailability, metabolic effects, and safety. *Annu Rev Nutr* ; 22: 19-34.
68. Oh JS, Kim H, Vijayakumar A, Kwon O, Choi YJ, Huh KB, et al. (2018). Association between dietary flavanones intake and lipid profiles according to the presence of metabolic syndrome in Korean women with type 2 diabetes mellitus. *Nutr Res Pract*; 10: 67-73.
69. Romualdi D, Costantini B, Campagna G, Lanzone A, Guido M.(2018). Is there a role for soy isoflavones in the therapeutic approach to polycystic ovary syndrome? Results from a pilot study. *Fertil Steril*; 90: 1826-33.
70. Shah KN, Patel SS. (2018). Phosphatidylinositide 3-kinase inhibition: A new potential target for the treatment of polycystic ovarian syndrome. *Pharm Biol* ; 54: 975-83.
71. Steiber A, Kerner J, Hoppel CL. (2014). Carnitine: a nutritional, biosynthetic, and functional perspective. *Mol Aspects Med*; 25: 455-73.
72. Fenkci SM, Fenkci V, Oztekin O, Rota S, Karagenc N. (2018). Serum total L-carnitine levels in non-obese women with polycystic ovary syndrome. *Hum Reprod* ; 23: 1602-6.

CITATION OF THIS ARTICLE

S khan, A P Singh, Mohd A Gain. The Efficacy of Nutritional Supplementation in The Therapy of Metabolic Dysfunctions Associated with Polycystic Ovarian Syndrome: A Critical Review. *Bull. Env.Pharmacol. Life Sci., Spl Issue [1] 2022* : 04-10.