



Media Optimization for the Bacteriocin Production Of *Lactobacillus plantarum* BLN39 against *Mycobacterium fortuitum*

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ABSTRACT

Lactic Acid Bacteria (LAB) is a diverse group of gram positive bacteria which grabs the researchers attention in recent times due to their Generally Recognized As Safe (GRAS) status. Bacteriocins of LAB found numerous applications in food and dairy industries as they are pH and heat stable, easy to produce and retains their activity after storage. Bacteriocin production by LAB highly depends on the culture condition, physicochemical properties, media composition, etc., In media composition, carbon and nitrogen sources, tween concentration are essential for the bacteriocin production. Most commonly, de Man Rogosa Sharpe (MRS) broth is the medium of choice for the LAB fermentation process. However, this medium is quite expensive and provides hindrance to the large scale economical production of bacteriocin from LAB. There is always a necessity to find an alternative and cheapest source of carbon and nitrogen source and optimization of media composition for the effective bacteriocin production. *Mycobacterium fortuitum* is one of the rapidly growing non tuberculous mycobacteria which causes diseases in humans includes pulmonary diseases, skin, joints and bone infections and disseminated diseases, etc., The treatment for *M. fortuitum* disease involves multiple antibiotics, long course treatment and drug related toxicities. This necessitates the development of new drug candidates against *M. fortuitum*. In this study, eight different media were prepared by supplementing and replacing the existing carbon and nitrogen source with molasses and saw dust and with 2X composition of existing carbon source and tween 80. *Lactobacillus plantarum* BLN39 was subjected to grown in this different optimized media and their arbitrary unit (AU/ml) was calculated by screening *L. plantarum* BLN39 against *M. fortuitum*. Among eight different media optimized, MRS broth supplemented with molasses showed 1600 AU/ml which is double the AU/ml obtained for neat MRS broth (800 AU/ml). MRS broth supplemented with saw dust and MRS broth with 2X tween 80 showed the same AU/ml as of neat MRS broth. Hence, molasses proved as an effective and cheapest supplement for the bacteriocin production of *L. plantarum* BLN39 against *M. fortuitum*.

Keywords: *Lactobacillus plantarum*, *Mycobacterium fortuitum*, Non tuberculous mycobacteria, media optimization, molasses.

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INTRODUCTION

Lactic Acid Bacteria is a diverse group of microorganisms which is gram positive, catalase negative bacteria which exists in either cocci or bacilli in shape. LAB grabs a significant attention towards researchers and in biopreservation techniques due to their Generally Recognized As Safe (GRAS) status [6]. LAB could inhibit the growth of other virulent microorganisms by production of bacteriocins. Bacteriocins is an antimicrobial peptide, ribosomally synthesized, can be produced by many other bacteria [18].

Bacteriocins of LAB have numerous biotechnology applications as they are easy to produce, stable at low pH, non toxic to humans, sensitive to proteases, heat stable and they are able to retain their activity after long term of storage, etc., [20, 21, 4]. Bacteriocin production can be influenced by incubation atmosphere, physicochemical conditions, etc. and also by various medium composition [9, 21-24]. The most important parameters in the medium composition on bacteriocin production are the concentration of carbon source, nitrogen source and tween 80 [1, 2, 14, 15]. Most commonly, de Man Rogosa Sharpe (MRS) medium is the choice of medium for LAB fermentations but it has its own limitations [1]. It is quite expensive and acts as barrier for economical large scale of production. Few studies focused on the

cheaper source of carbon and nitrogen for LAB fermentations and their bacteriocin production [22, 19, 11].

M. fortuitum is a clinically significant rapidly growing mycobacteria which causes mostly pulmonary diseases and also extrapulmonary disease, causing localized skin, soft tissue, wound or bone-infections following traumatic injuries or surgery. The treatment for *M. fortuitum* involved with long course, combination of antibiotics and their drug related toxicities necessitates the need for the development of new candidates against *M. fortuitum* [5, 8, 9, 13, 2].

In this study, eight different media has been prepared for the growth of LAB strain BLN39 which includes neat MRS broth, MRS broth supplemented with cheaper carbon sources like molasses and saw dust, MRS broth in which dextrose replaced by molasses and saw dust, MRS broth with 2X carbon and nitrogen source and Tween 80. BLN39 was subjected to grow in this media and screened their bacteriocin production by screening against *M. fortuitum*.

MATERIAL AND METHODS

Chemicals and Cultures:

All the chemicals and reagents used in this study were purchased from Himedia (Mumbai). The indicator organism used in this research work, *Mycobacterium fortuitum* (MTCC1902) was obtained from Microbial Type Culture Collection, Chandigarh, India.

Media Preparation:

Eight different media for the growth and bacteriocin production of BLN39 was prepared as mentioned in Table 1. The pH of all the prepared media was adjusted to pH 6.5 and autoclaved at 121°C for 15 minutes.

Bacteriocin Production:

An overnight grown culture of BLN39 was inoculated into the prepared eight different media and incubated in shaking condition (100rpm) at 30°C for 18 hours. After incubation, the culture was centrifuged at 5000rpm for 10 mins and the cell free supernatant (CFS) was collected to assess the bacteriocin production of BLN39. Then the collected CFS from each prepared media was serially diluted in two fold dilution to measure the arbitrary units per ml (AU/ml) of each optimized media.

Anti *M. fortuitum* activity:

Anti *M. fortuitum* activity of BLN39 with different optimized media was assessed using agar well diffusion method. Briefly, the suspension of an indicator organism, *M. fortuitum* (MTCC1902) was prepared by inoculating a loopful of colonies into 0.3ml of middlebrook 7H9 broth in bijou bottle containing sterile glass beads. The colonies were homogenized by vortex for 30 seconds and kept undisturbed to allow the clumps to settle down. Then the volume of the suspension was made to 5ml using middlebrook 7H9 broth. 200µl of prepared suspension was added to 5ml of molten agar and poured onto middlebrook 7H9 agar plate. Subsequently, the wells were made on the surface of the agar plate. 100µl of the CFS collected and serially diluted was added into the well. Then the plates were incubated at 37°C for 24 hours and the zone of inhibition was measured to calculate AU/ml.

Taxonomy of the strain BLN39:

The genomic DNA of the LAB strain BLN39 was isolated using solute ready genomic DNA kit. DNA was analyzed by gel electrophoresis and quantified using spectrophotometer (NanoDrop ND-1000, Thermo Scientific, Gloucester, UK). The 16S rRNA gene sequence of the strain was amplified using the primers: 27F 5'AGAGTTTGATCMTGGCTCAG3' (forward) and 1492R 5'TACGGYTACCTTGTTACGACTT3' (reverse) (Kumar Gothwal *et al.*, 2007). The PCR amplified product of the strain was sequenced and analyzed at National Collection of Industrial Microorganisms (NCIM), National Chemical Laboratory (CSIR-NCL), Pune, India. The partial 16S rRNA nucleotide sequence of BLN39 has been deposited in GenBank database.

RESULTS AND DISCUSSION:

Anti *M. fortuitum* activity:

BLN39 exhibited different level of anti *M. fortuitum* activity in the different optimized media (Figure 1 & 2). MRS broth supplemented with molasses showed 1600 AU/ml which is double the AU/ml obtained for neat MRS broth (800 AU/ml). Molasses was used as an efficient and cheap carbon source for large scale production of various microbial strains [3, 17]. This study supported the result obtained by Sridevi *et al.*, [19] in which molasses were proved as effective and cheaper source of carbon for the bacteriocin production by *Lactobacillus plantarum* sp., A study by Mulyani *et al* [16] also showed that molasses can act as low cost component for effective bacteriocin production by *Pediococcus pentosaceus*. MRS broth supplemented with saw dust and MRS broth with 2X tween 80 showed the same AU/ml as of neat MRS broth. MRS broth in which saw dust replaced for dextrose showed the least inhibitory activity i.e, 100 AU/ml. A study by Hoda *et al.*, [12] have showed that MRS broth supplemented with Tween 80 along with

other components resulted in high bacteriocin production by *Lactobacillus acidophilus*. A study by Todorov and Dicks [20] have proved that inclusion of tween 80 increased bacteriocin production by 50% in *Enterococcus mundtii*. Elvina *et al.*, showed that tween supply is essential for the bacteriocin activity of *L. plantarum* [7]. Therefore, molasses act as good carbon supplementary source for the growth and bacteriocin production of BLN39.

Taxonomy of the strain BLN39:

Amplification of 16S rRNA gene from the strain BLN39 resulted in 1455bp sequences. BLAST analysis showed 99.79% sequence similarity with *Lactobacillus plantarum* strain DSM 16365. Hence the strain BLN39 was identified as *Lactobacillus plantarum*.

Table 1: Media Preparation for bacteriocin production

Media	Components	w/v
Media 1	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Dextrose	20g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
Media 2	Proteose Peptone	20g/l
	Beef extract	20g/l
	Yeast extract	10g/l
	Dextrose	40g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
Media 3	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Dextrose	20g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
Media 4	Molasses	20g/l
	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
Media 5	Molasses	20g/l
	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Dextrose	20g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
Magnesium sulphate	0.1g/l	

	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
	Saw dust	20g/l
Media 6	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	8g/l
	Saw dust	20g/l
Media 7	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Dextrose	20g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
Media 8	Proteose Peptone	10g/l
	Beef extract	10g/l
	Yeast extract	5g/l
	Dextrose	20g/l
	Ammonium citrate	2g/l
	Sodium acetate	5g/l
	Magnesium sulphate	0.1g/l
	Manganese sulphate	0.05g/l
	Dipotassium phosphate	2g/l
	Tween 80	16g/l

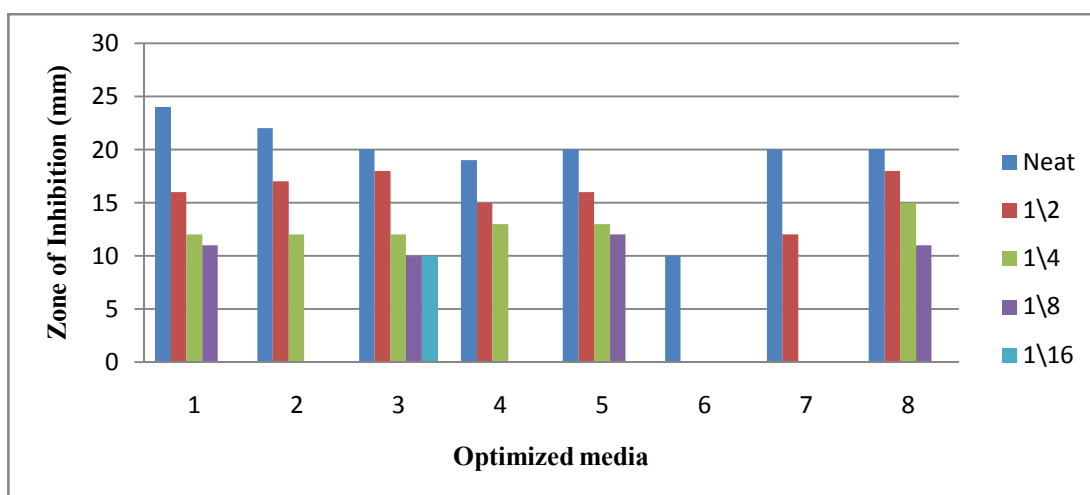


Figure 1: Anti *M. fortuitum* activity of optimized media

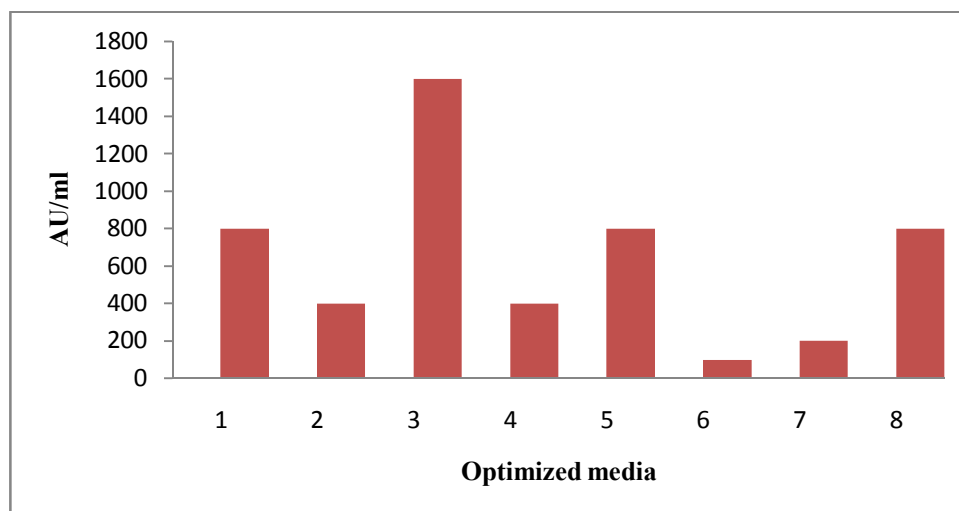


Figure 2: AU/ml evaluation of optimized media

CONCLUSION

Lactobacillus plantarum BLN39 showed better bacteriocin production against *M. fortuitum* MTCC1902 in the MRS broth medium supplemented with molasses. Molasses proved to be an effective and cheapest carbon source supplement for the bacteriocin production of *L. plantarum*.

REFERENCES

1. Abo-Amer. (2011). Optimization of bacteriocin production by *Lactobacillus acidophilus* AA11, a strain isolated from Egyptian cheese. *Ann Microbiol*, :445-452
2. Arakawa, K.; Kawai, Y.; Fujitani, K.; Nishimura, J.; Kitizawa, H.; Komine, K.; Kai, K.; Saito, T. (2008). Bacteriocin production of probiotic *Lactobacillus gasseri* LA39 isolated from human feces in milk-based media. *Anim. Sci. J.* 79, 634-640.
3. Barbosa HS, de Silveira EA, Miranda M, Ernandes JR (2016). Efficient very-highgravity fermentation of sugarcane molasses by industrial yeast strains. *J Inst Brew* 122:329-333
4. Bari ML, Ukuku DO, Kawasaki T, Inatsu Y, Isshiki K, Kawamoto S (2005). Combined efficacy of nisin and pediocin with sodium lactate, citric acid, phytic acid, and potassium sorbate and EDTA in reducing the *Listeria monocytogenes* population of inoculated fresh-cut produce. *J Food Prot* 68(7):1381-1387
5. Brown-Elliott BA, Wallace RJ Jr. (2002). Clinical and taxonomic status of pathogenic nonpigmented or late-pigmenting rapidly growing mycobacteria. *Clin Microbiol Rev.* 15(4):716-46.
6. Carr FJ, Chill D, Maida N (2002). The lactic acid bacteria: a literature survey. *Crit Rev Microbiol* 28(4):281-370
7. Elvina Parlidungan, Chaitali Dekiwadia, Oliver A.H. Jones. (2021). Factors that influence growth and bacteriocin production in *Lactiplantibacillus plantarum* B21. *Process Biochemistry*, 107 (8), 20-23.
8. G. Sathiyarayanan . R. Gandhimathi, B. Sabarathnam, G. Seghal Kiran, Joseph Selvin (2008). Optimization and production of pyrrolidone antimicrobial agent from marine sponge-associated *Streptomyces* sp. *MAPS15. Bioprocess Biosyst Eng.*
9. Ganzle MG, Weber S, Hammes WP: Effect of ecological factors on the inhibitory spectrum and activity of bacteriocins. *Int J Food Microbiol* 1999, 46:207-217.
10. Griffith DE, Aksamit T, Brown-Elliott BA, Catanzaro A, Daley C, Gordin F, Holland SM, Horsburgh R, Huitt G, Iademarco MF, et al. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med.* 2007;175(4):367-416.
11. Han B, Zhang R, Yu Z, Liu B and Ma Q. 2011. Optimization of bacteriocin production by *Lactobacillus plantarum* YJG isolated from the mucosa of the gut of healthy chickens. *African J Microbiol Res*, 5: 1147-1155.
12. Hoda Mahrous, Abeer Mohamed, M. Abd El-Mongy, A. I. El-Batal, H. A. Hamza. Study Bacteriocin Production and Optimization Using New Isolates of *Lactobacillus* spp. Isolated from Some Dairy Products under Different Culture Conditions. *Food and Nutrition Sciences*, 2013, 4, 342-356
13. Johanna Erber, Simon Weidlich, Tristan Tschaikowsky, Kathrin Rothe, Roland M. Schmid, Jochen Schneider and Christoph D. Spinne. Successful bedaquiline-containing antimycobacterial treatment in posttraumatic skin and soft-tissue infection by *Mycobacterium fortuitum* complex: a case report. *BMC Infectious Diseases* (2020) 20:365
14. Keren T, Yarmus M, Halevy G, Shapira R: (2004). Immunodetection of the bacteriocin lactacin RM: analysis of the influence of temperature and Tween 80 on its expression and activity. *Appl Environ Microbiol*, 70:2098-2104.
15. Mataragas M, Drosinos EH, Tsakalidou E, Metaxopoulos E:(2004). Influence of nutrients on growth and bacteriocin production by *Leuconostoc mesenteroides* L124 and *Lactobacillus curvatus* L442. *Antonie Van Leeuwenhoek*, 85:191-198.
16. Mulyani, S., Jenie, B. S. L., Kusumaningrum, H. D. and Arief, I. I. (2019). Characterisation of crude bacteriocin produced by *Pediococcus pentosaceus* 2A2 in enriched molasses medium. *International Food Research Journal* 26(1): 187 - 192.

17. Okafor N, Okeke BC (2017). Nutrient media for cultivation of industrial microorganisms and generation of microbial products. *Modern industrial microbiology and biotechnology*, 2nd edn. CRC Press, Taylor and Francis Group, pp 43–60
18. Sahar Abbasiliasi, Joo Shun Tan, Tengku Azmi Tengku Ibrahim, Fatemeh Bashokouh, Nagasundara Ramanan Ramakrishnan, Shuhaimi Mustafa and Arbakariya B. Ariff. (2017). Fermentation factors influencing the production of bacteriocins by lactic acid bacteria: a review. *RSC Adv.*, 2017, 7, 29395-29420
19. Sridevi Venigalla, Yasarapu N. Sindhuja, Kancharana Srujana, Silarapu Swathi, Yerri Naidu and Garapati H. Rao (2011). Optimized Production of Bacteriocin from Cheaper Carbon and Nitrogen Sources Using Response Surface Methodology. *Research Journal of Microbiology*. 12: 42-49
20. Todorov SD, Dicks LMT (2009). Effect of modified MRS medium on production and purification of antimicrobial peptide ST4SA produced by *Enterococcus mundtii*. *Anaerobe* 15(3):65–73
21. Turgis M, Vu KD, Millette M, Dupont C, Lacroix M (2016) Influence of environmental factors on bacteriocin production by human isolates of *Lactococcus lactis* MM19 and *Pediococcus acidilactici* MM33. *Probiotics Antimicrob Proteins* 8(1):53–59
22. Verluyten J, Leroy F, de Vuyst L, (2004). Influence of complex nutrient source on growth of and curvacin a production by sausage isolate *Lactobacillus curvatus* LTH1174. *Applied Environmental Microbiology*, 70: 5081-5088.
23. Zhang J, Zhang Y, Liu SN, Ye H, Zhou ZJ (2012) Modelling growth and bacteriocin production by *Pediococcus acidilactici* PA003 as a function of temperature and pH value. *Appl Biochem Biotechnol* 166(6):1388–1400
24. Zhou K, Zeng YT, Han XF, Liu SL (2015) Modelling growth and bacteriocin production by *Lactobacillus plantarum* BC-25 in response to temperature and pH in batch fermentation. *Appl Biochem Biotechnol* 176(6):1627–1637

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