



Cadmium–Zinc Interaction in Relation to Lettuce Growth

Bechan Singh*, Dinesh Mani, Vipin Sahu, Adarsh Bhushan, Pravesh Kumar, Devi Prasad Shukla and Himanchal Vishwakarma

Sheila Dhar Institute of Soil Science, Department of Chemistry, University of Allahabad, Prayagraj-211002, (U.P.), India.

*Corresponding Author's Email id- bechansinghch@gmail.com

ABSTRACT

A field experiment was conducted on alluvial soil (Entisols) of Sheila Dhar Institute Experiment Farm, Prayagraj to find out the possible effect of Zinc on yield and phytoaccumulation of Cadmium in Lettuce. Cd was applied as CdCl_2 @ 0, 20 and 40 mg kg^{-1} and Zn was applied as ZnSO_4 @ 0, 40 and 80 mg kg^{-1} . It was observed that Zn application up to 80 mg kg^{-1} increased the dry biomass yield of lettuce by 20.37% over the control and decreased Cadmium concentration particularly in shoot and root of lettuce. However, the applications of higher dose of Cadmium without Zn reduced the yield of lettuce and increased the Cd concentration in shoot and root of lettuce by 3.92 and 4.73 mg kg^{-1} respectively. The reduced uptake of Cadmium was observed in Zinc treated plots. The authors conclude that the application of ZnSO_4 @ 80 mg kg^{-1} reduced the Cd uptake in shoot and root of lettuce grown in Cd-contaminated soils.

Keywords: Cadmium, Zinc, Lettuce, phytoaccumulation

Received 24.02.2021

Revised 28.04.2021

Accepted 12.05.2021

INTRODUCTION

Toxic heavy metals are accumulated in agricultural soils due to human activities such as industrialization, mining, and agricultural practices [1,2,3,4]. Among these metals, cadmium (Cd) needs special attention due to its high mobility mainly in the soil-plant system as well as Cd toxicity to humans even at very low concentrations [5,6]. Cadmium enters into the food chain mainly through food crops grown in Cd-contaminated soil [7,8,9,10]. Higher Cd concentrations in plants caused toxicity at physiological, morphological, and molecular levels [11,12,13].

The rapid development of urbanization and the expansion of industrial sectors in and around urban cities are characteristically caused by the accumulation and contamination of urban soil heavy metals, which are greatly affected by the urban soil environment. Typically, soil heavy metals are extremely introduced into the urban environment through urban waste, waste disposal, industrial effluents, vehicle emissions, construction waste, and huge usage of agrochemicals [14, 15, 16, 17]. As a result, larger than five million sites are severely contaminated by various soil heavy metals around the world [18,16].

Vegetables are an important source of human diets, and their contamination can cause serious health problems [19,20,21,22,23,24]. Leafy vegetables like lettuce are considered as potential hyperaccumulators of heavy metals [25]. One of the properties of green leafy vegetables is the accumulation of heavy metals in their tissues without exhibiting any toxicity symptoms [26].

The risk of Cd uptake by crops, followed by the transfer in the food chain, is an issue of high concern is now a day. Lettuce is a worldwide important crop, one of the most consumed leafy vegetables in the human diet [27] and a high Cd-accumulating species [28]. The bioavailability of heavy metals in a plant varies for different plant organs, and the absorption and bioaccumulation rate is highest for roots as compared to other parts [29]. The mean heavy metal uptake by plants increases as the contents of these metals increase in the soil environment [30].

Several studies have been conducted on the effect of Cd and Zn interaction on lettuce but the studies regarding the impact of Zinc on the Cd-contaminated soil are hardly observed. In recent years, the concentrations of heavy metals are continuously increasing considerably in this area due to untreated sewage irrigation. However, Zn deficiency has also been reported in the study area. Lettuce is a leafy vegetable that can uptake a high concentration of heavy metals than other crops. Therefore, the present study was undertaken to assess the effect of Cadmium, Zinc interaction on dry biomass yield of lettuce and uptake of Cadmium in the shoot and root of lettuce.

MATERIAL AND METHODS

Plant material and experimental layout

The Sheila Dhar Institute Experimental Site covers an area of 1 hectare, is located at Prayagraj in northern India at 25° 57' N latitude, 81° 50' E longitude and at 120 ± 1.4 m altitude. A sandy clay loam soil, derived from Indo-Gangetic alluvial soils, situated on the confluence of rivers Ganga and Yamuna alluvial deposit, was sampled for the study. The texture was sand (>0.2 mm) 55.54 %, silt (0.002–0.2 mm) 20.22 % and clay (<0.002 mm) 24.24 %. The physical properties were: pH-(7.8 ± 0.2), EC(dS/m) at °C-(0.28 ± 0.03), (%) - 0.038 ± 0.01, Total Cd (mg kg⁻¹) 0.17 ± 0.02 DTPA-extractable Cd (mg kg⁻¹) 1.56 ± 0.57 organic carbon % -(0.56 ± 0.15), CEC [C mol (p+)/kg]- 19.6 ± 0.6, Total nitrogen (%) - 0.07 ± 0.02.

(Note- ± values indicate standard error having three replications, EC- electrical conductivity, CEC-cation exchange capacity, DTPA diethyl)

After a systematic survey factorial experiment was conducted to study the effect of Cadmium-Zinc Interaction in relation to lettuce growth. The experiment was replicated thrice with nine treatments and conducted in a completely factorial Randomized Block Design (factorial RBD). After 24 hrs of the treatment seeds were sown. Soil moisture was maintained by irrigating the crops at an interval of 5-6 days. Lettuce was grown successively in the 27 plots (each of 1m² in the area). The treatments of Cd × Zn consisted of 0, 20, 40 mg kg⁻¹ Cd along with 0, 40, 80 mg kg⁻¹ Zn. The source of Cd and Zn were CdCl₂ and ZnSO₄.

Soil sampling

The larger fields were divided into suitable and uniform parts, and each of these uniform parts was considered a separate sampling unit. In each sampling unit, soil samples were drawn from several spots in a zigzag pattern, leaving about a 2-m area along the field margins. Silt and clay were separated by the pipette method and fine sand by decantation [31]

Soil chemical analysis

Soil pH was measured in a suspension of 1:2.5 soil water ratio by Elico digital pH meter (Model LI127, Elico Ltd, Hyderabad, India). Double Distilled Water was used for preparing of all kinds of solutions. Organic carbon of soil was determined by rapid chromic acid digestion method of Walkley and Black. Cation exchange capacity (CEC) was determined by neutral 1N ammonium acetate. Total nitrogen was determined by micro-Kjeldahl method (Glass Agencies, Ambala, India) containing digestion mixture of sulphuric acid, selenium dioxide and salicylic acid. Available sulphur extracted with CaCl₂ solution (0.15%) was determined by turbidimetric method. Total phosphorus was determined by hot plate digestion using HNO₃ (16M, 71%) and extracted by standard ammonium molybdate solution [31].

Extraction for Cadmium (Cd) content in the soil

For total Cd content, one gram of soil was mixed in 5 ml of HNO₃ (16 M, 71 %) and 5 ml of HClO₄ (11 M, 71 %). The composite was heated up to dryness. The hot distilled water was added. The contents were filtrated, and the volume was made up to 50 ml. The clean filtrate was used for the estimation of heavy metals (Cd) by atomic absorption spectrophotometer (AAS) (Analyst 600, PerkinElmer Inc., MA, USA). For available Cd, 5 gram of soil was mixed with 20 ml DTPA solution {Di-ethyl-triamine-penta acetic acid (DTPA) solution [1.97 g (0.05 M) DTPA powder, 13.3 ml (0.1 M) Tri-ethanol amine and 1.47 g (0.01 M) CaCl₂ were dissolved in distilled water [32] and were made up to 1 l after adjusting the pH to 7.3] was added} and the contents were shaken for 2 h and then filtered through Whatman filter paper No. 42. The clean filtrate was used for the estimation of Cd by the aforesaid spectrophotometer.

Processing plant samples

Plants were harvested after 60 days having higher phytochemicals at their maturity stage as suggested by Mani *et al.* (2012). Plant samples were carefully rinsed with tap water followed by 0.2 % detergent solution, 0.1 N HCl, deionizer water and double-distilled water. Samples were then soaked with tissue paper, air-dried for 2–3 days in adjust and contaminant-free environment, placed in clean paper envelopes, dried in a hot-air oven at a temperature of 45 C, and ground to a fine powder. Plant biomass dry weights were recorded. Shoot and root were separated and analyzed.

Determination of Cadmium in plant extract

One gram of ground plant material was digested with 15 ml of a tri-acid mixture containing conc. HNO₃ (16 M, 71 %), H₂SO₄ (18 M, 96 %) and HClO₄ (11 M, 71 %) in 5:1:2. The composite was heated on a hot plate at low heat (60 C) for 30 min, and the volume was reduced to about 5 ml until a transparent solution was obtained. After cooling, 20 ml distilled water was added and the content was filtered through Whatman filter paper No. 42 [33]. Total Cd was determined by the AAS.

Data analysis

The experimental results were expressed as mean ± standard error of mean (SEM) of three replicates. Graph pad Prism (version 9, Graph Pad Software, USA) software was used for drawing Figures.

RESULTS AND DISCUSSION

Effect of Cd × Zn interaction on the yield of lettuce:

Data (fig.1) indicate a significant influence of Cd, Zn & Cd × Zn interaction on the dry biomass yield of lettuce. The application of the maximum dose of Zn 80 mg kg⁻¹ (T₃) increased the highest yield of plant 20.37% over the control. Application of Zn boosts the yield of lettuce and was found to play an ameliorative role in Cd-contaminated soil. Treatment of Cd 20 mg kg⁻¹ and 40 mg kg⁻¹ in without Zn treated plot significantly decreased the dry matter 15.99% and 21.89% content over the control respectively as the dose of Cd increased from 0-40 mg kg⁻¹. Combined treatment of Cd 20 mg kg⁻¹ and Zn 40 mg kg⁻¹ (T₅) and Cd 40 mg kg⁻¹ and Zn 40 mg kg⁻¹ (T₇) decreased the yield 3.40% and 8.68 % respectively. Integrated application of Cd 20 mg kg⁻¹ and Zn 80 mg kg⁻¹ (T₆) and Cd 40 mg kg⁻¹ and Zn 80 mg kg⁻¹ (T₉) increased the yield of plant 7.92 % and 3.77% over the control respectively.

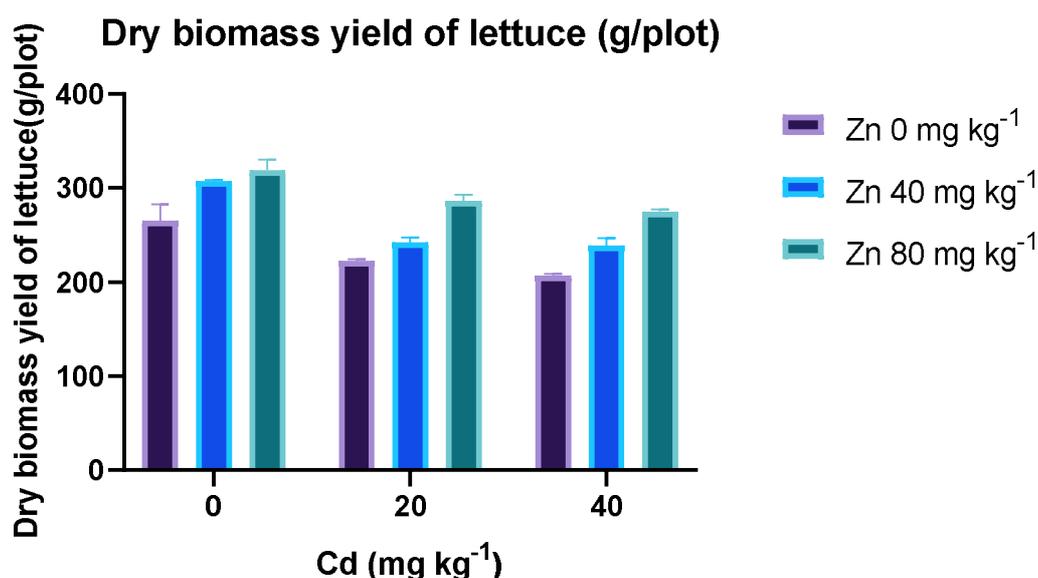


Fig.1: Effect of Cd × Zn interaction and ZSB on dry biomass yield of lettuce

[Note-Treatment combinations used under the pot experiment at Sheila Dhar, T₁-Control, T₂-Cd 0 mg kg⁻¹+Zn 40 mg kg⁻¹, T₃-Cd 0 mg kg⁻¹+ Zn 80, T₄-Cd 20 mg kg⁻¹, T₅-Cd 20 mg kg⁻¹+ Zn 40 mg kg⁻¹, T₆-Cd 20 mg kg⁻¹+Zn 80 mg kg⁻¹, T₇-Cd 40 mg kg⁻¹, T₈-Cd 40 mg kg⁻¹+40 mg kg⁻¹, T₉-Cd 40 mg kg⁻¹+80 mg kg⁻¹.]

Heavy metals have significantly negative effects on plant growth [34] other toxic effects may include root browning, alteration of mineral concentrations, and changes in photosynthesis [34]. Plants grown in soil containing high levels of Cd show visible symptoms of injury reflected in terms of chlorosis, growth inhibition, browning of root tips and finally death [35,36,37,38].

Effect of Cd × Zn interaction on Cd concentration in root and shoot of lettuce

The data graphically presented in figures (2a) and (2b) indicated that the effect of different treatments on the uptake of Cd in shoot contaminated and root of lettuce at experimental sites was observed highly significant. Cadmium concentration varies from 0.09 mg kg⁻¹ to 3.92 mg kg⁻¹ and 0.12 mg kg⁻¹ to 4.73 mg kg⁻¹ in the shoot and root respectively. Individual application of Zinc 80 mg kg⁻¹ (T₃) decreased the minimum concentration in the shoot and root of the plant by 0.09 mg kg⁻¹ and 0.12 mg kg⁻¹. Treatment of maximum dose of Cd 40 mg kg⁻¹ (T₇) without Zn was observed the maximum concentration of 3.92 and 4.73 mg kg⁻¹ in the shoot and root of lettuce. Combined application of Cd 20 mg kg⁻¹ and Zn 80 mg kg⁻¹ (T₆) reduced Cd uptake 1.52 and 1.82 mg kg⁻¹ in shoot and root of the plant. Integrated application of Cd 40 mg kg⁻¹ and Zinc 80 mg kg⁻¹ (T₉) mitigates Cd uptake 2.46 and 2.69 mg kg⁻¹ in the shoot and root of lettuce, respectively. Zinc decreased Cd translocation to shoots of wheat [39] and lettuce [40]. The plant accumulates a large portion of heavy metal in root followed by stem and leaf. The dry biomass of both roots and shoots was significantly reduced in Cd-treated plants compared to the control plants [41].

Cd concentration in root of lettuce (mg kg^{-1})

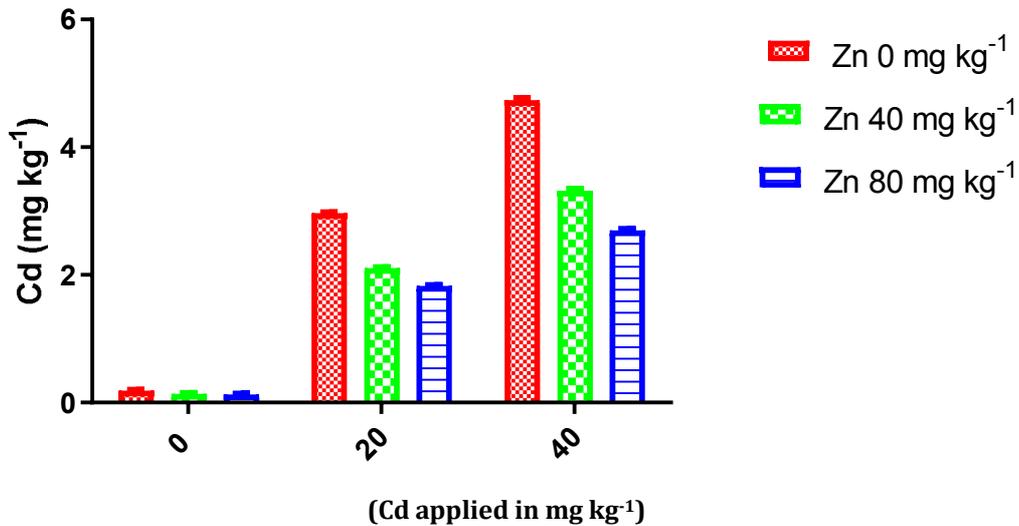


Fig. 2a: Effect of Cd \times Zn interaction on concentration in the root of lettuce

Cd concentration in shoot of lettuce (mg kg^{-1})

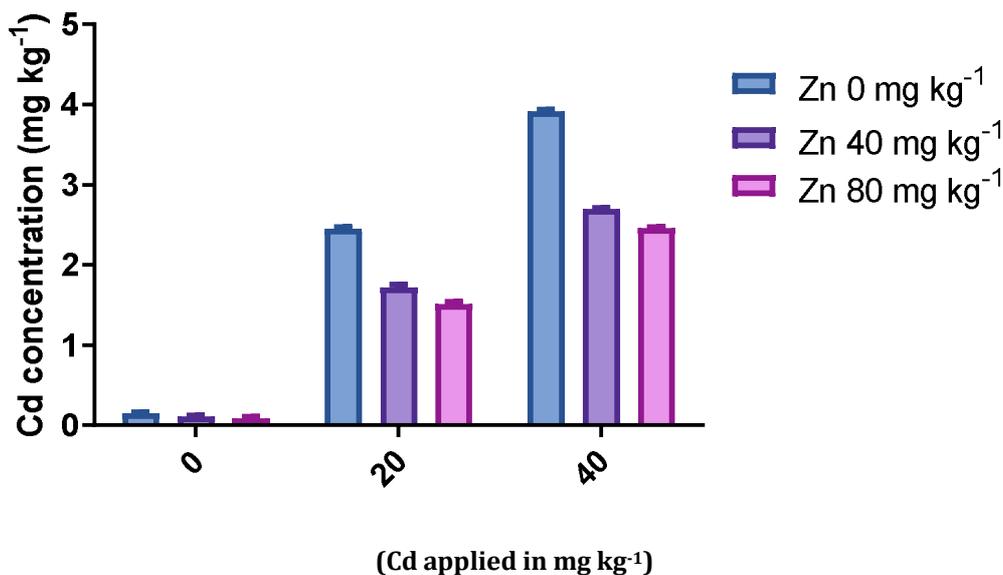


Fig.2b: Effect of Cd \times Zn interaction on Cd concentration in the shoot of lettuce

CONCLUSION

Application of maximum dose of Zn increased the highest potential to dry biomass of the plant and decreases the concentration of Cd in lettuce due to the reduction in the solubility of Cd in soil. Thus, a more detailed study is required to grow lettuce crops in Cd-polluted soil and evaluate their growth and distribution of Cd contamination in the different edible parts of the plant. Cd is not beneficial for plant growth and reduces the plant growth and yield of the plant because it tends to accumulate in the shoot of the vegetables grown in Cd-polluted soils. Application of Zn reduces Cd accumulation in plants, and a high concentration of Zn (80 mg kg^{-1}) should be applied in highly Cd-polluted soil. Application of Zn increases the growth and yield of plants and minimizes Cd-accumulation in plants.

ACKNOWLEDGEMENT

Authors are grateful to the contribution of University Grant Commission (UGC, India) for providing the D.Phil. scholarship under UGC scheme for the degree Doctor of Philosophy.

REFERENCES

- Nagajyoti, P.C., Lee, K.D. & Sreekanth, T.V.M. (2010). Heavy metals, occurrence and toxicity for plants: a review. *Environmental Chemistry Letters*, 8:199–216.
- Ali, B., Wang, B., Ali, S., Ghani, M.A., Hayat, M.T., Yang, C., Xu, L. & Zhou, W.J. (2013a). 5-Aminolevulinic acid ameliorates the growth, photosynthetic gas exchange capacity, and ultrastructural changes under cadmium stress in *Brassica napus* L. *Journal Plant Growth Regulation*, 32: 604–614.
- Abbas, T., Rizwan, M., Ali, S., Rehman, M.Z., Qayyum, M.F., Abbas F., Hannan, F., Rinklebe, J. & Ok, Y.S. (2017). Effect of biochar on Cadmium bioavailability and uptake in wheat (*Triticum aestivum* L.) grown in a soil with aged contamination. *Ecotoxicol Environmental Safety*, 140:37–47.
- Hou, D., O'Connor, D., Nathanail, P., Tian, L. & Ma, Y. (2017). Integrated GIS and multivariate statistical analysis for regional scale assessment of heavy metal soil contamination: a critical review. *Environmental Pollution*, 231:1188–1200.
- Ali, B., Huang, C.R., Qi, Z.Y., Ali, S., Daud, M.K., Geng, X.X, Liu, H.B. & Zhou, W.J. (2013b). 5-Aminolevulinic acid ameliorates cadmium-induced morphological, biochemical, and ultra structural changes in seedlings of oilseed rape. *Environmental Science Pollution Research*, 20:7256–7267.
- Rizwan, M., Ali, S., Adrees, M., Rizvi, H., Rehman, M.Z., Hannan, F., Qayyum, M.F., Hafeez, F. & OK, Y.S. (2016a). Cadmium stress in rice: toxic effects, tolerance mechanisms and management: a critical review. *Environmental Science Pollution Research*, 23:17859–17879.
- Harris, N.S. and Taylor, G.J. (2001). Remobilization of Cadmium in maturing shoots of near isogenic lines of durum wheat that differ in grain Cadmium accumulation. *Journal of Experiment Botany*, 52:1473–1481.
- Basnet, P., Amarasiriwardena, D., Wu, F., Fu, Z. & Zhang, T. (2014). Elemental bioimaging of tissue level trace metal distributions in rice seeds (*Oryza sativa* L.) from a mining area in China. *Environmental Pollution*, 195: 148–156.
- Fontes, R.L., Pereira, J. & Neves J.C. (2014). Uptake and translocation of Cd and Zn in two lettuce cultivars. *Anais da Academia Brasileira de Ciências*, 86(2): 907922.
- Martos, S., Gallego, B., Saez, L., Lopez-Alvarado, J., Cabot, C. & Poschenrieder, C. (2016). Characterization of Zinc and Cadmium hyperaccumulation in three *Noccaea* (Brassicaceae) populations' from non-metalliferous sites in the eastern Pyrenees. *Front Plant Science*, 7: 1–13.
- Erdem H., Tosun Y.K. and Ozturk M. (2012). Effect of Cadmium-Zinc interactions on growth and Cd-Zn concentration in durum and bread wheat. *Fresenius Environmental Bulletin*, 21: 1046–1051.
- Baycu G., Gevrek-Kürüm N., Moustaka J., Csatori I., Rognes S.E. and Moustakas M. (2017). Cadmium-zinc accumulation and photosystem II responses of *Noccaea caerulea* to Cd and Zn exposure. *Environmental Science Pollution Research*, 24: 2840–2850.
- Qayyum M.F., Rehman M.Z., Ali S., Rizwan M., Naeem A., Maqsood M.A., Khalid H., Rinklebe J. and Ok Y.S. (2017). Residual effects of mono ammonium phosphate, gypsum and elemental sulfur on cadmium phytoavailability and translocation from soil to wheat in an effluent irrigated field. *Chemosphere* 174:515–523.
- Adimalla, N. (2019). Heavy metals contamination in urban surface soils of Medak province, India, and its risk assessment and spatial distribution. *Environmental Geochemistry and Health*, 42: 59–75.
- Dong, B., Zhang, R., Gan, Y., Cai, L., Freidenreich, A. & Wang, K. (2019). Multiple methods for the identification of heavy metal sources in cropland soils from a resource based region. *Science of the Total Environ*, 651: 3127–3138.
- Sun, L., Guo, D., Liu, K., Meng, H., Zheng, Y. & Yuan, F. (2019). Levels, sources and spatial distribution of heavy metals in soils from a typical coal industrial city of Tangshan, China. *CATENA*, 175: 101–109.
- Zhao, K., Fu, W., Qiu, Q., Ye, Z., Li, Y. & Tunney, H. (2019). Spatial patterns of potentially hazardous metals in paddy soils in a typical electrical waste dismantling area and their pollution characteristics. *Geoderma*, 337: 453–462.
- Liu, L., Li, W., Song, W. & Guo, M. (2018). Remediation techniques for heavy metal-contaminated soils: Principles and applicability. *Science of the Total Environment*, 633: 206–219.
- Bi X., Feng X., Yang Y., Qiu G., Li G., Li F., Liu T., Fu Z. & Jin Z. (2006). Environmental contamination of heavy metals from Zinc smelting areas in Hezhang County, western Guizhou, China. *Environmental international*, 32(7): 883–890.
- Khan S., Aijun L., Zhang S., Hu Q. & Zhu Y.G. (2008a). Accumulation of polycyclic aromatic hydrocarbons and heavy metals in lettuce grown in the soils contaminated with long-term wastewater irrigation. *Journal of Hazardous Materials*, 152 (2): 506–515.
- Li W., Khan, M.A., Yamaguchi, S. & Kamiya, Y. (2005a). Effects of heavy metals on seed germination and early seedling growth of *Arabidopsis thaliana*. *Plant Growth Regulation*, 46: 45–50.
- Lim, H.S., Lee, J.S., Chon, H.T. & Sager, M. (2008). Heavy metal contamination and health risk assessment in the vicinity of the abandoned Songcheon Au–Ag mine in Korea. *Journal of geochemical Exploration*, 96: 223–230.
- Pruvot, C., Douay, F., Herve, F. & Waterlot, C. (2006). Heavy metals in soil, crops and grass as a source of human exposure in the former mining areas. *Journal of Soils Sediments*, 215–220.
- Radwan, M.A. & Salama, A.K. (2006). Market basket survey for some heavy metals in Egyptian fruits and vegetables. *Food and Chemical Toxicology*, 44(8): 1273–1278.
- Ramos, I., Esteban, E., Lucena, J.J. & Garate, A. (2002). Cadmium uptake and sub cellular distribution in plants of *Lactuca* sp. Cd-Mn interaction. *Plant Science*, 162(5): 761–767.
- Intawongse, M. & Dean, J.R. (2006). Uptake of heavy metals by vegetable plants grown on contaminated soil and their bioavailability in the human gastrointestinal tract. *Food Addit. Contam.*, 23(1):36–48.

27. McBride M.B. (2003). Cadmium concentration limits in agricultural soils: weaknesses in USEPA's risk assessment and the 503 rule. *Human and Ecological Risk Assessment: An International Journal*,9(3):661–674.
28. Monteiro, M.S., Santos, C., Soares, A.M. & Mann, R.M. (2009). Assessment of biomarkers of cadmium stress in lettuce. *Ecotoxicol Environmental Safety*, 72:811–819.
29. Verma S. & Dubey R. (2003). Lead toxicity induces lipid peroxidation and alters the activities of antioxidant enzymes in growing rice plants. *Plant Science*, 164(4):645–655.
30. Chaves, L.H.G., Estrela, M.A. & Sena de Souza, R. (2011). Effect on plant growth and heavy metal accumulation by sunflower. *Journal of Phytology*,3(12):04–09.
31. Chopra, S.L. & Kanwar, J.S. (1999). *Analytical agricultural chemistry*. Kalyani Publication, New Delhi.
32. Lindsay, W.L. & Norvell, W.A. (1978). Development of DTPA soil test for zinc, iron, manganese and copper. *Soil Science Society of American Journal*, 42(3):421–428.
33. Kumar, C. & Mani, D. (2010). *Enrichment and management of heavy metals in sewage-irrigated soil*. Lap LAMBERT Academic Publishing, Dudweilerjermany.
34. Lopez-Millan, A.F., Sagardoy, R., Solanas, M., Abadía, A. & Abadía, J. (2009). Cadmium toxicity in tomato (*Lycopersicon esculentum*) plants grown in hydroponics. *Environmental and Experimental Botany*, 65:376–385.
35. Sanita, D.I. Toppi, L. & Gabbrielli, R. (1999). Response to cadmium in higher plants. *Environmental and Experimental Botany*, 41:105–130.
36. Wojcik, M. & Tukiendorf, A. (2004). Phytochelatin synthesis and cadmium localization in wild type of *Arabidopsis thaliana*. *Plant Growth Regulation*, 44:71–80.
37. Mohanpuria, P., Rana, N.K. & Yadav, S.K. (2007). Cadmium induced oxidative stress influence on glutathione metabolic genes of *Camella sinensis*(L.). O Kuntze. *Environmental Toxicology*,229(4):368–374.
38. Guo, J, Dai, X., Xu, W. & Ma, M. (2008). Over expressing GSH1 and AsPCSI simultaneously increases the tolerance and accumulation of cadmium and arsenic in *Arabidopsis thaliana*. *Chemosphere*,72(7):1020–1026.
39. Green, C.E., Chaney, R.L. & Bouwkamp, J. (2003). Interactions between cadmium uptake and phytotoxic levels of zinc in hard red spring wheat. *Journal of Plant Nutrition*, 26(2):417–430.
40. Zare, A.A., Khoshgoftarmanesh, A.H., Malakouti M.J., Bahrami, H.A. & Chaney, R.L. (2018). Root uptake and shoot accumulation of cadmium by lettuce at various Cd: Zn ratios in nutrient solution. *Ecotoxicology and Environmental Safety*, 148:441–446.
41. Mysliwa-Kurziel B., Prasad M.N.V. & Stralka K. (2004). Photosynthesis in heavy metals stress plants, In: Prasad MNV (ed) *Heavy metal stress in plants*, 3rd edition. Springer, Berlin, 182-200.

CITATION OF THIS ARTICLE

B Singh, D Mani, V Sahu, A Bhushan, P Kumar, D P Shukla and H Vishwakarma. Cadmium–Zinc Interaction in Relation to Lettuce Growth. *Bull. Env. Pharmacol. Life Sci.*, Vol10[6] May 2021 : 89-94