



Effect of salinity stress on proline content in *Prosopis cineraria* and *Prosopis juliflora*

Monika Sharma¹, Rachana Dinesh^{2,*}, Seema Sen²

¹Research scholar, Department of Botany, Jai Narain Vyas University, Jodhpur – Raj. (India)

²Assistant Professor, Department of Botany, Jai Narain Vyas University, Jodhpur – Raj. (India)

*Email: rachanadinesh.dinesh@gmail.com

ABSTRACT

The objective of the present study is to find out the effect of salinity stress on proline content in two species of genus Prosopis i.e., Prosopis cineraria and Prosopis juliflora. Prosopis is a plant that is well adapted to grow in the challenging conditions of arid and semi-arid region. Proline plays an important role in mitigating the abiotic stress conditions in plants. Therefore, experiments were designed to find out the role of proline and its content in Prosopis cineraria and Prosopis juliflora. Proline content was determined by method given by Bates et al. (1973). Proline content of the seedling under different salinity treatments (ranging from 0.0mM to 300.00mM concentrations) were observed after two weeks and then a regular interval. The observations showed that with increase in salinity, the level of proline increased in both the species up to a certain limit. After that when the salinity was increased further the proline content starts decreasing. In case of Prosopis cineraria as well as Prosopis juliflora proline content was highest at 150mM salt concentration that is 10.46±0.41^a and 12.97±0.13^a respectively. The proline content in Prosopis juliflora was higher than Prosopis cineraria, in controlled as well as in different salt treatment.

Keywords: salinity stress, proline content, *Prosopis cineraria*, *Prosopis juliflora*, arid region

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INTRODUCTION

Prosopis is a very important plant of arid and semi-arid regions. Survival of this genus in the adverse environmental conditions of desert make it an important plant of this region. Timbers of these plants are used as fuel and for many other purposes. Pods of these trees are highly nutritious, used to feed the cattle. Pods of Prosopis cineraria are also consumed by local people, it becomes the only food during the famine in these regions. The two species of Prosopis, Prosopis cineraria and Prosopis juliflora are the lifeline trees of the arid region. Prosopis cineraria is worshiped by many communities of these regions for its usefulness and hence is called 'Kalpvriksha' [1], [2]. It withstands lots of abiotic stress conditions which is offered to this plant by nature. This makes these plants interesting for physiological studies [3]. Not many reports are available on the study of physiological parameters of this genus. Both of these plants can tolerate a high level of soil salinity. Sodium chloride contributes largely to soil salinity [4]. Salinity leads to the accumulation of Na⁺ and Cl⁻ ions and causes the water deficiency in plants. This osmotic stress ultimately leads to the death of plants [5]. To avoid the osmotic stress plant requires the osmo-protectors. Osmo-protectors are the nontoxic molecules which accumulate in the plant cell up to the required level to protect it from dehydration without affecting the metabolism of plants [6]. Synthesis of some macromolecules such as proline is affected in response to salt stress [7]. Proline is an osmolyte which actively participate to maintain the osmotic balance in plants. Proline is known to be the only osmolyte which can scavenge singlet oxygen as well as free radicals such as hydroxyl ions (OH⁻) [5]. It increases the turgor to help the plant in cell expansion [8], [9]. Proline helps in biochemical reactions [10], [11] and does not have any adverse effect on enzymatic activity [12]. Plants are having the ability to accumulate as well as degrade proline quickly when required [13], [14]. According to the study of many researchers, accumulation of proline is a physiological process which increases under abiotic stress [15], [16], [17], [18], [19]. This indicates that proline is a sign of stress resistance. During salt stress the osmotic adjustment is primarily the function of proline [20], [21]. Synthesis of proline takes place under Glu pathway which dominates under stress [22]. It protects proteins, DNA and other macromolecules and also helps to regulate redox potential. Enzyme denaturation due to stress can be reduced by proline [8]. Proline biosynthesis, accumulation and transportation under salinity stress studied by KaviKishor and Sreenivasulu, 2014 [23]. There are many researchers who believes that proline accumulation often related with the salt

tolerance. According to them salt tolerant genotype shows higher accumulation of proline as compare to the salt sensitive ones. According to [24] salinity does not affect the proline accumulation significantly. It is reported by [5] that over-expression of an enzyme, P5CS (D1-pyrroline-5-carboxylate synthetase) in transgenics enhances oxidative stress tolerance caused by drought and salt stresses. This is the enzyme used in the rate limiting step of proline biosynthesis reaction. Over expression of this enzyme during stress which leads to proline accumulation and enhanced stress tolerance reveals that proline has some role in stress tolerance. This is supported by different scientists in different transgenics plants such as in rice [25], [26], chickpea [27], tobacco [28], wheat [29], potato [30]. The purpose of this study is to investigate the effect of salinity on proline content in *Prosopis cineraria* and *Prosopis juliflora* and to find out that if proline plays some role in the salt tolerance of these plants.

MATERIAL AND METHODS

To study the proline content under control and stress condition, 500 seeds of each of the selected plants (*Prosopis cineraria* & *Prosopis juliflora*) were soaked in separate beakers. The imbibed seeds were transferred to the petri dishes of 5cm diameter containing three layers of moist filter papers. After seven days the germinated seeds with a length more than 3.5cm were transferred to the pots containing Hoagland solution. Ten seedlings were placed in each pot. After two weeks (when each of the plants got at least one leaf) the Hoagland solution was supplemented with salt solution (i.e., Hoagland with different salt concentrations). Proline content of the seedling under different salinity treatments (control, 25mM, 50mM, 75 mM, 100 mM, 125 mM, 150 mM, 175 mM, 200 mM, 250 mM, 300 mM) were observed after two and four weeks. For this the leaves of the seedling under control and different salt concentrations were harvested after two and four weeks.

Proline determination

Proline content was estimated by total proline content determination assay by Bates et al. (1973) [31]. Standard curve was prepared using proline. Plant extract was prepared by blending 250mg of fresh leaves of the treated plants with 5ml of 3% sulpho-salicylic acid. The mixture was centrifuged at 10,000 rpm for 20 minutes. 2ml of supernatant was mixed with equal amount of glacial acetic acid and ninhydrin. The mixture was boiled for 1hour. After cooling 4ml of toluene reagent was added using cyclo-mixture. Optical density of the upper layer was read using spectrophotometer at 520nm. The experiment was done in triplets and was repeated five times.

RESULT

Proline content:

The experiments were conducted to determine the proline content of the plants (*Prosopis cineraria* and *Prosopis juliflora*) after two and four weeks. Initially the proline content in the plants under study increased with the increase in salinity and time, it started decreasing when the salinity increased beyond 150 mM (fig. a, b). The proline content was always higher in *Prosopis juliflora* than *Prosopis cineraria* (fig. c) The increase in the particular treatments was also more prominent in case of *Prosopis juliflora*. Under controlled condition the proline content in *Prosopis juliflora* were 5.47 ± 0.31^g , 6.16 ± 0.13^h while in *Prosopis cineraria* 3.43 ± 0.28^h , 4.69 ± 0.22^f after two and four weeks respectively. After two weeks proline content was recorded 4.29 ± 0.26^g and 6.33 ± 0.11^f in *Prosopis cineraria* and *Prosopis juliflora* respectively under 25 mM salt concentration. At 50mM salt concentration proline content was 5.74 ± 0.17^f and 6.86 ± 0.05^e in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 75 mM salt concentration proline content was 7.24 ± 0.20^d and 7.70 ± 0.16^d in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 100 mM salt concentration also proline content was higher in *Prosopis juliflora* 8.52 ± 0.19^c as compared to *Prosopis cineraria* 7.80 ± 0.12^c . Proline content recorded at 125 mM was higher in *Prosopis juliflora* 10.6 ± 0.31^b as compared to *Prosopis cineraria* 8.39 ± 0.25^b . At 150 mM salt concentration proline content was 10.46 ± 0.41^a and 12.47 ± 0.34^a in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 175 mM salt concentration proline content was 6.33 ± 0.28^e and 7.41 ± 0.33^d in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 200mM salt concentration also proline content was higher in *Prosopis juliflora* 4.39 ± 0.32^h as compared to *Prosopis cineraria* 5.37 ± 0.38^f . After four weeks proline content was recorded 4.69 ± 0.22^f and 7.26 ± 0.16^f in *Prosopis cineraria* and *Prosopis juliflora* respectively under 25 mM salt concentration. At 50mM salt concentration proline content was 6.69 ± 0.22^d and 7.81 ± 0.17^e in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 75 mM salt concentration proline content was 8.62 ± 0.31^c and 8.56 ± 0.23^d in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 100 mM salt concentration also proline content was higher in *Prosopis juliflora* 9.77 ± 0.15^c as compared to *Prosopis cineraria* 9.32 ± 0.29^b . Proline content recorded at 125 mM was higher in *Prosopis juliflora* 11.50 ± 0.28^b as compared to *Prosopis cineraria* 10.28 ± 0.23^a . At 150 mM salt concentration proline content was 9.60 ± 0.25^b and 12.97 ± 0.13^a in *Prosopis cineraria* and *Prosopis juliflora* respectively. At

175 mM salt concentration proline content was 5.54 ± 0.32^e and 6.76 ± 0.47^s in *Prosopis cineraria* and *Prosopis juliflora* respectively. At 200mM salt concentration proline content was 3.50 ± 0.33^i in *Prosopis juliflora*, *Prosopiscineraria* seedling did not survive at 250mM salt concentration. In *Prosopis juliflora* proline content was highest under 150mM (12.97 ± 0.13^a) (Table. 1), lowest under 250mM (3.21 ± 0.14^i) after two weeks. In case of *Prosopis cineraria* proline content was highest (10.46 ± 0.41^a) under 150mM salt concentration and lowest (5.37 ± 0.38^f) under 200mM after two weeks (Table. 2).

Table. 1 Proline content of *P. cineraria*

	N	<i>P. cineraria</i> after two weeks	Std. Error	<i>P. cineraria</i> after four weeks	Std. Error
Control	8	3.43 ± 0.28^h	0.10	4.69 ± 0.22^f	0.08
25	8	4.29 ± 0.26^g	0.09	5.34 ± 0.30^e	0.11
50	8	5.74 ± 0.17^f	0.06	6.69 ± 0.22^d	0.08
75	8	7.24 ± 0.20^d	0.07	8.62 ± 0.31^c	0.11
100	8	7.80 ± 0.12^c	0.04	9.32 ± 0.29^b	0.10
125	8	8.39 ± 0.25^b	0.09	10.28 ± 0.23^a	0.08
150	8	10.46 ± 0.41^a	0.15	9.60 ± 0.25^b	0.09
175	8	6.33 ± 0.28^e	0.10	5.54 ± 0.32^e	0.11
200	8	5.37 ± 0.38^f	0.14	0.00 ± 0.00^g	0.00
250	8	0.00 ± 0.00^i	0.00	0.00 ± 0.00^g	0.00

Table. 2 Proline content of *P. juliflora*

	N	<i>P. juliflora</i> after two weeks	Std. Error	<i>P. juliflora</i> after four weeks	Std. Error
Control	8	5.47 ± 0.31^g	0.11	6.16 ± 0.13^h	0.05
25	8	6.33 ± 0.11^f	0.04	7.26 ± 0.16^f	0.06
50	8	6.86 ± 0.05^e	0.02	7.81 ± 0.17^e	0.06
75	8	7.70 ± 0.16^d	0.06	8.56 ± 0.23^d	0.08
100	8	8.52 ± 0.19^c	0.07	9.77 ± 0.15^c	0.05
125	8	10.6 ± 0.31^b	0.11	11.50 ± 0.28^b	0.10
150	8	12.47 ± 0.34^a	0.12	12.97 ± 0.13^a	0.04
175	8	7.41 ± 0.33^d	0.12	6.76 ± 0.47^s	0.17
200	8	4.39 ± 0.32^h	0.11	3.50 ± 0.33^i	0.12
250	8	3.97 ± 0.11^i	0.04	3.21 ± 0.14^i	0.05

Figure 1. Proline content of *P. juliflora*

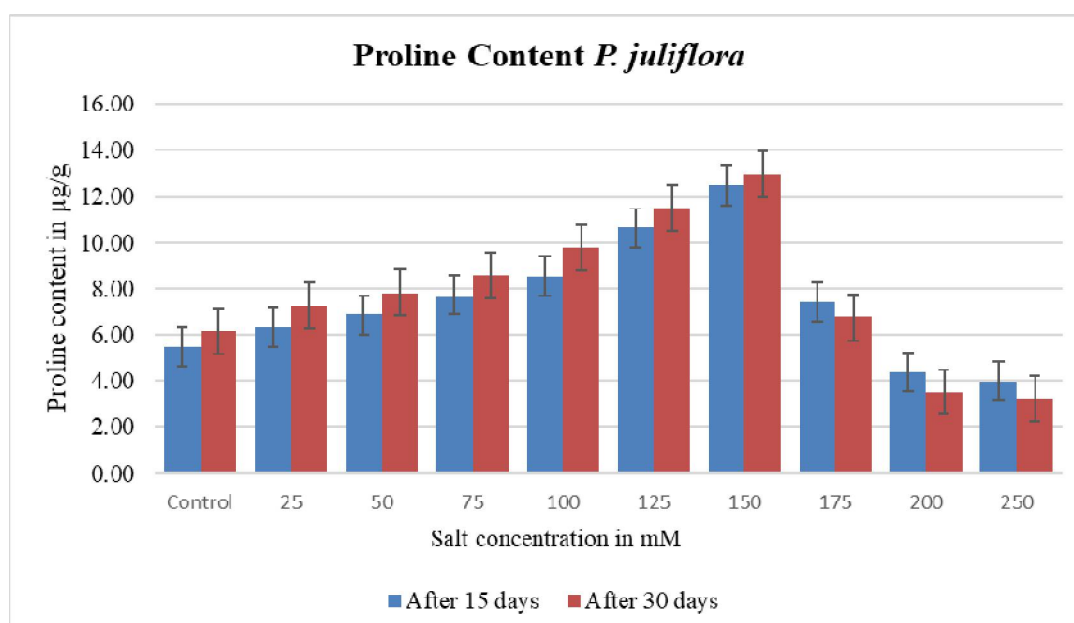
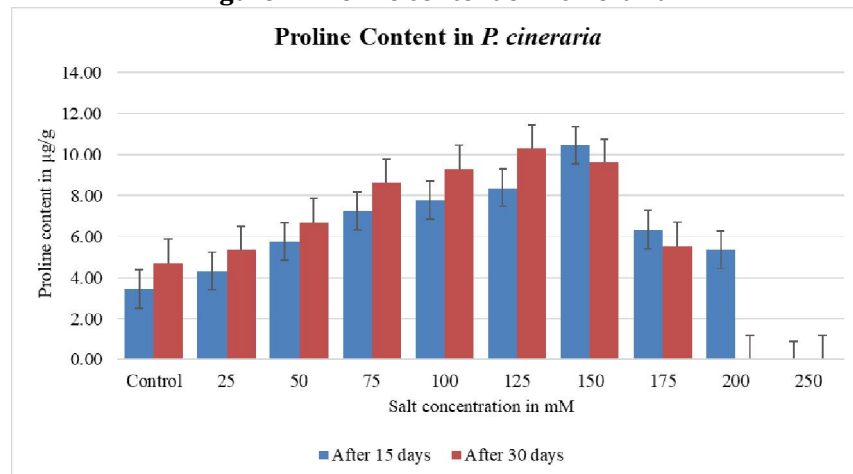
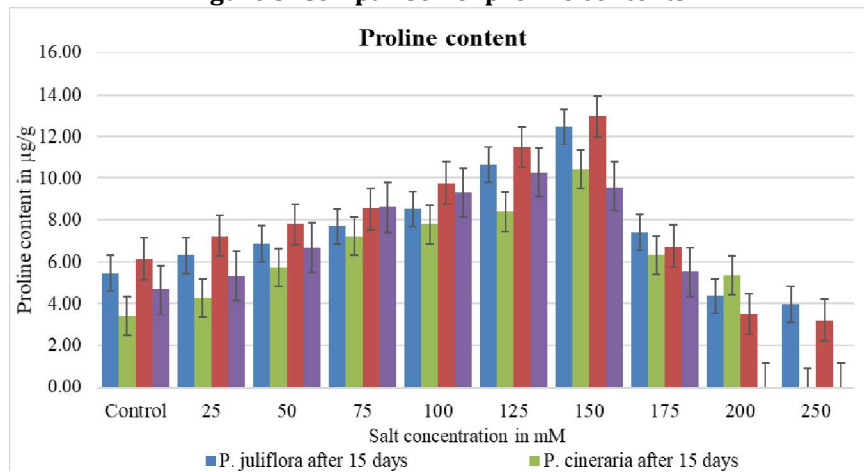


Figure 1. Proline content of *P.cineraria***Figure 3. Comparison of proline contents**

DISCUSSION

Accumulation of proline as osmolyte is the most common phenomenon used by the plants and other organisms to maintain their osmotic stability [32]. The scientists believe that proline accumulation makes the plants more tolerant and somehow helps the plant to overcome stress, here are some supporting views by different scientists. Proline is a compatible solute [12], must have some important role in osmotic adjustment during stress condition [33], [34] that is why metabolic stress leads to proline accumulation [35]. According to [36] proline can improve salt tolerance. Even the exogenous application of proline can improve the plant growth [37]. It was reported by [38] that foliar application of proline showed positive effect on plant growth and yield characteristics under salt stress. The level of proline accumulation was found higher in the tolerant plant species as compared to the sensitive ones. [23], [39], [40]. Increased proline content was recorded with increasing salinity in *Phaseolus vulgaris*, L. [41]. Proline content was found to increase in maize when treated with NaCl concentrations higher than 100mM [42]. Proline content of rice seedlings increased with the increase in salinity [43]. Proline content increased with the salt concentration in *Acacia auriculiformis* when this plant species was treated with different salt concentrations (0.3, 3.9, 6.0, 7.9, 10.0, 12.1, and 13.9 dS m⁻¹) [44]. The similar results were reported in tobacco [45], [28]), sorghum [46], green gram [47], wheat [48] and mulberry [49]. On the other hand, some scientists found the contrasting results. They recorded that excess of salinity inhibits the proline accumulation in Citrus [34]. Excessive use of proline could be the reason behind the decline in proline content under higher salinity stress. This view is supported by many scientists in Sorghum [50], rice [51] cotton [52], tomato [53]. The above studies explain the increase in proline content with the increase of salinity and then decrease in proline content at higher salinity in the plant under the present study.

CONCLUSION

The present studies reveal that proline has an important role in mitigating stress in many plant species. Change in level of proline content is directly related to stress condition. The *Prosopis juliflora* is more

tolerant as compare to *Prosopis cineraria*. As the proline content of *P. juliflora* were reported higher as compare to *P. cineraria* under all the treatment as well as controlled conditions.

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REFERENCES

- Rani B, Singh U, Sharma R, Gupta A, Dhawan N, Sharma A and Maheshwari R (2013). *Prosopis cineraria* (L.) druce: a desert tree to brace livelihood in Rajasthan. *Asian Journal of Pharmaceutical Research & Health Care* 5(2):58.
- Pareek AK, Garg S and Kuma M (2016). *Prosopis cineraria*: a gift of nature for pharmacy. *International Journal of Pharmaceutical* 6(6):958- 964.
- Khaliq T, Hussain N, Ali A, Ullah A, Ahmad M, Ahmad A (2016). Quantification of root-shoot development and water use efficiency in autumn maize (*Zea mays* L.) under different irrigation strategies. *J. Environ. Agric. Sci.*6:16-22.
- Koca H, Bor M, Ozdemir F, Turkan I (2007). The effect of salt stress on lipid peroxidation, antioxidative enzymes and proline content of sesame cultivars. *Environmental and Experimental Botany* 60:344-351.
- Surekha CH, Kumari KN, Aruna LV, Suneetha G, Arundhati A, Kishor PK (2014). Expression of the *Vigna acconitifolia* P5CSF129A gene in transgenic pigeon pea enhances proline accumulation and salt tolerance. *Plant Cell, Tissue and Organ Culture (PCTOC)*116(1):27-36.
- Yamaguchi-Shinozaki K, Shinozaki K (2005). Organization of cisacting regulatory elements in osmotic- and cold-stress-responsive promoters. *Trends Plant Sci* 10:88-94
- Santos CLV, Campos A, Azevedo H, Caldeira G (2001). In situ and in vitro senescence induced by KCl stress: nutritional imbalance, lipid peroxidation and antioxidant mechanisms. *J Exp Bot* 52:351-360.
- Matysik J, Alia, Bhalu B, Mohanty P (2002). Molecular mechanisms of quenching of reactive oxygen species by proline under stress in plants. *Current Science* 10:525-32.
- Sairam RK, Tyagi A (2004). Physiology and molecular biology of salinity stress tolerance in plants. *Current science* 10:407-21.
- Tester M and Davenport R (2003). Na⁺ tolerance and Na⁺ transport in higher plants. *Annals of Botany* 91:503-527.
- Cavagnaro TR, Jackson LE, Six J, Ferris H, Goyal S, Asami D, Scow KM (2006). Arbuscular mycorrhizas, microbial communities, nutrient availability, and soil aggregates in organic tomato production. *Plant & Soil* 282:209-225.
- Rathinasabapathi B (2000). Metabolic Engineering for Stress Tolerance: Installing Osmoprotectant Synthesis Pathways. *Annals of Botany*. 86. 10.1006/anbo.2000.1254.
- Trovato M, Mattioli R, Costantino P (2008). Multiple roles of proline in plant stress tolerance and development. *RendicontiLincei* 19:325-346
- Dar MI, Naikoo MI, Rehman F, Naushin F, Khan FA (2016). Proline accumulation in plants: roles in stress tolerance and plant development. In *osmolytes and plants acclimation to changing environment: emerging omics technologies* 155-166.
- Ahmad S, Ullah F, Sadiq A, Ayaz M, Imran M, Ali I, Zeb A, Ullah F, Shah MR (2016). Chemical composition, antioxidant and anticholinesterase potentials of essential oil of *Rumex hastatus* D. Don collected from the North West of Pakistan. *BMC complementary and alternative medicine* 16(1):1-1.
- Nahar K, Rahman M, Hasanuzzaman M, Alam MM, Rahman A, Suzuki T, Fujita M (2016). Physiological and biochemical mechanisms of spermine-induced cadmium stress tolerance in mung bean (*Vigna radiata* L.) seedlings. *Environmental Science and Pollution Research* 23(21):21206-18.
- Hayat S, Hayat Q, Alyemini MN, Wani AS, Pichtel J, Ahmad A (2012). Role of proline under changing environments: a review. *Plant signaling & behavior*. 7(11):1456-1466.
- Parida AK, Das AB (2005). Salt tolerance and salinity effects on plants: a review. *Ecotoxicology and environmental safety* 1;60(3):324-49.
- Mansour MM (2000). Nitrogen containing compounds and adaptation of plants to salinity stress. *BiologiaPlantarum* 43(4):491-500.
- Mansour MM, Stadelmann EJ (2014). NaCl-induced changes in protoplasmic characteristics of *Hordeum vulgare* cultivars differing in salt tolerance. *PhysiologiaPlantarum* 91(3):389-394.
- Hasegawa PM, Bressan RA, Zhu JK, Bohnert HJ (2000). Plant cellular and molecular responses to high salinity. *Annual review of plant biology*. 51(1):463-99.
- Yamada M, Morishita H, Urano K, Shiozaki N, Yamaguchi-Shinozaki K, Shinozaki K, Yoshida Y (2005). Effects of free proline accumulation in petunias under drought stress. *Journal of Experimental Botany* 56(417):1975-1981.
- KaviKishor PB, Sreenivasulu N (2014). Is proline accumulation per se correlated with stress tolerance or is proline homeostasis a more critical issue *Plant, cell & environment* 37(2):300-11.
- Datta P, Kulkarni M (2014). Arbuscularmycorrhizal colonization improves growth and biochemical profile in *Acacia arabica* under salt stress. *Journal of Bio Science & Biotechnology*1;3(3).
- Su J, Wu R (2004) Stress-inducible synthesis of proline in transgenic rice confers faster growth under stress conditions than that with constitutive synthesis. *Plant Sci* 166:941–948

26. Anoop N, Gupta AK (2003) Transgenic indica rice cv IR-50 over expressing *Vigna aconitifolia* d (1)-pyrroline-5-carboxylate synthetase cDNA shows tolerance to high salt. *J Plant Biochem Biotechnol* 12:109-116
27. Kiran Kumar Ghanti S, Sujata KG, Vijay Kumar BM, Nataraja Karba N, Janardhan Reddy K, Srinath Rao M, KaviKishor PB (2011) Heterologous expression of P5CS gene in chickpea enhances salt tolerance without affecting yield. *Biol Plant* 55(4):634-640
28. Yamchi A, Jazii FR, Mousav A, Karkhane AA, Renu (2007) Proline accumulation in transgenic tobacco as a result of expression of Arabidopsis D1-pyrroline-5-carboxylate synthetase (P5CS) during osmotic stress. *J Plant BiochemBiotechnol* 16:9-15
29. Vendruscolo ECG, Schuster I, Pileggi M, Scapim CA, Molinari HBC, Marur CJ, Vieira LGE (2007) Stress-induced synthesis of proline confers tolerance to water deficit in transgenic wheat. *J Plant Physiol* 16:1367-1376
30. Hmida-Sayari A, Gargouri-Bouzzid R, Bidani A, Jaoua L, Savoure A, Jaoua S (2005) Overexpression of D1-pyrroline-5-carboxylate synthetase increases proline production and confers salt tolerance in transgenic potato plants. *Plant Sci* 169:746-752
31. Bates LS, Waldran RP, Teare ID (1973). Rapid determination of free proline for water studies. *Plant Soil* 39: 205-208
32. Bartels D and Sunkar R (2005). Drought and salt tolerance in plants. *Crit. Rev. Plant Sci.*24:1-36.
33. Mademba F, Boucherea UA & Larher FR (2003). Proline accumulation in cultivated citrus and its relationship with salt tolerance *J. Hort. Sci. Biotechnol.* 78:617-623.
34. Cushman JC (2001). Crasulacean acid metabolism. A plastic photosynthetic adaptation to arid environments. *Plant Physiol.* 127:1439-1448
35. Heuer B (2010). Role of proline in plant response to drought and salinity. in *Handbook of Plant and Crop Stress* 3:213-238.
36. El Moukhtari A, Cabassa-Hourton C, Farissi M, Savouré A (2020). How does proline treatment promote salt stress tolerance during crop plant development? *Front. Plant Sci.*11:1127.
37. Alam, R, Das D, Islam M, Murata Y, Hoque M (2016). Exogenous proline enhances nutrient uptake and confers tolerance to salt stress in maize (*Zea mays* L.). *Progr. Agric.* 27:409-417.
38. Heidari B (2011). Knee osteoarthritis prevalence, risk factors, pathogenesis and features: Part I. *Caspian journal of internal medicine.* 2(2):205.
39. Demiral T, Türkan I (2006). Exogenous glycine betaine affects growth and proline accumulation and retards senescence in two rice cultivars under NaCl stress. *Environmental and Experimental Botany*1;56(1):72-9.
40. Stoeva N, Kaymakanova M. Effect of salt stress on the growth and photosynthesis rate of bean plants (*Phaseolus vulgaris* L.). *Journal of Central European Agriculture.* 2008 Nov 28;9(3):385-91.
41. Camara TR, Willadino L, Torne AM, and Santos MA (2000). Efeito do estresse salino e da prolina exógena em calos de milho," *Revista Brasileira De Fisiologia Vegetal.* 12:146-155.
42. Pushpam R and Rangasamy SRS (2000). Effect of salinity on protein and proline contents of callus and seedlings of rice. *Crop Research* 20:192-196.
43. Patel AD, Jadeja HR & Pandey AN (2010). Effect of soil salinity on growth, water status and nutrient accumulation in seedlings of *Acacia auriculiformis* (Fabaceae). *Journal of Plant Nutrition* 33:914-932.
44. Gubis J, Vankova R, Cervena V, Dragunova M, Hudcovicova M, Lichtnerova H, Dokupil T, Jurekova Z (2007) Transformed tobacco plants with increased tolerance to drought. *S Afr J Bot* 73:505-511
45. Jogeswar G, Pallela R, Jakka NM, Reddy PS, Rao JV, Sreenivasulu N, KaviKishor PB (2006) Antioxidative response in different sorghum species under short term salinity stress. *ActaPhysiol Plant* 28:465-475
46. Misra N, Gupta AK (2005) Effect of salt stress on proline metabolism in two high yielding genotypes of green gram. *Plant Sci* 169:331-339
47. Sairam RK, Srivastava GC, Agarwal S, Meena RC. Differences in antioxidant activity in response to salinity stress in tolerant and susceptible wheat genotypes. *Biologia Plantarum.* 2005 Mar;49(1):85-91.
48. Kumar SG, Reddy A, Sudhakar C (2003) NaCl effects on proline metabolism in two high yielding genotypes of mulberry (*Morus alba* L.) with contrasting salt tolerance. *Plant Sci* 165:1245-1251
49. Lutts S, Kinet JM and Bouharmont J (1996). Effects of various salts and of mannitol on ion and proline accumulation in relation to osmotic adjustment in rice (*Oryza sativa* L.) callus cultures. *J. Plant Physiol.*149:186-195.
50. Feitosa FL, Birgel EH, Mirandola RM, Perri SH. Diagnosis of failure of passive transfer of immunity in the calf by determining total protein and its electrophoretic fractions, immunoglobulins and serum gammaglutamyl transferase activity. *Ciência Rural.* 2001 Apr;31(2):251-5.
51. Meloni DA, Oliva MA, Ruiz HA, Martinez CA. (2001). Contribution of proline and inorganic solutes to osmotic adjustment in cotton under salt stress. *Journal of Plant Nutrition.*28;24(3):599-612.
52. Heuer B (2003). Influence of exogenous application of proline and glycine betaine on growth of salt-stressed tomato plants. *Plant Science* 1;165(4):693-9..

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