**Bulletin of Environment, Pharmacology and Life Sciences** Bull. Env. Pharmacol. Life Sci., Vol 4 [8] July 2015: 16-25 ©2015 Academy for Environment and Life Sciences, India Online ISSN 2277-1808 Journal's URL:http://www.bepls.com CODEN: BEPLAD Global Impact Factor 0.533 Universal Impact Factor 0.9804

**ORIGINAL ARTICLE** 



**OPEN ACCESS** 

# Examining the Effect of the Aqueous Extract of Anchusa italica on abortion and Fetal-placental parameters in Wistar Female Mice

Arezoo Zangeneh<sup>1</sup>, Shahla Roozbahani<sup>2</sup>

1Department of Biology, Islamic Azad University Hamedan, Hamadan, Iran 2- Department of Biology, Islamic Azad University Flavarjan, Esfahan. Iran

### ABSTRACT

The borage plant with scientific name of Anchusa italica belongs to the Boraginaceaae family. These plants contain fatty acids, seed oils, and chemical compounds. Any uterine bleeding associated with closed uterine cervix and bloody discharge in the first half of pregnancy is called a threatened to miscarriage, and in 50% of cases results in abortion. The placenta is the main organ for maintaining healthy pregnancy and has an important role in transferring the foodstuffs from mother to fetus. Fetus weight gaining during pregnancy is a response to placenta growing that is considered as an independent factor in predicting the neonatal weight. The aim of this study is to evaluate the effect of aqueous extract of Anchusa italica on miscarriage and related parameters and Fetal-placental parameters. In this experimental study, after mating and observation of vaginal plug, 30 Wistar female mice were randomly divided into five groups including one group receiving physiological saline intraperitoneally, and four treatment groups receiving aqueous extract of Anchusa italica at concentrations of 100, 200, 300, and 400 mg/kg. Injection was daily conducted for 5 days after the 7<sup>th</sup> day of pregnancy, and cesarean operation was performed on 16th day of pregnancy. The number of absorbed fetus were counted and considered as abortion also Fetal-placental parameters were evaluated and the results were analyzed using SPSS software. The results of this study suggest that Injecting Anchusa italica extract affected the number of fetus absorbed(p=0.016), The number of fetus formed(p=0.000), The number of fetus observed(p=0.000), Fetus weight(p=0.000), placental weight(p=0.016), Placental size(p=0.002), The number of placental(p=0.000).\_Has been effective because the standard deviation is less than 0.05. Therefore, the differences shown in the figure were statistically significant. The results suggest that the aqueous extract of Anchusa italica affected the number of fetus absorbed, the number of fetus formed, and the number of healthy fetus observed and Fetal-placental parameters. Keywords: Anchusa italica, fetus weight, Placental size

Received 12.05.2015

Revised 21.06. 2015

Accepted 05.07.2015

# **INTRODUCTION**

The new studies show that herbal plants can influence on all body organs and systems including the productive system [1]. Extracts of several plants can manifest its effect during the pregnancy or during delivery which indicates the importance of mothers' diet during pregnancy [2]. The borage plant with scientific name of Anchusa italica belongs to the Boraginaceaae family. This plant has contain fatty acids (Linoleic, oleic,  $\alpha$ - Linoleic, y- Linoleic, Palmitic, Stearic, Eicosnoic, Erucic and Steroosonic) and seed oils [3]. Raw polysaccharides of *Anchusa italica* roots include poly (oxy-1-carboxy-2-3,4-dihydroxy-phenyl ethylene) [4]. The plant petals also contain flavonoids, saponin and phenolic compounds [5]. Endulatocid, Tri-terpenes glycoside and Flavonoid glycoside were first separated from the Anchusa plant.(6). Phytosterols are found in large quantities in plant oils such as Anchusa italic. In addition, beta-Sitosterol and Campesterol were identified as the major sterols [7]. After fertilization, the womb becomes a place for keeping cell mass which will be transferred to the fetus. If no disorder occurs in it, it leads to childbirth after passing the pregnancy dumouseion with the uterine contractions, while if there occurs inconsistency in this part of body due to disordering effects, it will be influenced by these factors regarding the pregnancy step, there occur adverse effects on fetus that sometimes lead to abortion [9] The underlying mechanisms of miscarriage include fetus - maternal and environmental [10]. Numerous studies conducted in different countries have obtained different results. Mental problems (depression, anxiety, suicide) are likely to occur in women who experience miscarriage increase (11). The placenta is the main organ for maintaining a healthy pregnancy and has an important role in transferring the foodstuffs from mother to fetus [11]. As the fetus grows in the womb, main changes occur in shape and

function of the placenta to be a proper solution and good response to fetus needs during growth stages. Reaching such consistency requires unique metabolic, immunologic and endocrine changes in placenta trophoblasts [11, 12]. Fetus weight gaining during pregnancy is a response to placenta growing that is considered as an independent factor in predicting the neonatal weight [13]. Infant underweight is due to reducing the receiving function of placenta such that it reduces by placenta of exchanging area between mother and fetus means the velocity placental level and capillary level of fetus reduce therefore the ability of transferring oxygen and food from mother to fetus reduces (11). As it is expected, the capacity of weight growth is mostly determined by fetus and fetus growth belongs to placental weight [14].Meanwhile, placenta as an important auxiliary and additive source contributes the estrogen production and if pregnancy is natural, its production will be increased since 10<sup>th</sup> pregnancy week by its end especially in 5 last weeks [15]. Such hyper-estrogen is one of the progressive criteria of pregnancy which is removed immediately after placenta and fetus removal [16].During delivery, the plasma level of progesterone increases from 1-2 ng/m to above 100 ng/m. During the week 6-36<sup>th</sup> weeks of pregnancy the level of 17-alpha-hydroxy of plasma progesterone increases from 0.5 ng/m to 50-60 ng/m, and in the 32<sup>nd</sup> week of pregnancy, its plasma levels gradually increases from 0.5 ng/m to 140-120 ng/m. [17].The metabolic clearance of progesterone in pregnant women is similar to that of non-pregnant women and men. During pregnancy, a disproportionate increase occurs in the plasma concentration of  $\alpha$ -5 d-hydroxyprogesterone, and the concentration of this metabolite to progesterone concentration reduces in pregnant women. The mechanism is still unknown; perhaps it is in relation to the development of resistance against shrinkage factors that occur during a normal pregnancy. In pregnant women and the fetus, the progesterone converts into the strong mineral corticosteroid dexa corticosterone (DOC) [18].Termination of pregnancy, whether spontaneous or induced, before the fetus is developed enough to evolve is called abortion [19]. A genetic factors influencing in miscarriage is the progesterone receptor modulator UPA as a contraceptives factor in women [20]. [21] concluded that Runx3 (Runt3) plays an important role during implantation and decidualization. Press35 gene is also involved in capital letter development, ovulation, implantation, and decidualization [22]. IGFBP7 inhibition through specific immunization of DNA significantly reduces the pregnancy rate and implantation of fetus [23]. [24] study that RU-486 is an anti-progesterone factor that can cause premature labor.Fatty acid amide hydrolase (FAAH) is effective in the development and implantation of the embryo [25]. Wnt7a has an important role in the formation and function of the uterus gland required for implantation of the blastocyst and reproductivity in adult uterine [26]. The results of studies by [27]. showed that the lack of N and O sets of Glycon in growing oocytes damages the primary stages of fetus growth and leads to delaying in implantation and a small infant. During the pre-implantation stage, the uterine growth influences on the process of implantation and fetus growth. Uterine unnatural growth causes many productive diseases in human. Among adjusting factors of uterine growth, vascular remodeling promoters are important for uterine function and fertilization. Meanwhile VEGF vascular endothelial growth factor is important in endothelial cell growth and development of blood vessels and nonendothelial cells [28]. IGFBP7 is mainly in Granular epithelium and platform and has more exposition after fetus implantation. IGFBP7-t, PCR3/1 is made by exposing IGFBP7. IGFBP7 inhibition via special DNA immunization considerably reduces the pregnancy and implanting fetuses [29]. The methanol extract of Iranian Anchusa italica is effective in seizure induced by picrotoxin (30). Anchusa italica is harmful for pregnant women due to alkaloid compounds [31]. Anchusa italica liver toxicity effects is generally accepted for its alkaloid compounds [32]. Anchusa italica and fish oil causes inflammation and bone repair [33]. Anchusa italica extract has higher liver toxic effects compared to valerian extract [34]. Anchusa italica has obvious effect of increasing blood pressure, especially in high blood pressures, and using dried limes brewed mixed with Anchusa italica is recommended to reduce heart beat rate [35]. Aqueous extract of Anchusa italica is an effective and safe drug for the treatment of patients with obsessive-compulsive disorder [36]. Methanolic extract of Anchusa italica shows anti-inflammatory effects [37]. Anti-oxidizing, anesthetic, immune modulating and anxiolytic anxioletic of (Iranian) Anchusa italica show that this plant has a beneficial effect in acute pancreatitis, and has anti-inflammatory, antioxidant and immune modulating effects [38]. Indian Anchusa italica essence oil is a natural resource to control African malaria-carrying mosquito [39]. The antiviral activity of flu was evaluated in aqueous and alcoholic extracts of the Anchusa italica plant. These findings suggest that the antiviral activity of the plant is likely due to interference with the replication and transcription of the virus [40]. The Anchusa italica effects on pregnancy outcome in this study was to evaluate the effects of the extract on abortion, placenta and fetus.

## MATERIALS AND PROCEDURE

Preparing Anchusa italica:

*Anchusa italica* was prepared from semirom and them systematic diagnosis was done in herbarium plant, and their flower and shoot were dried and then the dried plant was grinded and for preparing the extract, the powder was soaked in distilled water in vitro for 24 hours and then having used shaker for 48 hours, the pure extract was obtained, this extract was prepared in concentmouseion of 100, 200, 300 and 400 mg/lit and then extracts were ready for injection.



Fig1 : Experimental Plant and Extraction

Treatment animals:

In this research, 30 wistar female mice and 20 wistar male mice in mean weight of 25 gram to 30 gram were bought from Islamic Azad University, Shiraz branch. Then the female and males mice were put sepamouseely in special cages and they were kept for 7 days for compatibility with labomouseory conditions in 27°C and natural light. Food and water were available for animals unlimitedly. Meanwhile the mice were pathologically studied daily. After 7 days, the male and female mice were put next to each other randomly for intercrossing and having controlled the vaginal plaque in female mice, the mice whose vaginal plaque were observed were sepamou seed from the remaining mice in sepamousee cages. G0 at  $1^{st}$  day of pregnancy was written for them, then pregnant female mouses were randomly divided into 5 groups, these groups included control group, group receiving 100 concentmouseion, 200 concentmouseion, 300 concentmouseion and 400 concentmouseion. After 7st pregnancy day, Anchusa *italica* was Intraperitoneally injected to the mice in treatment group for 5 pregnancy days and physiology serum was intraperitoneally injected to mice in control group and the caesarean was done on 16<sup>th</sup> day of pregnancy. For doing this openouseion, at first the animal was anesthetized by chloroform and having establishing transverse cleft on abdomen and removing Retroperitoneal, the uterine horns containing fetus are observed. Having used narrow edge of surgical scissors, the wall of uterine horn was opened precisely and fetus was got out of it. Thus, healthy fetus were counted. Then, in order to determine the number of fetuses aborted, by examining the lining of the uterine horn was examined and the number of placenta without fetus and the number of born prematurely babies were added were considered as the number of aborted fetuses. its pill-shape and easily get out if wall of uterine horn. For measuring placental weight, they were put on digital scale after getting out the placenta in an animal and weight was registered in each animal and finally mean of placental weight is obtained in a treatment group. For measuring placental size, its diameter is determined by caliper and/or ruler and then total mean was determined in each treatment and finally the results were analyzed by SPSS18.

# RESULTS

Effect of *Anchusa italica* on The number of fetus absorbed, The number of fetus formed, The number of fetus observed:

Injecting *Anchusa italica* extract affected the number of fetus absorbed, since the standard deviation is less than 0.05 (p =0.016). Therefore, the differences shown in the figure are statistically significant. Injecting *Anchusa italica* extract affected the number of fetus formed, since the standard deviation is less than 0.05 (p =0.000). Therefore, the differences shown in the figure are statistically significant. Injecting *Anchusa italica* extract affected the number of healthy fetus observed, since the standard deviation is less than 0.05 (p =0.000). Therefore, the differences shown in the figure are statistically significant (Table 1).

Variable	Changes	Sum of squares	Degree of freedom	Mean square	Fischer Z	p_value
The number of fetus absorbed	Between Groups	36.795	4	9.199	3.889	0.016*
The number of fetus formed	Between Groups	136.449	4	34.112	8.125	0/000*
The number of fetus observed	Between Groups	224.705	4	56.176	12.440	0/000*

Zangeneh and Roozbahani



Diagram 1. Comparison of the average number of fetus formed in different treatments of plant



extract levels of anchusa italica

Diagram2. The correlation between the number of fetus absorbed in different treatments of plant



Diagram 3. Comparison of the average number of fetus absorbed in different treatments of plant

Effect of *Anchusa italica* on, Fetus weight, placental weight, Placental size, The number of placental: The injection of *Anchusa italica* is effective on fetus weight since standard deviation is less than 0.05 (P= 0.000) and the differences in graph are statistically significant. The injection of *Anchusa italica* is effective on placental weight since standard deviation is less than 0.05 (P= 0.016) and the differences in graph are statistically significant. The injective on placental size since standard deviation of *Anchusa italica* is effective on placental size since standard deviation is less than 0.05 (P= 0.016) and the differences in graph are statistically significant. The injection of *Anchusa italica* is effective on placental size since standard deviation is less than 0.05 (P= 0.002) and the differences in graph are statistically significant. The

injection of *Anchusa italica* is effective on number of placenta since standard deviation is less than 0.05 (P= 0.000) and the differences in graph are statistically significant. (Table 2).

Variable	Changes	Sum of squares	Degree of freedom	Mean square	Fischer Z	p_value
Fetus weight	Between Groups	0.757	4	-1.189	10.298	0/000*
placental weight	Between Groups	0.506	4	0.126	3.899	0.016*
Placental size	Between Groups	121.643	4	30.411	6.243	0.002*
The number of placental	Between Groups	136.449	4	34.112	8.125	0/000*



extract levels of anchusa italica

Graph 1: comparing mean of fetus weight in different treatments of plant









Graph 4: comparing mean of number of placenta in different treatments of plant

# **RESULTS AND DISCUSSION**

Effect of *Anchusa italica* on The number of fetus absorbed:

The results of the study showed that the number of embryos absorbed in the 100 and 200 control is a significant difference. In contrast, the number of embryos absorbed in level control significant difference was observed in the control group (Table 1).

[41] introduced Anethum graveolens as a contraceptive and a factor to regulate the menstrual cycle .Studies by (42) showed that *Stachys lavandulifolia* perforatum extract has properties that cause miscarriage and it should be used cautiously during pregnancy .In a study on the aqueous extract of pjyllanthus amarus (AEPA), results showed that the AEPA resulted in miscarriage in pregnant mice [43]. MSX homeobox genes regulate embryonic implantation through paracrine signaling between the substrate and epithelium type 1 and type 2, which encodes the transcription factors and control organogenesis and tissue interactions during the growth of fetus. Lack of these genes increases Wnt signaling in substrate cells and enables  $\beta$ -catenin that causes production of fibroblast growth factors (FGFs) in these cells. The secreted FGFs act in paracrine method through FGF receptors in epithelium to increase proliferation of epithelial cells. These findings indicate a unique signaling network, which includes Msx 1/2, Wnts, and FGFs, and at the time of implantation, work to control the mesenchyme epithelial relationship [44]. [45] showed that exposure Using genistein leads to major changes in gene expression of fallopian tubes, Other changes in gene expression of fallopian tubes were observed only in early pregnancy. After the FRT is exposed to inflammatory or antigenic stimuli resulted from ovulation and mating, changes in fallopian tubes affect the growth of developing fetus through increased branching and a reduced the cell ratio of trophectoderm cellular mass in blastocyst stage. The effects of Bisphenol A (BPA) on uterus and embryo factors in implantation shows negative effects of high doses of BPA on the processes that are critical for embryo implantation. These processes include fetus transportation, fetus development before implantation, and stabilization of the uterus receptivity [46]. [47] Yellow Daphne consumption in early pregnancy showed that it can cause destruction and necrosis in decidual tissues and release prostaglandins, while increasing the levels of alpha 2PGF and 2PGE and decreasing progesterone level. On the other hand, the plasma levels of alpha PGF2 and PGE2 remained unchanged, and estrogen and progesterone levels dropped. In general, the contraceptive mechanism of Yellow Daphne mainly intervenes in intrauterine environment to maintain pregnancy, which in turn lead to abortion [1].In contrast, studies by [48] suggest that using ethanolic extract of eremomastax speciosa leaves has toxic effects on the endocrine system of female mice, and it is not dangerous as a drug in the management of infertility in female mice.

Flavonoids are polyphenolic compounds. Flavonoids have strong antioxidant effects and inhibit free radicals and reduce the risk of some chronic diseases [49, 50]. Luteolin, and kaempferol phytophenols apply anti-inflammatory properties in the pregnancy tissues. Anti-inflammatory properties in the pregnancy tissues are shown by inhibiting activity of binding to DNA NF-Kb, the AP1 path, and its target genes. In addition to anti-inflammatory activity, phytophenols have a wide range of biological activities. Kaempferol increases the length of pregnancy. Luteolin and kaempferol does not affect secretion of cytokines on the pregnant myometrial cells. In myometrium cells, luteolin, and kaempferol have substantially reduced incidence of COX-2, prostaglandin release, and MMP-9 activity resulted from the

B1-L1. The two substances perform their activity through AP-1 and NF-Kb paths, and prevent binding of DNA 65 NF-kBp and c-jun in myometrial cells [51].TNF- $\alpha$  and B1-L1 apply inflammatory activity, such as elevating prostaglandin and ECM destructive enzymes. This results in the beginning of three important stages in human delivery: fetus membranes rupture, cervix maturation, and uterine contractions [52]. The COX2 produces prostaglandins that are important in the initiation and maintenance of labor through increased uterine contractions and cervical maturation and activation of fetus-decidual membrane (52, 53).Linoleic acid is an essential fatty acid for the formation of cholesterol sterols through ACAT enzyme in the membrane of the yolk sac. This acid has a negative impact on the absorption of lipids in chicken fetus at the last embryonic stage that leads to increased fetus mortality [54]. Studies have shown that saponins have a contraceptive effect with a negative feedback mechanism. The effects of this extract occur before and after implantation. The anti-implantation activity is probably due to estrogenic activity that excretes the ovule from the fallopian tube and disrupts the luteotrophic activity of blastocyst [55]. Recent studies have shown that long-term use of phytosterols can increase reproductive success in short-tailed mice [56], and stimulate egg production in hermaphrodite marshy snails. However, despite the increased quantity, the quality of eggs will be reduced [57].

Effect of *Anchusa italica* on, Fetus weight, placental weight, Placental size, The number of placental:

The results of this study indicate that 100 with 200 pairs showed a significant difference in weight (p=0.000). In contrast, the control group showed no significant difference with other levels.

The results of this study suggest that placental size is a significant difference in the level of control with the 100 and 200 levels (p=0.11). As well as the 300 there is a significant difference(p=0.45). The size of the pair in the 100 to 200 showed a significant difference (p=0.000). In contrast, other levels did not show significant difference.

The results indicate that the number of pairs in the control group with a significant difference in the levels of 100 and 200(p=0.000). As well as the 300 there is a significant difference (p=0.009). In contrast, the number of pairs in other levels did not show significant differences.

Zingiber officinale causes wide disorders and anomalies in fetus maybe due to its effect of smooth muscles and also due to its effect on synthesis of leukotriene and prostaglandins [58]. Tmouseogenic effects of Stachys lavandulifolia on skeleton system and fetus growth of mouse showed that Stachys ivandulifolia at different doses causes disorder in fetus growth (height and weight) and such disorder upon increasing the dose increases. Meanwhile this extract has tmouseogenic characteristics and reduces growth and reduces interparital ossification belonged to dose [59]. The findings of a research showed that both Anchusa italica and matricarica chamomile causes disorder in natural process of pregnant mousee weight gaining and it reduces the length of rumps-seats and their fetus weight [60]. The consumption of pigeon pea extract as edible increases the infant size of mouse while the infants' weight reduces. Meanwhile, this extract reduces the mother's weight gaining [61].[62] showed that *Peganum hermla* influences of skeleton system and fetus growth of mouse and reduces the fetus height and weight. The consumption of Zataria multiflora in second week of pregnancy causes partial changes like tail size and placental diameter [63].The studies about the effect of Physalis alkekengion anomalies showed that this extract reduces the diameter, weight and size of placenta, meanwhile, Physalis alkekengi may have negative effect on quantitative growth of fetus and distinction of some cells through preventing the cellular division and distinction and reducing the function of several enzymes and hormones (64). On the other hand, the results of studies of [65] showed that Green tea catechins (GTC-H) has no effect on mean of uterine weight of the uterus growth or living. Palmitic acid is a satumouseed fatty acid and this acid has distinctive effects on fetus trofektrom and internal cell mass and influence on placenta growth (66). Palmitic contact in fetus after implantation will include changing R1IGF and increasing GPT2 activity. Such metabolic changes with cellular nucleon are in fetuses which influenced by palmitic acid which is probably the result of increasing epoptosis and reducing cellular prolifemouseion in fetus and TS cells are exposed to increasing the palmitic concentmouseion. Meanwhile, metabolic changes can influence on long-term consequences merely via influencing on cellular prolifemouseion before implantation and epoptosis via changing the availability of energy. The other probability is that these metabolic changes indicates changes in genetic immune adjustment of metabolism or abnormal planning of Mitochondrial function organisms. The fetus shows the change of growth and metabolic patterns connected to extra palmitic acid before implantation and these fetus shows growth limitation in uterine (67, 68). Several long fatty acid supplements of Omega 3 chains increases the pregnancy dumouseion in humans and animals [69].Deco Hexanoic acid (DHA) with CLA causes delay in delivery time in induced lambing by beta-methasone and fish oil in human can increase the pregnancy dumouseion by 4 days (70, 69). Increasing pregnancy dumouseion maybe related to the inhibitory effect of CLA on Prostaglandins production. the recent researches knows the inhibitory effect of CLA in prostaglandin as decisive and related to PPAR activation since PPAR inhibits the activity of nuclear polygraph factor (NF $\kappa$ B) after

activation [71]. Several researchers believe that the fatty acids in diets influence on homeostasis adjusting hormone of body weight and energy balance (liptin) (72, 73).

Therefore regarding the results obtaining, *Anchusa italica* shall be used with caution.

## RREFERENCES

- 1. Zou, A.M., Fang, J.Y.& Ye, D.(1993). Effect of yellow daphne on levels of plasma and decidual estradiol, prostaglandins, progesterone and its receptor in early pregnancy. Zhongguo. Zhong. Xi. Yi. Jie. He. Za. Zhi.,9(531-2):516.
- Magee, L., Vohra, S., Matsui, D., Bérard, A., Johnson, B., Moretti, M., & Einarson A.(2008).MotherNature: Establishing a Canadian Research Network for Natural Health Products (NHPs) During Pregnancy and Lactation. *J. Altern. Complement. Med*; 14 (4): 369 – 72.
- 3. Tamer, O.(2008). Fatty Acid Profiles of the Seed Oils in Two Groups of Anchusa officinalis L. IUFS. J. Biol.,(1):65.
- Barbakadze ,V., Gogilashvili, L., Amiranashvili, L., Merlani, M., Mulkijanyan, K., Churadze, M., Salgado, A. & Chankvetadze, B.(2010), Poly[3-(3,4-dihydroxyphenyl)glyceric acid] from *Anchusa italica* roots. Nat. Prod .Commun., 5(7): 1091.
- 5. Naderi Haji Bagher Kandi, M.& Rezaei, M.B.(2004). The phytochemical study of Echium amoenum. Iranian Medicinal and Aromatic Plants Research Quarterly; 20(3): 377-383.
- 6. Koz, O., Pizza, C. & Kirmizigül S.(2009), Triterpene and flavone glycosides from Anchors undulata subsp. Hybrid. Nat Prod Res., 23(3):284.
- 7. Abbaszadeh, S., Rajabian, T. & Taghizadeh, M.(2012). Identification of phytosterols in the oil seed of populations of two species of Echium in Iran. Iranian Medicinal and Aromatic Plants Research Quarterly., 28 (4): 741-742.
- 8. Guyton, A.C. & Hall, J.E. (2006). Textbook of Medical Physiology. 11th ed. Chapter 82, Elsevier-Saunders, p. 1027 41
- 9. Cunningham, F.G. Leveno, K.J. Bloom, S.L. Hauth, J.C. Gilstrap III, L.C. & Wenstrom, K. D.( 2005).New York, McGraw-Hill, p.232.
- 10. Fergusson, D.M., Horwood, L.J.& Ridder E.M.(2006). Abortion in young women and subsequent mental health. J Child Psychol Psychiatry.,47(1):16.
- 11. Yu, Km.(1992). Relation between placental morphometry and fetal growth. zhonghue .fu. Chan. Ke. za .zhi .,27(4):187-9.
- 12. Chunnigham, F.C, et al. (2001). Williams Obsteric. 21 st ed. Philadelphia : Mc Graw HILL,: 558-9, 572, 745, 758.
- 13. Kinare, A.S., Natekar, A.S., Chinchwadkar, M.C., Yajnik, C.S., Coyaji, K.J., Fall, C.H.& Howe, D.T. (2000). Low midpregnancy placental volume in Rural Indian women: A cause for low birth weight?. Am. J. Obstet. Gynecol., 182(2):443-8.
- 14. Lo, Y.F.Jeng, M.J., Lee, Y.S., Soong, W.J. &Hwang ,B.(2002). Placental Weight and Birth Charactristics of healthy Singlton Newborns. Acta. Paediatr. Taiwan ., 43(1):21-5.
- 15. Sanadgol, H. (1992). Human Physiology, Yazd publication, Volume 3, p.1627-1577-1576-1573.
- 16. Voightrich, W.J. (1997). Pregnancy and childbirth, Eshtiagh publication, Volume 1,p.179-104-103-92.
- 17. Hajihosseini, R.(2008). Hormones biochemistry, Payam-e-Noor University, p.131-130-59-58
- 18. Voightrich W.J. (2001). Pregnancy and childbirth, Golban publication, Volume 2.
- 19. Berek & Novak gynecology. (2007). Philadelphia, Lippincott, Williams & Wilkins.
- 20. Nallasamy, S., Li, Q., Bagchi, M.K.& Bagchi I.C.(2012). Msx homeobox genes critically regulate embryo implantation by controlling paracrine signaling between uterine stroma and epithelium. PLoS. Genet., 8(2):1.
- 21. Bai, Z.K., Guo, B., Tian, X.C., Li, D.D., Wang, S.T., Cao, H., Wang, Q.Y.& Yue, Z.P.(2013). Expression and regulation of Runx3 in mouse uterus during the peri-implantation period. J. Mol. Histol., 44(5):519-26.
- 22. Honglu, D., Shuo, X., Rong, L.i., Fei, Z. & Xiaoqin, Y.m.(2013). Distinct Spatiotemporal Expression of Serine Proteases Prss23 and Prss35 in Periimplantation Mouse Uterus and Dispensable Function of Prss35 in Fertility. PLoS One., 8(2):1.
- Zhen-Kun, L., Rong-Chun, W., Bing-Chen, H., Ying, Y. &Jing-Pian, P.(2012). A Novel Role of IGFBP7 in Mouse Uterus: Regulating Uterine Receptivity through Th1/Th2 Lymphocyte Balance and Decidualization. PLoS One., 7( 9):1.
- 24. Li, Y., Je, H.D., Malek, S. & Morgan, K.G.(2004). Role of ERK1/2 in uterine contractility and preterm labor in rats. Am. J. Physiol. Regul. Integr. Comp. Physiol., 287(2):328.
- 25. Xiao, A.Z., Zhao, Y.G. & Duan, E.K.(2002). Expression and regulation of the fatty acid amide hydrolase gene in the rat uterus during the estrous cycle and peri-implantation period. Mol. Hum. Reprod., 8(7):651.
- Dunlap, Kathrin, A., Filant, Justyna., Hayashi, Kanako., Rucker III, Edmund. B., Song, Gwonhwa., Min Deng, Jian., Behringer, Richard, R., Franco, J. DeMayo., Lydon, John., Jeong, Jae-Wook. & Spencer, Thomas. E.(2011). Postnatal Deletion of *Wnt7a* Inhibits Uterine Gland Morphogenesis and Compromises Adult Fertility in Mice. Biology of Reproduction., 85(2): 386.
- 27. Grasa, Patricia, Kaune, Heidy. & Williams, Suzannah. A.(2012). Embryos generated from oocytes lacking complex *N* and *O*-glycans have compromised development and implantation. Endocrinology., 144(4):455.
- Yan, Ji, Xiaodan, Lu., Qingping, Zhong., Peng, Liu., Yao, An., Yuntao, Zhang., Shujie, Zhang., Ruirui, Jia., Isaias, G., Tesfamariam, Abraha G., Kahsay, Luqing ., mail, Zhang., Zhu mail, Wensheng. & Zheng mail, Yaowu. (2013).Transcriptional Profiling of Mouse Uterus at Pre-Implantation Stage under VEGF Repression. PLoS One., 8(2):1.

- 29. Zhen-Kun, Liu., Rong-Chun, Wang., Bing-Chen, Han., Ying, Yang., & Jing-Pian, Peng. (2012). A Novel Role of IGFBP7 in Mouse Uterus: Regulating Uterine Receptivity through Th1/Th2 Lymphocyte Balance and Decidualization. PLoS One., 7(9):1.
- 30. Hosseini, A. &Shariatifar, N.(2004). The anticonvulsant effect of methanol extract of Iranian *Anchusa italica* on seizure caused by picrotoxin in mice, Journal of Gonabad Medical Sciences and Health University., 10(2): 20.
- 31. Smith, P., Konstantin, K., Hansell, R. & Chandler, F.(2004). The side effects of herbal medicines, translated by Tafaghodi, M., Mashhad University of Medical Sciences, Vol. 3, p.257.
- 32. Zamansoltani, Farzaneh., Nassiri Asl, Marjan., Karimi, Rana. & Mamaghani-Rad, Parvaneh. (2008). hepatotoxicity effects of aqueous extract of echium amoenum in rats. Pharmacologyonline, 1:432-438
- 33. Wauquier, F., Barquissau, V., Léotoing, L., Davicco, M.J., Lebecque, P., Mercier, S., Philippe, C., Miot-Noirault, E., Chardigny, J.M., Morio, B., Wittrant, Y. & Coxam V.(2012). Borage and fish oils lifelong supplementation decreases inflammation and improves bone health in a murine model of senile osteoporosis. Elsevier., 50( 2):553.
- 34. Zahedi, M.J., Heydari, M.R., & Mohajeri, M.(2003). The impact of valerian and *Anchusa italica* on kidney and liver function tests in rats. Journal of Kerman University of Medical Sciences., , 11(1): 22.
- 35. Hamidi Ravari, M., Khaksari, M. & Hojabri, K.H.(2011). The effects of aqueous extracts of *Anchusa italica* and dried limes on blood pressure and heart beat rate before and after injection of phenylephrine in male rats. Kerman University of Medical Sciences., 18(4):349-57.
- 36. Sayyah Bargard, M., Boustani, H., Sayyah, M., Fazileh, F., Kamalinejad, M. & Akhundzadeh, Sh.(2005) The effectiveness of the aqueous extract of chium amoenum in the treatment of obsessive-compulsive disorder, Journal of Medicinal Plants., 4(15): 43.
- Kuruuzum-Uz, A., Suleyman, H., Cadirci, E., Guvenalp, Z.& Demirezer, L.O.(2012). Investigation on antiinflammatory and antiulcer activities of Anchusa azurea extracts and their major constituent rosmarinic acid. Z. Naturforsch. C., 67(7-8):360.
- Abed, Alireza., Minaiyan, Mohsen., Ghannadi, Alireza., Mahzouni, Parvin.& Babavalian, Mohammad. Reza.(2012). Effect of Echium amoenum Fisch. etMey a Traditional Iranian Herbal Remedy in an ExperimentalModel of Acute Pancreatitis. ISRN Gastroenterology., 1.
- 39. Kweka, E.J., Senthilkumar, A.& Venkatesalu, V.(2012). Toxicity of essential oil from Indian borage on the larvae of the African malaria vector mosquito, Anopheles gambiae. Parasit Vectors., 3(5-227):1.
- 40. Ketabchi, Saghar. , Moatari, Afagh., Shadram ,Mostafa. & Rostami, Younes. (2011). Society of Applied SciencesThe Anti Influenza Virus Activity of Anchusa Italica. ASIAN J. EXP. BIOL. SCI., 2(4).
- 41. Monsefi, Malihezaman., Ghasemi, Maedeh.& Bahaoddini, Aminollah.(2006). The effects of anethum graveolens l. On female reproductive system of rats. Pharmaceutical Sciences., 14(3):131.
- 42. Jafarzadeh, L., Asgari, A., Golshan Iranpour, F., Kheiri, S., Parvin, N., Rafieian, M., Taji, F., Shahinfar, N., Rahimian, A. & Azadegan, F.(2009). The effect of Stachys lavandulifolia in induced abortion in mice. Shahrekord Medical Sciences Journal., 11(4): 26.
- 43. Iranloye, B., Oyeusi, K. &Alada, A.(2010). Effect of aqueous extract of phyllantus amarus Leaves on implantation and pregnancy in rats. Niger. J. Physiol. Sci., 25(1): 63
- 44. Nallasamy, S., Li, Q., Bagchi, M.K.& Bagchi, I.C.(2012).Msx homeobox genes critically regulate embryo implantation by controlling paracrine signaling between uterine stroma and epithelium. PLoS Genet., 8(2):1.
- 45. Jefferson, Wendy, N., Padilla-Banks, Elizabeth., Phelps Jazma, Y., Cantor, Amy, M. & Williams, Carmen, J.(2012). Neonatal Phytoestrogen Exposure Alters Oviduct Mucosal Immune Response to Pregnancy and Affects Preimplantation Embryo Development in the Mouse. Biology of Reproduction., 87(1-10):1.
- 46. Xiao, S., Diao, H., Smith, M.A., Song, X. & Ye, X.(2011).. Preimplantation exposure to bisphenol A (BPA) affects embryo transport, preimplantation embryo development, and uterine receptivity in mice. Elsevier., 32(4):434.
- 47. Liu, Y. & Wu, J.Z.(2006). Effect of Gutai Decoction on the abortion rate of in vitro fertilization and embryo transfer. Chin. J. Integr. Med., 12(3):189
- 48. Mboso, O.E., Eyong, E.U., Ebong, P.E., Iwara, A. & Odey, M.(2013), The effects of the ethanolic extract of Ereromastax speciosa leaf on the serum levels of leuteinizing hormone, follicle stimulating hormone, progesterone and estradiol in female pubertal rats. Annals of Biological Research., 42:136.
- 49. Ghasemi, Sh., Hemati, Kh., Bashiri Sadr, Z., Ghasemnejad, A. & Ghasemi, M.(2011). Some phenolic compounds of Citrus aurantifolia tissues in various stages of growth. Journal of Food Science and Technology., 8(31).
- 50. Gattuso, G., Barreca, David., Gargiulli, C., Leuzzi, U.& Caristi, C.(2007). Flavonoid composition of citrus juices. Molecules., 12: 1641-1673.
- 51. Steinborn, A., Günes, H., Röddiger, S. & Halberstadt, E.(1996). Elevated placental cytokine release, a process associated with preterm labor in the absence of intrauterine infection. *Obstetrics and Gynecology*, 88(4):534–539.
- 52. Romero, R., Mazor, M., Munoz, H., Gomez, R., Galasso, M.& Sherer, D.M.(1994). The preterm labor syndrome. *Annals of the New York Academy of Sciences.*, 734:414–429.
- 53. Bennett P.R., Elder M.G., Myatt L.(1987), The effects of lipoxygenase metabolites of arachidonic acid on human myometrial contractility. *Prostaglandins*,33(6):837–844.
- 54. Fujii, M., Horizoe, I., Fukunaga, T., Koga, K.& Aizono, Y.(1986). Appearance of triacylglycerol lipase in egg yolk sac of Japanese quail during embryonic development, its partial purification and some properties. Agric. Biol. Chem., 50:461–467.
- 55. Payal, dande. & suraj, patil.(2012). Evaluation of saponins from trigonella foenum graecum seeds for its antifertility activity. Asian Journal of Pharmaceutical and Clinical Research., 5(3).
- 56. Nieminen, P., Mustonen, A.M., Päiväläinen, P. & Kukkonen, J.V.K.(2004). Reproduction of the tundra vole (Microtus oeconomus) with dietary phytosterol supplement. Food Chem Toxicol., 42: 945-951.

- 57. Czech, P., Weber, K. & Dietrich, D.R.(2001). Effects of endocrine modulating substances on reproduction in the hermaphroditic snail Lymnaea stagnalis L. Aquat Toxicol., 53(2): 103-114.
- 58. Moallem, SA., Tafazzoli, M. & Niapour, M.(2003). studying the tratogenic effects of zingiber officinale on mouse, Iran journal of medical basic sciences., 6(1): 43.
- 59. Goolshan Iranpour, F., Jafarzadeh, L.& Asgari, A. (2012). tratogenic effects of Stachys lavandulifolia in skeleton system and growth of mouse fetus (Balb/c). Shahrkord university of medical sciences journal., 14(4): 21.
- 60. Baharara, J.& Rostampour, M.(2006). anomalies effects of matricaria chamomilla on small rats, Iran journal of gynecology, obstetrics and infertility., 9(2): 37.
- 61. Luqman Aribidesi, Olayaki., Ibiyemi, Olatunji-Bello., Ayodele Olufemi, Soladoye., Olusegun Rabiu, Jimoh., Olaide, Ghazal. & Martins, Ighodalo.(2009). Effects of aqueous leaf extract of Cajanus cajan on litter size and serum progesterone in pregnant rats. Journal of Pharmacognosy and Phytotherapy., 1(2):21.
- 62. Kermanian, F., Jaghtaei., MT.& Mehdizadeh, M.(2002). Studying tratogenic effects of Peganum.hermla on skeleton system and growth of mouse fetus having used red Alizarin painting, Iran journal of anatomy sciences., 1(1): 35.
- 63. Anvari, M., Dashti., MH., Zeinali, F.& Hosseini Biyouki, SM.(2011). Studying zataria multiflora boiss on status of bid laboratory rats fetus and placenta. herbal plants quarterly., 2(38): 19.
- 64. Nasimi, M., Heydari Nasrabadi, M.& Shiravi, A.(2008). Effects of Physalis alkekengi on placental growth of wistar pregnant rats. animal biology., 1(2): 51.
- 65. Morita, O., Knapp, J.F., Tamaki, Y., Stump, D.G., Moore, J.S. &Nemec, M.D.(2009). Effects of green tea catechin on embryo/fetal development in rats. Food Chem Toxicol., 47(6):1296.
- Chakravarthy, M.V., Zhu, Y., Wice, M.B., Coleman, T., Pappan, K.L., Marshall, C.A., McDaniel, M.L.& Semenkovich, C.F.(2008). Decreased fetal size is associated with beta-cell hyperfunction in early life and failure with age. Diabetes., 57:2698–2707.
- 67. Catalano, P.M. & Ehrenberg, H.M.(2006). The short- and long-term implications of maternal obesity on the mother and her offspring. B.J.O.G., 113:1126–1133
- 68. Warner, M.J. & Ozanne, S.E.(2010). Mechanisms involved in the developmental programming of adulthood disease. Biochem. J., 427:333–347.
- 69. Castañeda-Gutiérrez, E., Benefield, B.C., de Veth, M.J., Santos, N.R., Gilbert, R.O., Butler, W.R.& Bauman, D.E. (2007). Evaluation of, the mechanism of action of conjugated linoleic acid isomers on reproduction in dairy cows. J. Dairy. Sci., 90(9):4253-64.
- 70. Harris, M.A., Hansen, R.A., Vidsudhiphan, P., Koslo, J.L., Thomas, J.B., Watkins, B.A. & Allen, K.G.(2001). Effects of conjugated mlinoleic acids and docosahexaenoic acid on rat liver and reproductive tissue fatty acids, prostaglandins and mmatrix metalloproteinase production. Prostaglandins sLeukot Essent Fatty Acids.,65(1):23-9
- 71. Flint, AP., Sheldrick, E.L.& Fisher, P.A.(2002). Ligand-independent activation of steroid receptors. Domest Anim Endocrinol., 23(1-2):13-24.
- 72. Akahoshi, Asuka., Koba, Kazunori ., Ohkura-Kak, Shihoko u, Kaneda, Naoko ., Goto, Chikage , Sano, Hiroe ., Iwata, Toshio ·, Yamauchi, Yoshie ·,Tsutsumi, Kentaro.& Sugano ,Michihiro .(2003). Metabolic effects of dietary conjugated linoleic acid (CLA) isomers in rats. Nutr Res.,23(12):1691-701.
- 73. Yamasaki, M., Ikeda, A., Oji, M., Tanaka, Y., Hirao ,A., Kasai, M., Iwata, T., Tachibana ,H.& Yamada, K.(2003). Modulation of body fat and serum leptin levels by dietary conjugated linoleic acid in Sprague-Dawley rats fed various fat-level diets.Nutrition.,19

# **CITATION OF THIS ARTICLE**

Arezoo Z , Shahla R Examining the Effect of the Aqueous Extract of *Anchusa italica* on abortion and Fetal-placental parameters in Wistar Female Mice. Bull. Env. Pharmacol. Life Sci., Vol 4 [8] July 2015: 16-25