



Nuclear Polyhedrosis Virus (NPV): An overview

Prasun Mondal, Adesh Kumar* and Tammana

School of Agriculture, Lovely Professional University, Phagwara (Punjab), India-144411

E-mail: adesh.19078@lpu.co.in, phytopath06@gmail.com

ABSTRACT

*Nuclear polyhedrosis viruses (NPV) are baculoviruses, obligate pathogens with rod-shaped nucleocapsid containing double stranded circular DNA. The normal way of infection is by ingestion of polyhedra or in some cases virions can infect host insects. These days the use of chemical pesticides affects the environment and cause health hazards to humans, animals and beneficial insects also. Nuclear Polyhedrosis virus is a good alternative to the synthetic pesticides. This virus can control lepidopteran insects of cultivated crops like potato, cotton, cabbage etc. This narrow spectrum biocide only targets the specific host i.e., insect pests not the non-target hosts which includes beneficial insects. Mass culture of NPVs on *Spodopteralitura* is easily possible by using natural diet, castor leaves under laboratory condition in plastic buckets. Spraying of NPV formulation should be late within the day when peak sunshine or evening escapes sun's rays to safeguard the microorganism particles to enhance the effectiveness of NPV.*

Keywords: Biopesticide, Entomopathogens, Nuclear Polyhedrosis Virus (NPV), Baculoviruses, Lepidoptera, Helicoverpa spp., Spodoptera spp.

Received 21.02.2021

Revised 30.04.2021

Accepted 22.05.2021

INTRODUCTION

Biopesticides or 'biological pesticides' is a part of biological control where pest management is through organisms of biological origin or their products like virus, bacterium, fungus and protozoan [37]. In Pest management mechanisms involve parasitic, predatory, or chemical relationships. Pest refers to any organism (insect, non-insects, microbial pathogens, weeds etc. detrimental to man and his properties. European Union told biopesticides is a type of pesticide which is based on microorganisms or natural products.

HISTORY OF NPV

In history During 1892 Germany first introduced NPV into the population of *Lymantria monacha* in pine forest. During 1913 first field application of NPV was against gypsy moths in the U.S.A; a Californian farmer collected NPV killed larvae of *C.eurytheme* as an inoculum for application on alfalfa crop. In 1975, U.S.A registered first viral insecticide (Elcar) used against cotton bollworm (*Helicoverpa Zea*) [1, 39]. Baculoviruses give an alternate of chemical insecticide for controlling different insect pest because it don't have any residue effect and don't have any harmful effect on beneficial predators and parasites. The *Helicoverpa* and *Heliothis* attack more than 65 crops like maize, wheat, sorghum various legumes, solanaceous crop, malvaceous crop, etc.[8]. Many important spp. of *Helicoverpa* and *Heliothis* found sensitive to NPV i.e., *Helicoverpa armigera*, *Helicoverpa assulta*, *Heliothis virescens* and *Helicoverpa zea*.

STRUCTURE

Nuclear polyhedrosis virus is an obligate pathogen. The virus is rod-shaped nucleocapsid (250-400*40-70nm), containing double stranded circular DNA [36, 58]. Nucleocapsids are enclosed within an envelope called virions consisting of a proteinaceous polyhedral occlusion body inside which the virions or virus rods are embedded [4, 15]. Due to alkaline gut juice the viruses are liberated, which attack the nuclei of cell tissue, fat bodies, hemocytes, tracheal matrix, ganglia and brain [18].

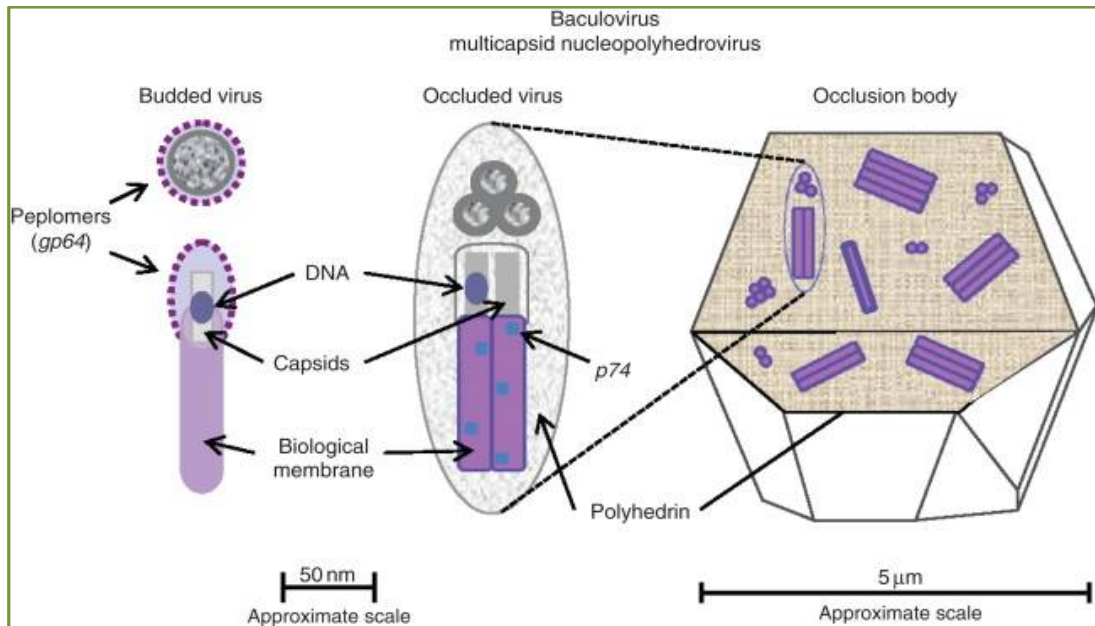


Fig. Structure of Baculovirus; a multicapsidnucleo-poly-hedro-virus[20]

INFECTION PROCESS

The normal way of infection is by ingestion of polyhedra or in some cases virions. Larvae should be susceptible; when polyhedra are ingested with the food the occlusion protinacious bodies start dissolve under the alkaline (pH>9)condition. The virions infect the epithelial cell of midgut; in the nucleus of infected cells newvirions are produce and infect cells of hemocoel and other tissue such as fat body [47]. In that tissue occlusion of virions in polyhedra take place & that possess keep going until cell lysis. After 24 to 72hr infection polyhedra can be detected in the nuclei of infected cells. After a few days larvae die and spilled on the foliage and soil. After that predators and parasite eat or distribute to other insect or place [13, 43].

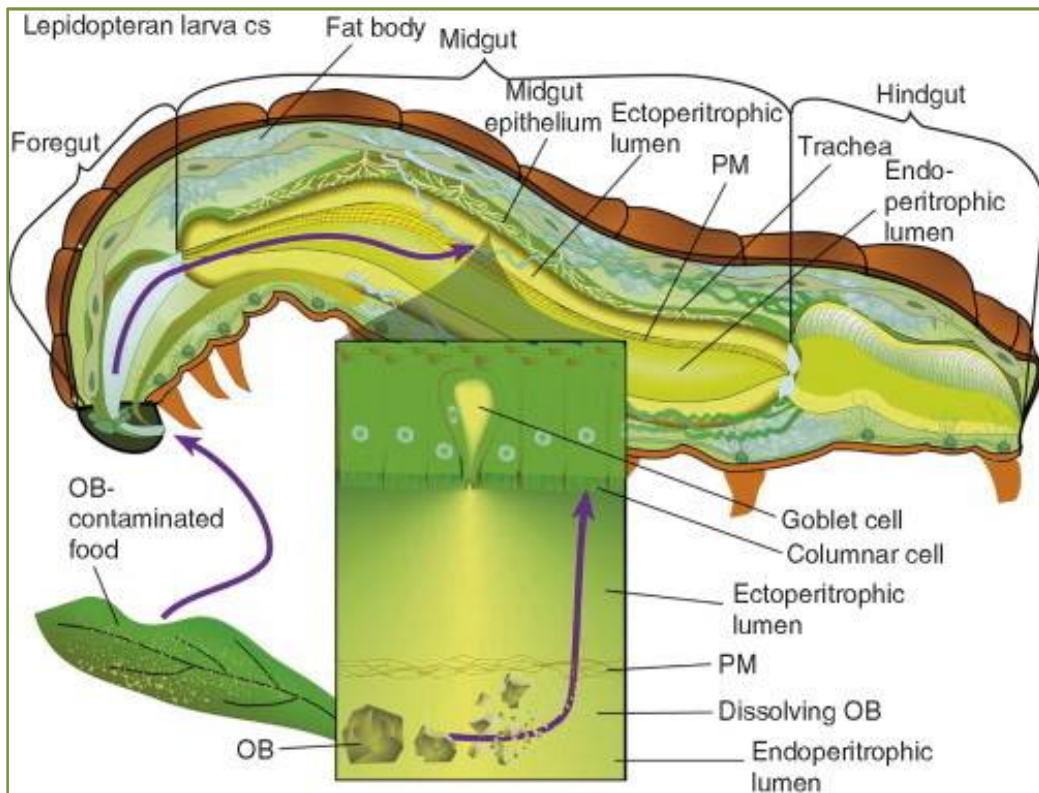


Fig. Mechanism of infection of lepidopteran larvae by NPV[57]

Infected insects are dull in color, less active and the larvae turn reddish pink on the ventral side. With advanced stages, the worms become flaccid, the skin becomes very fragile and eventually breaks down [11]. Infected larvae hang upside down on plants. This is called "tree top disease" or "wipfelkrankheit" Diseases caused by NPV in silkworms are called Grasserie [2].

THE EFFECT OF ENVIRONMENT ON NPV

The virus came out from the dead larvae which are resistant to abiotic factors such as humidity, drought, pressure etc. become inactive by sunlight and UV radiation range 280 to 320 nm is harmful for the NPV [32].

Effectiveness of NPV depends on environment

As foliar application virus can be inactive after a few days but in soil polyhedra can stay active for more than 10 years [26]. During rain splash polyhedra can contaminate foliage.

MASS PRODUCTION OF NPV

The mass production of the NPV is carried out in the early fifth instar stage of the *Spodoptera litura* which yields the maximum amount of the NPV. Therefore in the host culture laboratory a continuous culture of the insects is maintained with proper handling procedures [5].

The larvae are grown in a diet held in 5ml glass vials when the larvae reach the appropriate stage they are transferred to virus production facility. The NPV is multiplied by feeding the semi synthetic diet coated with a clean inoculum of the NPV that has previously been standardized. This is accomplished by placing aliquots of 10ml of viral suspension of concentration 1×10^8 polyhedral occlusion bodies (POB) in the center over the diet surface either in glass vials and spreading the suspension uniformly all over the surface with a polished glass rod. Larvae are released singly after 15 min, into each glass vial/cell and incubated at 25°C for 10 days. The larvae begin to die from 5th day onward. The cadavers are collected individually and transferred to 500ml plastic containers and frozen immediately until processing. The processing method is similar to that of *Helicoverpa armigera* (TNAU) [56].

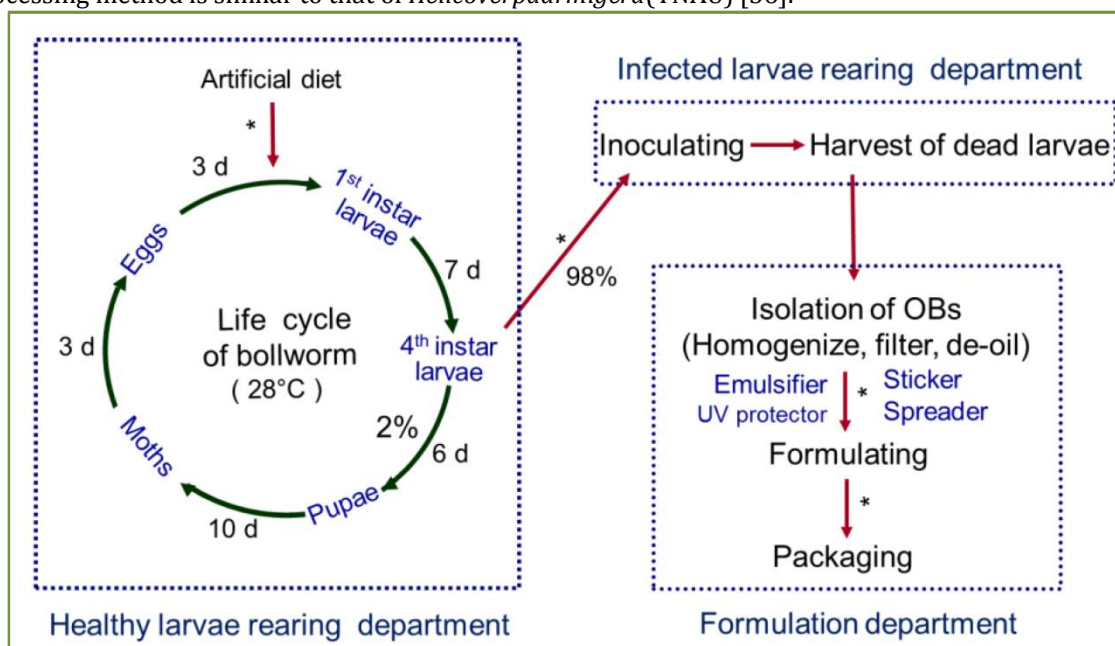


Fig. Infection cycle by NPV [54]

The cadavers are unit homogenized in sterile ice cold water at the magnitude relation 1:2.5 (w/v) during a liquidizer or cool all glass pestle and mortar. The material is filtered through double stratified cloth and repeatedly washed with water. The magnitude relation of water to be used for this purpose is 1:7.5-12.5 (w/v) for the first weight of the corpse processed [30]. The leftover mat on the cloth is discarded and also the filtrate may be semi-purified by differential activity. The filtrate is centrifuged for 30-60 sec at 500rpm to get rid of junk. The supernatant is next centrifuged for 20min at 5,000 rpm. Then the pellet containing the solid occlusion bodies (POB) is suspended in sterile water and washed 3 times by centrifuging the pellet in water at low rate followed by activity at high rate. The pellet finally collected is suspended in water and created upto a famed volume that is important to calculate the strength of the letter box within the refined suspension [60].

IN VIVO PRODUCTION TECHNIQUE

All industrial production of NPV for use as biopesticide is therefore currently done in vivo by infecting, rearing and harvesting the whole insect. Systems ranging from simple home pesticide involving farmers using the field collected larvae to the computer controlled robot operated mass production facilities with capacity of 0.25-1 million ha per annum. Examples of such modern mass production facilities have been built in France, Canada and USA [35, 58]

In vivo production technique completes into three steps: 1. Infect larvae susceptible species, 2. Grow infected larvae for a time to allow the infection to develop and the virus to replicate and 3. Harvest the larvae and extract the virus [35].

However in practice maintaining a sustained production of virus has not been found to be easy. It is only with well- trained staff, adequately developed production procedures, the appropriate equipment and a high standard of process quality control that effective production can be both attained and maintained [16].

TYPICAL PROBLEMS FOUND WITH PRODUCTION

1. Reduced production rate per insect over time.
2. Contamination of the system by other competing pathogens reduces the NPV yield as well as lowering the quality of the product.
3. Failure to maintain a supply of healthy insects [16].

The causes of problems are various but important factors are

1. Poor quality insects; impure inoculum.
2. Inappropriate dosing.
3. Poor rearing.
4. Unsuitable harvesting.
5. Poor sanitation.

MEASURING THE STANDARD OF VIRUS

Measure the standard of virus by Haemocytometer. Either dark field or section distinction magnifier is required to count the solid inclusion bodies (PIB) employing a Haemocytometer. The dose of virus is expressed as the larval equivalent (LE) and one LE is 6×10^9 POB [31, 32].

APPLICATION OF NPV

Spraying the NPV late within the day when peak sunshine or evening escape sun's rays to safeguard the microorganism particles to enhance the effectiveness of NPV. Adding ultraviolet light absorbent use of 1ml capacity unit of robin blue to a liter of spray solution has been reported as improving the effectiveness of the NPV [16]. Also, addition of an adhesive like teepoland and a phagostimulant like jiggery is suggested to improve the efficacy. The dosage lies between 250-500 LE/ ha relying upon the foliage density [1].

Application Systems**1. High-volume**

Many Spraying systems we can use for applying nuclear polyhedrosis viruses in crops. High-volume applications were made with an azo-propen hand sprayer using a birchmeier helicon sapphire nozzle 100, pressure 4 bar. This sprayer nozzle should be 50 cm over the crop during spray time [3].

2. Low-volume

Low-volume spinning disc applications were made by micron-ulva eight spinning discs. This sprayer nozzle should be 75 cm over the crop during spray and droplets angle should be 50 degree. Volume should be apply 3 ml virus suspension in water with 20% mineral oil per meter square [3].

3. Ultra-low volume electrodynamic sprayer

In ultra-low volume electrodynamic sprayer use of electrodynamic provided by ICI [7], in this sprayer nozzle position should be 50 cm above the crop [3].

MAJOR INSECT PESTS AND THEIR MANAGEMENT BY NPV***Spodoptera litura***

Spodoptera litura (Lepidoptera: Noctuidae) is a polyphagous pest of different crops and it has attained global importance due to its damaging range and insecticide resistance problems. SINPV can be good alternate for the control of *Spodoptera litura* on cabbage [33, 56].

MODE OF APPLICATION OF SINPV

SINPV is applied in the field @ rates of 500 lit/ha. Two sprays are required; first spray 45th days after transplanting of cabbage in the field and second spray at the time of the 60th day after transplanting.

Helicoverpa armigera

Helicoverpa armigera Hubner (Noctuidae: Lepidoptera) ranks as the most important

lepidopteran pest in South Africa [1, 17] Included in its wide range of economically important agricultural crops is citrus, on which it can be a serious pest. The moths lay their eggs on or near the blossoms during spring.

MODE OF APPLICATION OF HaNPV

A concentration of 1.15×10^7 OBs/ml of HaNPV is sprayed using a knapsack, resulting in a 100% reduction in *H. armigera* larval infestation within 7 days on tomato plants in a hot house environment. A 10-fold lower concentration, 1.15×10^6 OBs/mL, resulted in a 100% reduction within 16 days. **Table 1 and Table 2.**

Table 1 Worldwide infestation by larvae of *Spodoptera* spp. on various plant species

strawberry	Lettuce	sage	Citrus	coffee
sugar beat	Maize	Potato	Chilli	cotton
Sweet pepper	Marigold	Safflower	carrot	Eggplant
wheat	Tomato	sesame	Cabbage	Eucalyptus
Turnip	millet	Purslane	Bean	Grape
sunflower	Mint	Pear	Barley	Spinach
Rice	Onion	Tobacco	Apple	Jute
Pea	Lentil	Redish	Alfalfa	Indigo

Source: [16, 30, 60]

Table 2: Insect pests of plants and their management by NPV

Scientific name of Host insect	Common name in India	Associated crops	References
<i>Helicoverpa armigera</i>	cotton bollworm	Cotton, Tomato, Wheat	[38, 43]
<i>Helicoverpa zea</i>	Corn earworm	Maize	[17, 48]
<i>Heliothis virescens</i>	Tobacco Budworm	cotton	[24]
<i>Hyphantriacunea</i>	Fall webworm	Mulberry	[22, 61]
<i>Buzurasup pressaria</i>	Tea looper	Tea & Tung oil tree	[36, 61]
<i>Autographa californica</i>	Alfalfa looper	Cabbage, cotton & Ornamentals	[55]
<i>Anticarsia gemmatalis</i>	Bean caterpillar	Soybean	[41, 60]
<i>Spodopteralitura</i>	Tobacco cutworm	Vegetables, rice and peanuts	[33, 56, 58]
<i>Spodoptera exigua</i>	Army worm	Cabbage, Cotton, Amaranthus and Sunflower	[23, 42]
<i>Spodoptera frugiperda</i>	Fall armyworm	Maize	[44]
<i>Spodopteralittoralis</i>	cotton leaf worm	Cotton	[30]
<i>Lymantiradispar</i>	Gypsy moth	Forest	[31]
<i>Mamestrabraccae</i>	Cabbage moth	Cabbage	[66]

CONCLUSION

By this review paper we can conclude that Organic foods are the safe food for the human body. NPV is a very good option of bio pesticide available for farmers which will not cause any harm to nature. The initial cost may be little high but within *in vivo* technique we can replace the chemical pesticides in long term. For production of NPV skilled farmers are required which may be fulfilled by giving training to the farmers.

REFERENCES

- Arora, R., & Padmanaban, N. (2002). Field Evaluation of Native Nucleopolyhedrovirus for the Management of Sunflower Head borer, *Helicoverpa armigera* (Hübner). *Annals of Plant Protection Sciences*, 10(2), 184-187.
- Babu, K. R., Ramakrishna, S., Reddy, Y. H. K., Lakshmi, G., Naidu, N. V., Basha, S. S., & Bhaskar, M. (2009). Metabolic alterations and molecular mechanism in silkworm larvae during viral infection: A review. *African Journal of Biotechnology*, 8(6).
- Bateman, R. P., Matthews, G. A., & Hall, F. R. (2007). Ground-based application equipment. In *Field manual of techniques in invertebrate pathology* (pp. 73-98). Springer, Dordrecht. https://doi.org/10.1007/978-1-4020-5933-9_4
- Beniya, H., Funk, C. J., Rohrmann, G. F., & Weaver, R. F. (1996). Purification of a Virus-Induced RNA Polymerase from *Autographa californica* Nuclear Polyhedrosis Virus-Infected *Spodoptera frugiperda* Cells That Accurately Initiates Late and Very Late Transcription in Vitro. *Virology*, 216(1), 12-19. <https://doi.org/10.1006/viro.1996.0029>

5. Brown, E. S., & Dewhurst, C. F. (1975). The genus *Spodoptera* (Lepidoptera, Noctuidae) in Africa and the near east. *Bulletin of entomological research*, 65(2), 221-262. <https://doi.org/10.1017/S0007485300005939>
6. Cao, Q., Zhu, S. Y., Wu, Y., Liu, Y., Zhu, J., & Wang, W. (2012). The effect of a small conotoxin-like ctx gene from *Autographa californica* nuclear polyhedrosis virus (AcMNPV) on insect hemolymph melanization. *Polish journal of microbiology*, 61(3), 183.
7. Coffee, R. A. (1980). Electrodynamics energy-a new approach to pesticide application. In *Proceedings 1979 British Crop Protection Conference-Pests and Diseases*. (No. Volume 3, pp. 777-789).
8. Cunningham J.C. 1995. Cunningham, J. C. (1995). Baculoviruses as microbial insecticides. In "Novel Approaches to Integrated Pest Management", ed. R Reuveni, pp.261-92.
9. Im, D. J., Choi, K. M., Lee, M. H., Jin, B. R., & Kang, S. K. (1989). In vivo mass production of *Spodoptera litura* nuclear polyhedrosis virus. *Korean Journal of Applied Entomology*, 28(2), 82-87.
10. da Silveira, E. B., Cordeiro, B. A., Ribeiro, B. M., & Bao, S. N. (2005). In vivo apoptosis induction and reduction of infectivity by an *Autographa californica* multiple nucleopolyhedrovirus p35- recombinant in hemocytes from the velvet bean caterpillar *Anticarsia gemmatalis* (Hübner) (Lepidoptera: Noctuidae). *Research in microbiology*, 156(10), 1014-1025. <https://doi.org/10.1016/j.resmic.2005.06.001>
11. Deb, S., Chakraborty, S., Hazra, N., Saha, A., & Chakraborty, C. (2015). Grasserie in silkworm, *bombyx mori* in West Bengal. *Journal of Plant Protection and Environment*, 12(1), 19-25.
12. Doyle, C. J., Hirst, M. L., Cory, J. S., & Entwistle, P. F. (1990). Risk assessment studies: detailed host range testing of wild-type cabbage moth, *Mamestra brassicae* (Lepidoptera: Noctuidae), nuclear polyhedrosis virus. *Applied and environmental microbiology*, 56(9), 2704-2710.
13. Entwistle, P. F., Adams, P. H. W., Evans, H. F., & Rivers, C. F. (1983). Epizootiology of a nuclear polyhedrosis virus (Baculoviridae) in European spruce sawfly (*Gilpinia hercyniae*): spread of disease from small epicentres in comparison with spread of baculovirus diseases in other hosts. *Journal of Applied ecology*, 473-487. <https://doi.org/10.2307/2403521>
14. Fuxa, J. R., & Richter, A. R. (1989). Reversion of resistance by *Spodoptera frugiperda* to nuclear polyhedrosis virus. *Journal of Invertebrate Pathology*, 53(1), 52-56. [https://doi.org/10.1016/0022-2011\(89\)90073-6](https://doi.org/10.1016/0022-2011(89)90073-6)
15. Grula, M. A., Buller, P. L., & Weaver, R. F. (1981). α -Amanitin-resistant viral RNA synthesis in nuclei isolated from nuclear polyhedrosis virus-infected *Heliothis zea* larvae and *Spodoptera frugiperda* cells. *Journal of Virology*, 38(3), 916-921.
16. Grzywacz, D., Jones, K. A., Moawad, G., & Cherry, A. (1998). The in vivo production of *Spodoptera littoralis* nuclear polyhedrosis virus. *Journal of virological methods*, 71(1), 115-122. [https://doi.org/10.1016/S0166-0934\(97\)00209-7](https://doi.org/10.1016/S0166-0934(97)00209-7)
17. Hackett, K. J., Boore, A., Deming, C., Buckley, E., Camp, M., & Shapiro, M. (2000). *Helicoverpa armigera* granulovirus interference with progression of *H. zea* nucleopolyhedrovirus disease in *H. zea* larvae. *Journal of invertebrate pathology*, 75(2), 99-106. <https://doi.org/10.1006/jipa.1999.4914>
18. Harrap, K. A. (1970). Cell infection by a nuclear polyhedrosis virus. *Virology*, 42(2), 311-318. [https://doi.org/10.1016/0042-6822\(70\)90275-8](https://doi.org/10.1016/0042-6822(70)90275-8)
19. Harrap, K. A. (1972). The structure of nuclear polyhedrosis viruses: I. The inclusion body. *Virology*, 50(1), 114-123. [https://doi.org/10.1016/0042-6822\(72\)90351-0](https://doi.org/10.1016/0042-6822(72)90351-0)
20. Hitchman, R. B., Possee, R. D., & King, L. A. (2009). Baculovirus expression systems for recombinant protein production in insect cells. *Recent patents on biotechnology*, 3(1), 46-54.
21. Hughes, D. S., Possee, R. D., & King, L. A. (1993). Activation and detection of a latent baculovirus resembling *Mamestra brassicae* nuclear polyhedrosis virus in *M. brassicae* insects. *Virology*, 194(2), 608-615. <https://doi.org/10.1006/viro.1993.1300>
22. Hukuhara, T. (1973). Further studies on the distribution of a nuclear-polyhedrosis virus of the fall webworm, *Hyphantria cunea*, in soil. *Journal of Invertebrate Pathology*, 22(3), 345-350. [https://doi.org/10.1016/0022-2011\(73\)90161-4](https://doi.org/10.1016/0022-2011(73)90161-4)
23. Hussain, M. A., Pachori, R., & Choudhary, B. S. (2002). Polyhedral occlusion body counts of naturally diseased larvae of *Spodoptera litura* (Fab.) infected by SINPV on cabbage. *Journal of Entomological Research*, 26(1), 29-32.
24. Ignoffo, C. M., & Couch, T. L. (1981). nucleopolyhedrosis virus of *Heliothis* species as a microbial insecticide. *Microbial control of pests and plant diseases, 1970-1980*.
25. Jakubowska, A. K., Peters, S. A., Ziemnicka, J., Vlak, J. M., & van Oers, M. M. (2006). Genome sequence of an enhancer gene-rich nucleopolyhedrovirus (NPV) from *Agrotis segetum*: collinearity with *Spodoptera exigua* multiple NPV. *Journal of General Virology*, 87(3), 537-551. <https://doi.org/10.1099/vir.0.81461-0>
26. Jaques, R. P. (1985). Stability of insect viruses in the environment. In *Viral Insecticides for biological control* (eds.): Academic Press, pp. 285-360.
27. Jones, K. A., Moawad, G., McKinley, D. J., & Grzywacz, D. (1993). The effect of natural sunlight on *Spodoptera littoralis* nuclear polyhedrosis virus. *Biocontrol Science and Technology*, 3(2), 189-197. <https://doi.org/10.1080/09583159309355275>
28. Kalantari, M., Marzban, R., Imani, S., & Askari, H. (2014). Effects of *Bacillus thuringiensis* isolates and single nuclear polyhedrosis virus in combination and alone on *Helicoverpa armigera*. *Archives of Phytopathology and Plant Protection*, 47(1), 42-50. <https://doi.org/10.1080/03235408.2013.802460>
29. Kamita, S. G., & Maeda, S. (1993). Inhibition of *Bombyx mori* nuclear polyhedrosis virus (NPV) replication by the putative DNA helicase gene of *Autographa californica* NPV. *Journal of virology*, 67(10), 6239-6245.

30. Keddie, B. A., Aponte, G. W., & Volkman, L. E. (1989). The pathway of infection of *Autographa californica* nuclear polyhedrosis virus in an insect host. *Science*, 243(4899), 1728-1730. DOI: 10.1126/science.2648574
31. Kool, M., Voncken, J. W., Van Lier, F. L. J., Tramper, J., & Vlak, J. M. (1991). Detection and analysis of *Autographa californica* nuclear polyhedrosis virus mutants with defective interfering properties. *Virology*, 183(2), 739-746. [https://doi.org/10.1016/0042-6822\(91\)91003-Y](https://doi.org/10.1016/0042-6822(91)91003-Y)
32. Koul, O. (2012). Microbial biopesticides: Opportunities and challenges. *Biocontrol News and Information*, 33(2), 1R. *CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources* 6, No. 056. doi: 10.1079/PAVSNR20116056
33. Kumari, V., & Singh, N. P. (2009). *Spodoptera litura* nuclear polyhedrosis virus (NPV-S) as a component in Integrated Pest Management (IPM) of *Spodoptera litura* (Fab.) on cabbage. *Journal of Biopesticides*, 2(1), 84-86.
34. Lepore, L. S., Roelvink, P. R., & Granados, R. R. (1996). Enhancin, the granulosis virus protein that facilitates nucleopolyhedrovirus (NPV) infections, is a metalloprotease. *Journal of invertebrate pathology*, 68(2), 131-140. <https://doi.org/10.1006/jipa.1996.0070>
35. Lindegren, J. E., Valero, K. A., & Mackey, B. E. (1993). Simple in vivo production and storage methods for *Steinernemacarpocapsae* infective juveniles. *Journal of Nematology*, 25(2), 193.
36. Maeda, S. U. S. U. M. U., Kamita, S. G., & Kondo, A. (1993). Host range expansion of *Autographa californica* nuclear polyhedrosis virus (NPV) following recombination of a 0.6-kilobase-pair DNA fragment originating from *Bombyx mori* NPV. *Journal of Virology*, 67(10), 6234-6238.
37. Marrone, P. G. (2009). Barriers to adoption of biological control agents and biological pesticides. *Integrated Pest Management*. Cambridge University Press, Cambridge, UK, 163-178.
38. Moore, S. D., Pittaway, T., Bouwer, G., & Fourie, J. G. (2004). Evaluation of *Helicoverpa armigera* nucleopolyhedrovirus (HearNPV) for control of *Helicoverpa armigera* (Lepidoptera: Noctuidae) on citrus in South Africa. *Biocontrol Science and Technology*, 14(3), 239-250. <https://doi.org/10.1080/09583150310001655666>
39. Moscardi, F. (1999). Assessment of the application of baculoviruses for control of Lepidoptera. *Annual review of entomology*, 44(1), 257-289. <https://doi.org/10.1146/annurev.ento.44.1.257>
40. Moscardi, F., & Sosa-Gómez, D. R. (2007). Microbial control of insect pests of soybean. In *Field manual of techniques in invertebrate pathology* (pp. 411-426). Springer, Dordrecht. https://doi.org/10.1007/978-1-4020-5933-9_19
41. Moscardi, F., Allen, G. E., & Greene, G. L. (1981). Control of the velvetbean caterpillar by nuclear polyhedrosis virus and insecticides and impact of treatments on the natural incidence of the entomopathogenic fungus *Nomuraearileyi*. *Journal of Economic Entomology*, 74(4), 480-485. <https://doi.org/10.1093/jee/74.4.480>
42. Murray, D. A. H., Monsour, C. J., Teakle, R. E., Rynne, K. P., & Bean, J. A. (1995). Interactions between nuclear polyhedrosis virus and three larval parasitoids of *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae). *Australian Journal of Entomology*, 34(4), 319-322. <https://doi.org/10.1111/j.1440-6055.1995.tb01348.x>
43. Nawaz, A., Ali, H., Sufyan, M., Gogi, M. D., Arif, M. J., Ranjha, M. H., & Ghramh, H. A. (2019). Comparative bio-efficacy of nuclear polyhedrosis virus (NPV) and Spinosad against American bollworm, *Helicoverpa armigera* (Hubner). *Revista Brasileira de Entomologia*, 63(4), 277-282. <https://doi.org/10.1016/j.rbe.2019.09.001>
44. Podgwaite, J. D., Shields, K. S., Zerillo, R. T., & Bruen, R. B. (1979). Environmental persistence of the nucleopolyhedrosis virus of the gypsy moth, *Lymantria dispar*. *Environmental Entomology*, 8(3), 528-536. <https://doi.org/10.1093/ee/8.3.528>
45. Qayyum, M. A., Wakil, W., Arif, M. J., & Sahi, S. T. (2015). *Bacillus thuringiensis* and Nuclear Polyhedrosis Virus for the Enhanced Bio-control of *Helicoverpa armigera*. *International Journal of Agriculture and Biology*, 17(5).
46. Rabindra, R. J., Sathiah, N., & Jayaraj, S. (1992). Efficacy of nuclear polyhedrosis virus against *Helicoverpa armigera* (Hbn.) on *Helicoverpa*-resistant and susceptible varieties of chickpea. *Crop Protection*, 11(4), 320-322. [https://doi.org/10.1016/0261-2194\(92\)90056-B](https://doi.org/10.1016/0261-2194(92)90056-B)
47. Rahman, M. M., & Gopinathan, K. P. (2004). Systemic and in vitro infection process of *Bombyx mori* nucleopolyhedrovirus. *Virus research*, 101(2), 109-118. <https://doi.org/10.1016/j.virusres.2003.12.027>
48. Rowley, D. L., Popham, H. J., & Harrison, R. L. (2011). Genetic variation and virulence of nucleopolyhedroviruses isolated worldwide from the heliothine pests *Helicoverpa armigera*, *Helicoverpa zea*, and *Heliothis virescens*. *Journal of invertebrate pathology*, 107(2), 112-126. <https://doi.org/10.1016/j.jip.2011.03.007>
49. Santiago-Alvarez, C., & Ortiz-Garcia, R. (1992). The influence of host plant on the susceptibility of *Spodoptera littoralis* (Boisd.) (Lep., Noctuidae) larvae to *Spodoptera littoralis* NPV (Baculoviridae, Baculovirus). *Journal of Applied Entomology*, 114(1-5), 124-130. <https://doi.org/10.1111/j.1439-0418.1992.tb01106.x>
50. Sehgal, V. K., & Ujagir, R. (1990). Effect of synthetic pyrethroids, neem extracts and other insecticides for the control of pod damage by *Helicoverpa armigera* (Hübner) on chickpea and pod damage-yield relationship at Pantnagar in northern India. *Crop protection*, 9(1), 29-32. [https://doi.org/10.1016/0261-2194\(90\)90042-6](https://doi.org/10.1016/0261-2194(90)90042-6)
51. Slack, J., & Arif, B. M. (2006). The baculoviruses occlusion-derived virus: virion structure and function. *Advances in virus research*, 69, 99-165. [https://doi.org/10.1016/S0065-3527\(06\)69003-9](https://doi.org/10.1016/S0065-3527(06)69003-9)
52. Smits, P. H. (1987). *Nuclear polyhedrosis virus as biological control agent of Spodoptera exigua* (Doctoral dissertation, Smits).
53. Somasekar, S., Jayapragasam, M., & Rabindra, R. J. (1993). Characterization of five Indian isolates of the nuclear polyhedrosis virus of *Helicoverpa armigera*. *Phytoparasitica*, 21(4), 333-337. <https://doi.org/10.1007/BF02981052>

54. Sun, X. (2015). History and current status of development and use of viral insecticides in China. *Viruses*, 7(1), 306-319. <https://doi.org/10.3390/v7010306>
55. Sun, X. L., & Peng, H. Y. (2007). Recent advances in biological control of pest insects by using viruses in China. *Virologica Sinica*, 22(2), 158-162.
56. Takatsuka, J., Okuno, S., Ishii, T., Nakai, M., & Kunimi, Y. (2007). Productivity and quality of polyhedral occlusion bodies of a nucleopolyhedrovirus harvested from *Spodoptera litura* (Lepidoptera: Noctuidae) larvae. *Applied entomology and zoology*, 42(1), 21-26. <https://doi.org/10.1303/aez.2007.21>
57. Tettamanti, G., Grimaldi, A., Pennacchio, F., & de Eguileor, M. (2007). Lepidopteran larval midgut during prepupal instar: digestion or self-digestion?. *Autophagy*, 3(6), 630-631. <https://doi.org/10.4161/auto.4908>
58. Tinsley, T. W., & Kelly, D. C. (1985). Taxonomy and nomenclature of insect pathogenic viruses. In "Insecticides for Biological Control" (K. Maramorosch and KE Sherman, eds.), Vol. 3.
59. Trang, T. T. K., & Chaudhari, S. (2002). Bioassay of nuclear polyhedrosis virus (NPV) and in combination with insecticide on *Spodoptera litura* (Fab). *Omonrice*, 10, 45-53.
60. Veenakumari, K., Rabindra, R. J., Naik, C. S., & Shubha, M. R. (2007). In situ field level mass production of *Amsactaalbistriga* (Lepidoptera: Arctiidae) nucleopolyhedrovirus in a groundnut ecosystem in South India. *International Journal of Tropical Insect Science*, 27(1), 48-52. <https://doi.org/10.1017/S1742758407746623>
61. Yang, M. M., Li, M. L., an Zhang, Y., Wang, Y. Z., Qu, L. J., Wang, Q. H., & Ding, J. Y. (2012). Baculoviruses and insect pests control in China. *African Journal of Microbiology Research*, 6(2), 214-218. <https://doi.org/10.5897/AJMR11.1357>
62. Ye, S., & Tiong, R. L. (2000). NPV-at-risk method in infrastructure project investment evaluation. *Journal of construction engineering and management*, 126(3), 227-233. [https://doi.org/10.1061/\(ASCE\)0733-9364\(2000\)126:3\(227\)](https://doi.org/10.1061/(ASCE)0733-9364(2000)126:3(227))

CITATION OF THIS ARTICLE

P Mondal, A Kumar and Tammana. Nuclear Polyhedrosis Virus (NPV): An overview. *Bull. Env. Pharmacol. Life Sci.*, Vol10[6] May 2021 : 12-19