



ORIGINAL ARTICLE

Evaluating SiO₂ Nanoparticles Effects on Developmental Characteristic and Photosynthetic Pigment Contents of *Zea mays* L.

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ABSTRACT

*The usage of the nanoparticle is increasing in different fields, but till yet, very limited studies have been performed as far as consequence of nanomaterials and plant interactions is concerned. In this study, the effects of silica nanoparticles (SiO₂) on developmental stages of *Zea mays* L. viz. seed germination, rate of root and stem elongation, relative water content (RWC) and photosynthetic pigment content have been investigated. The results exhibited that exposure to the silica nanoparticles at 0, 400, 2000, and 4000 mg/L concentrations could significantly ($p < 0.05$) increase the root elongation and seed germination in comparison to control. In addition, shoot elongation, relative water content and photosynthetic pigment content, differed between treated and control samples.*

Keywords: Developmental characteristic, SiO₂ Nanoparticles, Photosynthetic pigment, *Zea mays* L.

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INTRODUCTION

Nanoparticles are exploring a wide range of applications in biomedical sciences, drug delivery, gene therapy, cell targeting, magnetics, optics, mechanics, catalysis and energy science [1-4]. In the past few years, nanomaterials and nanotechnology have been extensively used in various industry and commerce areas in many countries. Nanoparticles are atomic or molecular aggregates with at least one dimension between 1 and 100 nm, contrastingly, nanomaterials are great and versatile group of materials in which one or several dimensions are of size 1-100 nm [5]. This size can significantly change their properties compared to the bulk material [6]. Nanotechnology has an ability to improvise the agricultural and food industry with new tools like instant disease detectors and increasing plant's ability to absorb nutrients [7]. The exclusive properties of these materials, such as a large specific surface area and greater reactivity, have raised questions concerning potential adverse effects on human and environmental health [8]. In spite of the extensive growth of nanotechnology and nanomaterials through the last 20 years, the recent focus has been turned on to the potential toxicological effects on humans, animals, and the environment through the exposure of metal nanomaterials. Until now only a very limited number of phyto and eco-toxicological studies have been performed. The results of these studies have been reported

with an aim to supply more insight into the correlations between plants and nanomaterials. At present there is an extensive discussion in relation to the risks and benefits of the many nonmaterial into the environment [9]. Because of the widespread utilization of these products it is expected that nanoparticles will soon find their way into aquatic, terrestrial and atmosphere environments, where their fate and behavior are largely unknown. Therefore, it demands research related risk assessment evaluation in order to clarify all related aspects of the concern, but risk assessment is quite a difficult task for nanomaterials since an insignificant research has been carried out. Moreover, there are many unknown effects that nanomaterials have on ecosystems. The aim of this study is evaluating the effect of SiO₂ nanoparticle on *Zea mays* L. growth.

MATERIALS AND METHODS

Nanoparticle

The nanosilicon dioxide was procured from Nanoamor Co., Iran with the specifications shown in Figure 1 and Table 1. Average particle size of experimental nanoparticle was 20 nm and purity was 99.5% (Table 1). The shape of SiO₂ nanoparticles was spherical (Fig 1). The nanoparticles were dispersed in distilled water at four concentrations viz. 0, 400, 2000 and 4000 mg/L and then sterilized at 120°C for 30 minutes. Ultrasonicator was used for easy dispersion of nanoparticles without precipitation. There was sufficient shaking to break up agglomerates. The treatment concentration for each nanoparticle was prepared separately, devoid of dilutions. The particles were weighed and dispersed in water. The nanoparticles suspensions were dispersed under suction for 20 minutes before use.

Seeds

Seeds of *Zea mays* L. were surface sterilized in 5% (w/v) sodium hypochlorite with 20 minute incubation period. After three washes with distilled water, the seeds were germinated on wet filter paper in sterile Petri dishes. Corn seeds were grown hydroponically, in solution with known concentrations of SiO₂ nanoparticles. The growth of plants was visually observed and recorded (Fig 2). Germination percentages were determined by comparing the numbers of seeds that developed a primary root of at least 1 mm to the total number of seeds planted in each dish. Relative root growth inhibition was calculated as the difference between the average primary root length of the unexposed control plants and treatment plants, average root length divided by the primary root length of the control.

Relative water content (RWC) measurement

To determine the relative water content of different parts of the plant, the leaf samples were removed and their fresh weight (FW), dry weight (DW) and turgid weight (TW) was determined. Leaf relative water content, was measured by soaking leaf sample (0.5 g) in 100 ml of distilled water at 4°C in the dark for 24 h. The turgid leaves were quickly blotted dry prior to the turgid weight measurement. Dry weight of leaves was determined after oven-drying at 70°C for 48 h. The relative water content (RCW) was determined by subtracting fresh weight from dry weight and multiplying this number by the difference between turgid weight and dry weight. RWC was calculated according to Smart and Bingham [10], using the following equation:

$$RWC = [\text{fresh weight} - \text{dry weight} / \text{turgid weight} - \text{dry weight}] \times 100$$

Photosynthetic Pigment Measurement

The content of photosynthetic pigments was determined according to the method of Lichtenthaler and Wellburn [11]. Two hundred mg of leaf tissue was weighed and powdered using liquid nitrogen. After adding 80% acetone, the volume was brought to 25 ml. The resulting solution was centrifuged at 4800 rpm for 20 min. The supernatant was used for measurement of chlorophyll a, b and the carotenoids. Observance of the clear supernatant was read at 663.2, 646.8 and 470 NM (Shimadzu spectroscopy, A160 model, Japan) and pigment concentrations were calculated using the following formula:

$$\text{Chl a } (\mu\text{g/ml}) = 12.25 * A_{663.2} - 2.79 * A_{646.8}$$

$$\text{Chl b } (\mu\text{g/ml}) = 21.5 * A_{646.8} - 5.1 * A_{663.2}$$

$$\text{Chl a + Chl b } (\mu\text{g/ml}) = 7.15 * A_{663.2} + 18.71 * A_{646.8}$$

$$B\text{-carotene } (\mu\text{g/ml}) = (1000 * A_{470} - 1.82 * Ca - 85.02 * Cb) / 198$$

In this formula Chl a, Chl b, Chl T and CX+C are, chlorophyll a, chlorophyll b, total chlorophyll and carotenoid concentrations respectively.

Statistical Analysis

The experimental designs were randomized complete block and each value reported is the average of three replicates. The raw data were imported into Microsoft Excel 2007 program for calculations and graphical representation. SPSS (version 11.5) software was used for analysis of variance. Quantitative changes of different parameters were analyzed through analysis of variance (ANOVA), with Duncan's multiple range tests at $P < 0.05$ being used to determine significant differences among treatments. All

consequences are presented as the mean \pm standard deviation (SD). The result was considered significant if $P < 0.05$, when compared to the control.

RESULTS AND DISCUSSION

Effect of SiO₂ Nanoparticles on Seed Germination

Based on experimental results, it was proclaimed that seeds uptake the SiO₂ nanoparticles from the hydroponic solution. This study concluded that the nanoparticles enhanced the growth of the seed resulting in longer root elongation compared to a control group (Fig 3). By increasing the SiO₂ nanoparticles concentration, seed germination elevated at 400 mg/L but decreased at 2000, and 4000 mg/L respectively.

Effect of SiO₂ Nanoparticles on Root and Shoot Elongation

SiO₂ nanoparticles enhanced the growth of the root and seed elongation when compared to a control group (Fig 4). The result shows that with increasing concentration of nanoparticles at 0 to 4000 mg/L, size of largest root and mid-spaces of two root lengths increased but the size of the shoot decreased. This result illustrated that SiO₂ nanoparticles have positive effects on root and negative effects on shoot elongation.

Effect of SiO₂ nanoparticles on Plant RWC

Relative water content (RWC) is an index demonstrating the amount of water in the plant organs and shows the ability of a plant in maintaining water under stress conditions. Therefore, in an experimental controlled environment, the measured RWC clearly showed the response of a plant. The higher the measured amount, the greater the ability of the treatment to preserve water [12]. RWC in plant decreased by SiO₂ nanoparticles at 400 mg/L, 2000 mg/L and 4000 mg/L concentrations respectively in comparison to control sample, but this decrease were low in 4000 mg/L concentration. Total weight, weight of root, stem and leaf and weight of root (Fig 5), dry weight of leaf, stem and root (Fig 6), wet weight of leaf, stem and root (Fig 7) and relative water content estimates of *Z. mays* treated by SiO₂ nanoparticles at four concentrations; 0, 400, 2000, and 4000 mg/L (Fig 8) respectively, is demonstrated.

The Effect of SiO₂ Nanoparticles on Photosynthetic Pigment

The effect of SiO₂ nanoparticles on photosynthetic pigment contents is shown in Fig 9. It evidenced the effect of SiO₂ nanoparticles on the contents of chlorophyll a, b and carotenoids. However, nanoparticles at 400 mg/l, 2000 and 4000 mg/L concentrations caused an increased content in all the photosynthetic pigments in *Z. mays* in relation to the control. This increase was comparatively more in 400 mg/l concentration.

CONCLUSION

The present experimental study was focused on the potential effect of engineered nanoparticles on *Z. mays* L. plant. The growth rates of the plant roots were enhanced as a result of exposure to SiO₂ nanoparticles (Fig 4). As far as seed germination is concerned, with increase in concentration of SiO₂ nanoparticles, the seed germination increased at 400 mg/L but lowered in 2000 and 4000 mg/L. Prior studies have provided similar reports where positive effects of NPs on germination and growth of plants have been authenticated. For example, TiO₂ and SiO₂ NPs were found to enhance not only the germination but also the growth of *Glycine max* seeds [13]. Similarly, carbon nanotubes (CNT) were learned to progress germination and root elongation of tomato seeds [14]. The emergence of such observations is possibly due to an enlarged water uptake by seeds in the presence of high concentrations of NPs [15]. Till yet nil studies were found describing the mechanism by which SiO₂ nanoparticles affect the uptake, translocation and growth of plants. In the present study, RWC in plant decreased by SiO₂ nanoparticles at 400 mg/L, 2000 mg/L and 4000 mg/L concentrations as compared to control sample. Low decrease in 4000 mg/L concentration can be probably due to unusual increases of root growth in this concentration. Until now no studies were explored explaining the effects of nanoparticles on plants. Plants can efficiently utilize the absorbing light energy when treated with SiO₂ nanoparticles and it is recommended that an increase in photosynthetic pigment is a natural response of plants. Nanoscale SiO₂ increased leaf chlorophyll a, b and carotenoid content in treated plants irrespective of concentrations as compared to control. SiO₂ nanoparticles at 400 mg/L recorded the highest chlorophyll a, b and carotenoid content (Fig 9). Higher chlorophyll accumulation is caused by matching effect of other intrinsic nutrients like magnesium, iron and sulfur. Similar results were experienced by Zhang *et al.*, [16] where *Spinacia oleracea* seeds were treated with TiO₂ nanoparticles. This is in concurrence with our results wherein in 2000 mg/L and 4000 mg/L concentrations of SiO₂ nanoparticles, the rate of increase in chlorophyll a, b and carotenoids content, reduced. It can be safely said that exposure to higher concentrations of nanoparticles may create some damages in plant photosynthetic apparatus. Till date, there are few studies reported on the phytotoxicity effects of nanoparticles on plants. The toxic effects considerably

depend on many factors viz. the type of nanoparticles, plant species, and the stage of plant development that should be surveyed in future studies. Therefore it can be safely said that yet, there are many gaps in our understanding of the toxicity of nanoparticles and there are several unresolved problems and new challenges as far as the biological effects of these nanoparticles is concerned.

Table 1 Detail of Nano SiO₂ used in Present Study (Stock #: 4860MR) (A and B).

Components	Contents (%)	Nano Material	Silicon Dioxide (SiO _x , x=1.2-1.6, amorphous)
Al	0.001	Purity	99.5%
Fe	0.001	APS	20 nm
Sr	0.004	SSA	160 m ² /g
Ca	0.002	Color	White
Mg	0.001	Morphology	Spherical
Cl	0.001	Bulk density	0.08 g/cm ³
Cr	0.004	True density	2.2-2.6 g/cm ³

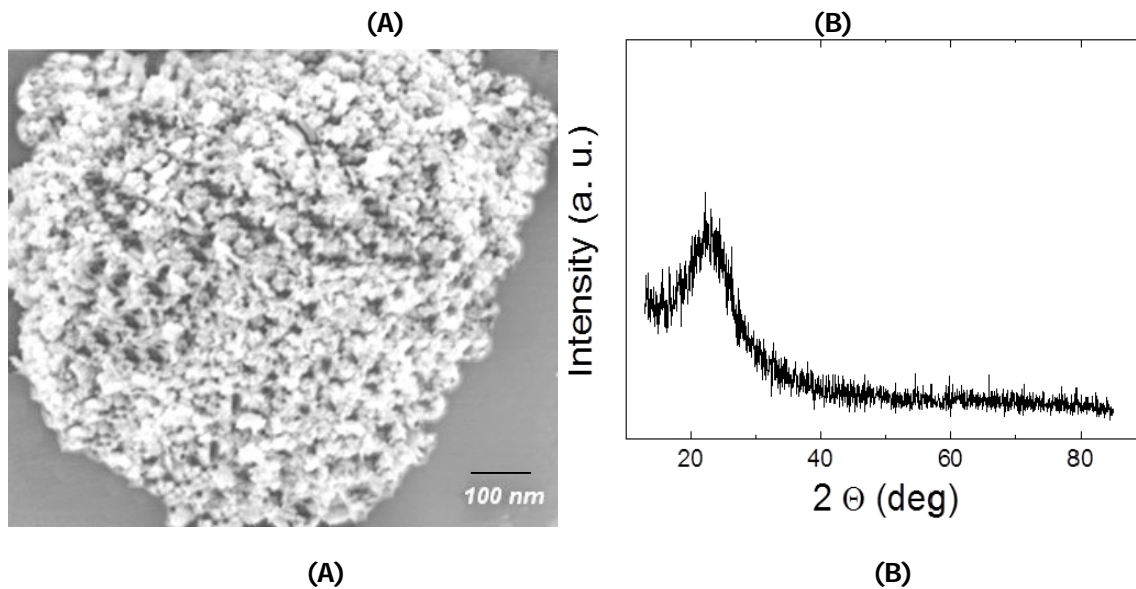


Fig. 1 Transmission Electron Microscopy (TEM) image of SiO₂ nanoparticles (A), X-Ray Diffraction Image of SiO₂ Nanoparticles (B).



Fig. 2 *Z.mays* after 21 days of planting. Right to left treated with 0 (control), 400, 2000, and 4000mg/L concentrations respectively.

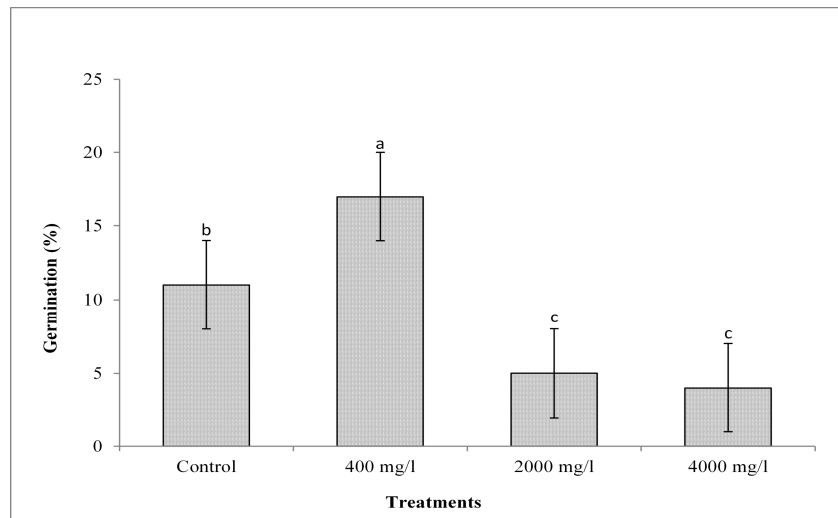


Fig. 3 Effects of varied concentrations of SiO₂ nanoparticles on seed germination. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

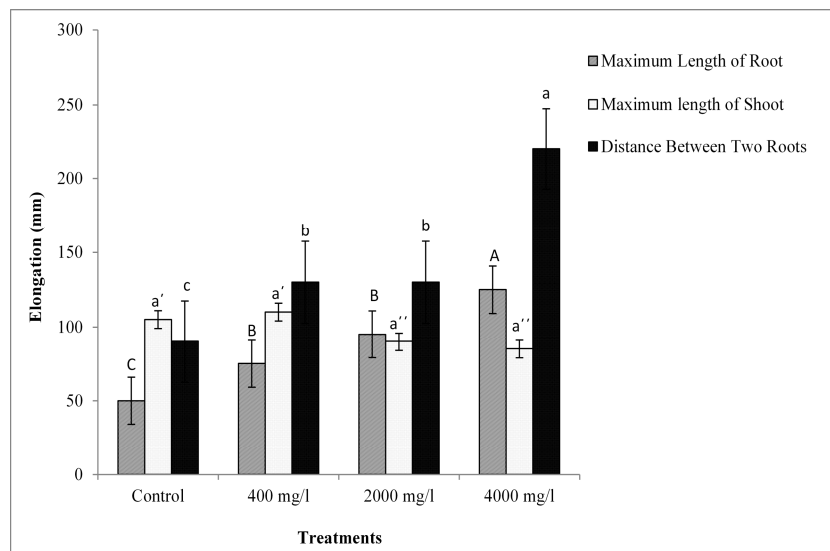


Fig. 4 Effects of varied concentrations of SiO₂ nanoparticles on root and shoot elongation. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

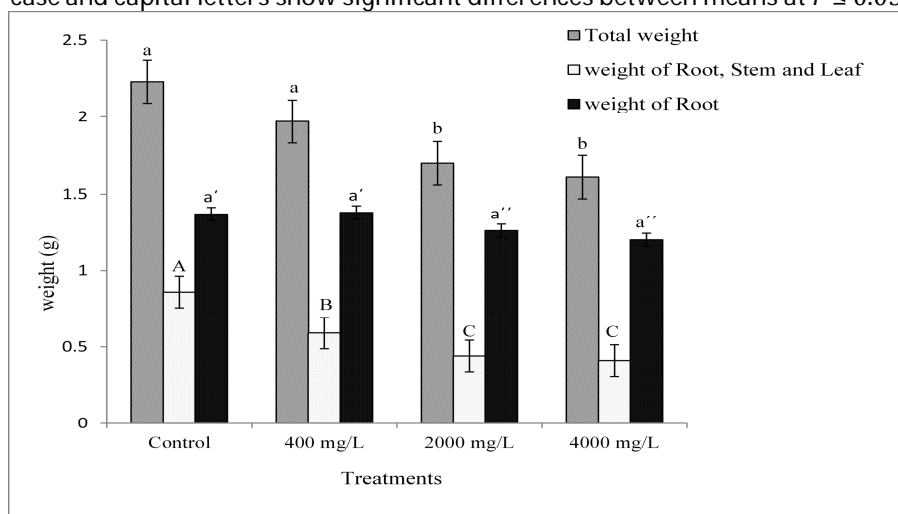


Fig. 5 Effects of varied concentrations of SiO₂ nanoparticles on total weight, weight of root, stem, leaf and weight of root. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

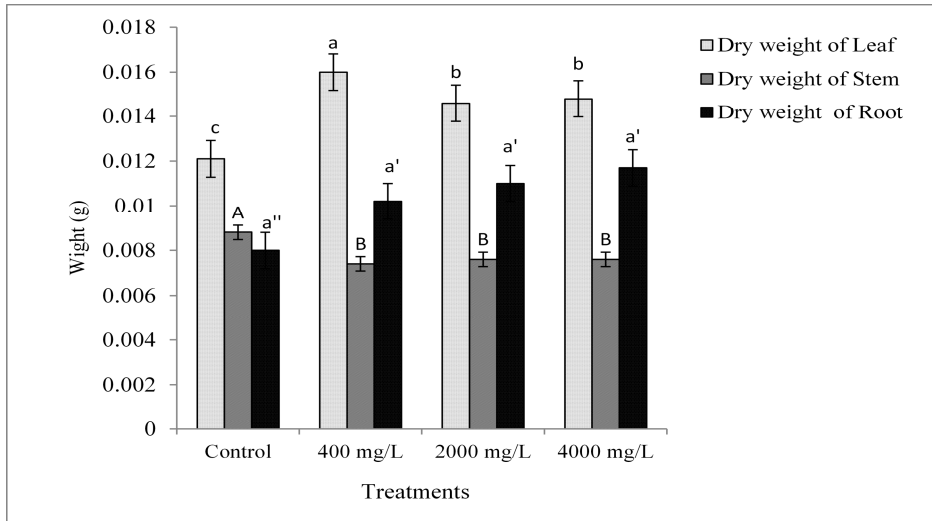


Fig. 6 Effects of varied concentrations of SiO₂ nanoparticles on dry weight of leaf, stem and root. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

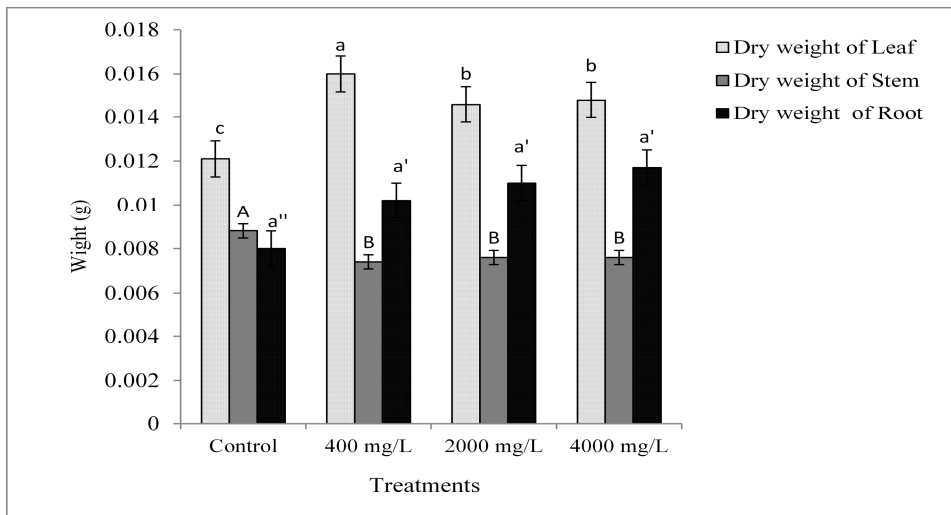


Fig. 6 Effects of varied concentrations of SiO₂ nanoparticles on dry weight of leaf, stem and root. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

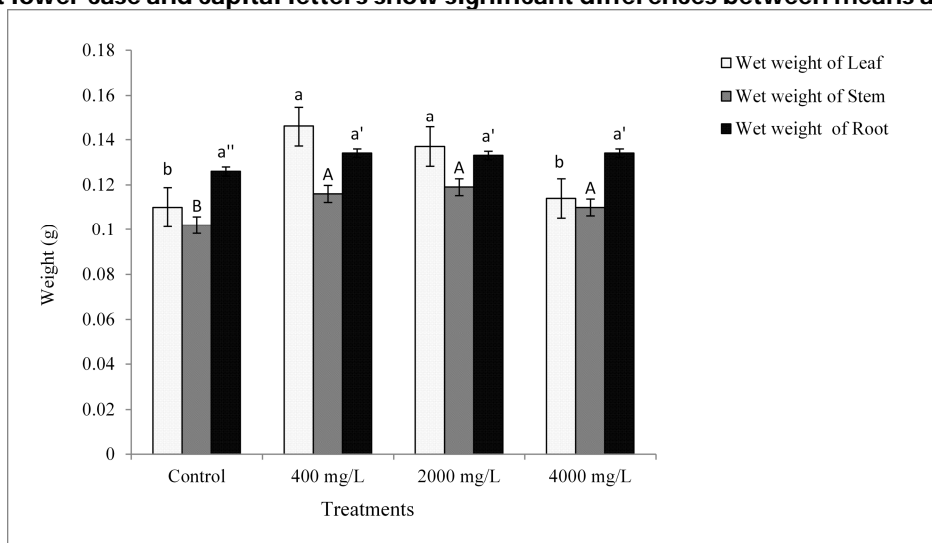


Fig. 7 Effects of varied concentrations of SiO₂ nanoparticles on wet weight of leaf, stem and root. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

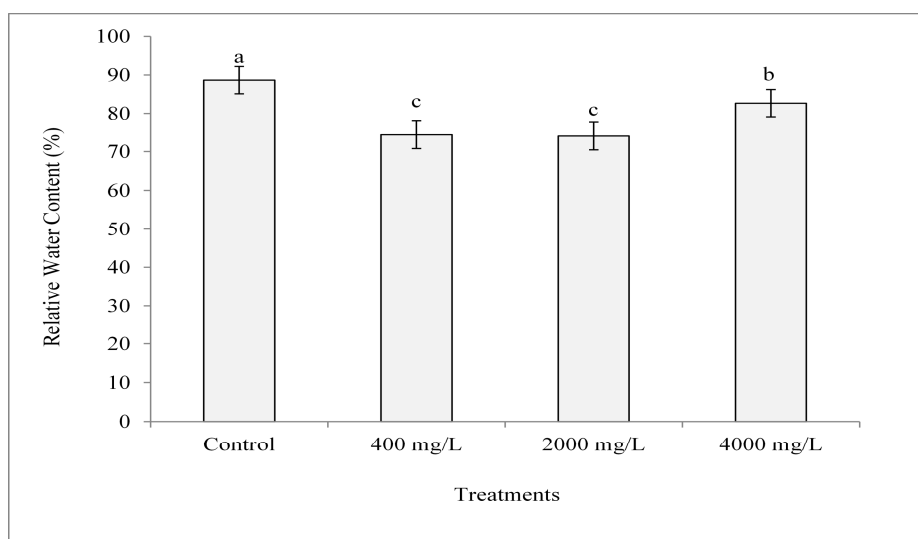


Fig. 8 Effects of varied concentrations of SiO₂ nanoparticles on relative water content. Different lower case and capital letters show significant differences between means at $P \leq 0.05$

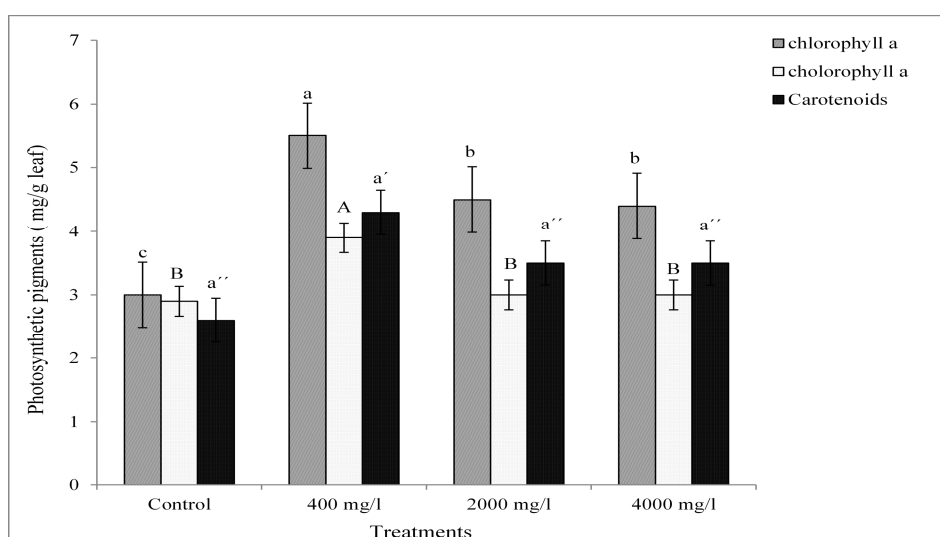


Fig. 9 Effects of varied concentrations of SiO₂ nanoparticles on photosynthetic pigments. Different lower case and capital letters show significant differences between means at $P \leq 0.05$.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

REFERENCES

- Daniel M C, Astruc D. (2004). Gold nanoparticles: assembly, supramolecular chemistry, quantum-size-related properties, and applications toward biology, catalysis, and nanotechnology. *Chemical Reviews*, **104**(1), 293–346.
- Berry C C, de la Fuente J M, Mullin M, Chu S W, Curtis A S. (2007). Nuclear localization of HIV-1 tat functionalized gold nanoparticles. *IEEE Transactions on Nano Bioscience*, **6**(4), 262-269.
- Tiwari P M, Vig K, Dennis V A, Singh S R. 2011. Functionalized Gold Nanoparticles and Their Biomedical Applications. *Nanomaterials*, **1**(1), 31-63.
- Tang F, Li L, Chen D. (2012). Mesoporous Silica Nanoparticles: Synthesis, Biocompatibility and Drug Delivery. *Advanced Materials*, **24**(12), 1504–1534.
- Ball P. 2002. Natural strategies for the molecular engineer. *Nanotechnology*, **13**(5), 15-28.
- Nel A, Xia T, Mädler L, Li N. 2006. Toxic potential of materials at the nano level. *Science*, **311**(5761), 622-627.
- Auffan M, Rose J, Wiesner M R, Bottero J Y. (2009). Chemical stability of metallic nanoparticles: A parameter controlling their potential cellular toxicity in vitro. *Environmental Pollution*, **157**(4), 1127-1133.
- Maynard A D, Aitken R J, Butz T, Colvin V, Donaldson K, Oberdörster G, Philbert M A, Ryan J, Seaton A, Stone V, Tinkle S S, Tran L, Walker N J, Warheit D B. (2006). Safe handling of nanotechnology. *Nature*, **444**, 267–269.
- Smart R E, Bingham G E. 1974. Rapid estimates of relative water content. *Plant Physiology*, **53**, 258-260.
- Haghighi M, Pourkhaloe A. (2013). Nanoparticles in agricultural soils: their risks and benefits for seed germination. *Minerva Biotechnologica*, **25**(2), 123-32.

11. Lichtenthaler H K , Wellburn A R. (1985). Determination of total carotenoids and chlorophylls a and b of leaf in different solvents. *Biochemical Society Transactions*, **11**, 591-592.
12. Abbaszadeh B, Ashourabadi E S, Lebaschi M H, Kandy M N H, Moghadami F. (2008). The Effect of Drought Stress on Proline Contents, Soluble Sugars, Chlorophyll and Relative Water Contents of Balm. *Iranian Journal Of Medicinal And Aromatic Plants*, **4**, 504-513.
13. Lu C M, Zhang C Y, Wen J Q, Wu G R, Tao M X .(2002). Research of the effect of nanometer materials on germination and growth enhancement of Glycine max and its mechanism. *Soybean Science*, **21**, 168-172.
14. Khodakovskaya M, Dervishi E, Mahmood M, Xu Y, Li Z, Watanabe F, Biris S. 2009. Carbon Nanotubes Are Able To Penetrate Plant Seed Coat and Dramatically Affect Seed Germination and Plant Growth. *ACS Nano*, **10(3)**, 3221-3227.
15. Nair R, Varghese S H, Nair B G, Maekawa T, Yoshida Y, Kumar DS .(2010). Nanoparticulate material delivery to plants. *Plant Science*, **179(3)**, 154-163.
16. Zhang L, Hong F, Lu S, Liu C. (2005). Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biological Trace Element Research*, **104(1)**, 83-91.