



Antimicrobial activities of scorpion and honey bee venom against some common selected pathogen

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ABSTRACT

Multidrug resistant pathogens are reported in high numbers in the world. This cause failure of the currently available antibiotics to treat these multidrug resistant pathogens. This lead to the global concern to discover alternative to antibiotics. Various researchers have conducted many studies about the biological activities of bee venom and scorpion venom. In the literature it is reported that the venom of bee and scorpion have some peptide which have antimicrobial activity. Our study was therefore conducted to determine the antimicrobial property of bee venom and scorpion venom against some common selected pathogens. Bee venom and scorpion venom antimicrobial activity was tested against common selected pathogenic bacteria and fungi that includes Staphylococcus aureus, Salmonella typhimurium, Escherichia coli, Pseudomonas aeruginosa, Candida albicans, Trichophyton mentagrophytes and Trichophyton rubrum. Disc diffusion method was used to evaluate the antimicrobial activity of these venom. Standard antibiotics and antifungal that includes Chloramphenicol, streptomycin and penicillin, fluconazole and amphotericin B were used as control for antimicrobial activity. Against all selected common pathogenic bacteria and fungi both of the venom of bee and scorpion shows antibacterial and antifungal activity. The bee venom give zone of inhibition 25mm, 18mm, 21mm against E. coli, S. aureus, and Salmonella typhimurium and Pseudomonas aeruginosa respectively while the scorpion venom gives 23mm, 16mm, 21mm and 17mm zone of inhibition against these bacteria respectively. While in case of fungi the zone of inhibition of bee venoms against C. albicans T. mentagrophytes and T. rubrum was 18mm, 20mm and 17mm while scorpion venom gives 17mm, 21mm and 15mm zone of inhibition against these fungi respectively. Our study shows that both bee venom and scorpion venom have the ability to inhibit bacteria and fungi which may be used complementary to antibiotics. Therefore, our study might conclude that specific mechanism, which is not well known, is used by bee venom and scorpion venom to inhibit the growth of both bacteria and fungi. Further in vivo and in vitro study based on a chemical, pharmacological, and clinical approach must be conducted to understand the exact mechanism of these venoms.

Key words: Antibacterial; Antifungal; scorpion venom; honey bee venom; inhibition zone

Received 12.07.2020

Revised 22.08.2020

Accepted 03.09.2020

INTRODUCTION

Currently the conventional available antibiotics are not working properly because most of the pathogenic microbes are reported to be antibiotic resistant. 70% of the hospital acquired infection causing bacteria have been reported to be resistant to one or more available antimicrobial agents. However some bacterial

strains are reported to be multi drug resistant while some of the bacterial strains are reported to be resistant to all available conventional antibiotics [1].

Due to drug resistance the global public health is at high risk and the risk is increasing with the time because the drug resistant pathogens are emerging continuously [2, 3]. The most threatening pathogenic bacteria for public health is methicillin-resistant *Staphylococcus aureus* (MRSA). This is because of increased death rate and cost in treating this multidrug resistant bacteria [4, 5]. Currently there is a need to develop new antimicrobial agents to decrease the threat of drug resistant pathogen to the public health [6].

Pharmaceutical products that are available commercially, mostly are direct or indirect derived product of microbes, animals and plants of both terrestrial and marine origin [7]. Generally, it is highly studied and reviewed to use plants for medicinal purposes in previous studies. On the other hand, the same attention has not been paid to the animals to be used for medicinal purposes as insects has the potential to be used for this purpose. By comparing the research per species between plants and insects, it has been observed that plants chemicals have been 7000 time more studied as compared to chemicals present in insects. Currently the attention of the researchers are attracted highly towards insects to contribute in novel discoveries [8, 9].

Evolutionarily Antimicrobial peptides are considered as prehistoric weapons. As the antimicrobial peptides are distributed widely all over the animal kingdom. This distribution suggested their role in complex multicellular organism's evolution successfully [10]. From antimicrobial secretions and venoms numerous antimicrobial peptides have been isolated. Numerous peptides have been derived from the scorpion's venom. These peptides are reported to have antibacterial and antifungal activity in a similar manner like broad spectrum antibiotics [11]. Against a large number of gram-positive bacteria, pandinin 1 and 2 of scorpion venom are reported to have strong antimicrobial activity [12]. Additionally, venom of the scorpion CsTX also shows antimicrobial activity [13]. Other than venom of scorpions of, the bee venom (*Apis mellifera*) also shows antimicrobial activity [14]. Mellitin component of the bee venom shows more potent antimicrobial activity against gram-positive bacteria as compared to gram negative bacteria. Moreover, in addition to wasps venom, honey bees and many snakes have antimicrobial peptides, however no investigation have been done on the functions of these peptides [15].

Bees belonging to the species *Apis mellifera* species of bee have many activities that are similar to human being. Traditionally these includes; pollination, honey production, resins, wax, jelly, pollen, and venom like apitoxin. Compounds synthesized by bees synthesized many compounds that have been widely studied because these compounds have many application therapeutically [16, 17, 18, 19]. Bee produce many substances, among these the most important substance is apitoxin. This complex chemical is synthesized by the gland located in the abdomen of these insects. Apitoxin of bee venom have 88% water content while 12% comprises of many components like phospholipase A2, hyaluronidase, melittin, histamine. Additionally it contains peptides such as apamin, secapin etc. [20]. Regarding the components of apitoxin, the highly studied compound is phospholipase A2. Samel et al. done their study in which they shows that *sn-2* fatty acyl ester bond of *sn-3* phosphoglycerides hydrolysis is catalyzed by phospholipases A2 due to which they give free lysophospholipids and fatty acids [21]. Phospholipases A2 protein have been found in numerous tissues of mammals and arthropods. It has been found in snakes, scorpions and bee venom. This constitute a large family of protein in these [22]. Among these 10 groups are of secretory phospholipase A2 [23, 24]. Molecular weight of Phospholipase A2 is low. They have high potential of immunogenicity and their catalytic activity is also high. Phospholipase A2 enzyme shows antibacterial and anticoagulant activity and shows vigorous role in chemical mediator's generation, proliferation of cell, contraction of muscle [25, 26]. Vital component of apitoxin is melittin. It contains 26 amino acids having amphipathic character. These amino acid chain let melittin interaction with lipid membranes. This also increase the erythrocytes and other membrane. About 50% of the bee apitoxin belong to species *Apis mellifera* are constituted by these amino acids [20]. Cytotoxic activity has been observed for melittin and it has potential activity of cell lysis and its cell lysis activity has been evidenced in human cell lysis of erythrocytes [27]. Moreover, cell membrane is directly acted upon by it [28, 29]. Melittin have numerous biological activities that includes activity against microbes like bacteria, fungi, viruses. They also have anti-inflammatory activity, inhibitory effect on cell growth and different cancer cell line apoptosis [30, 31].

Scorpions are considered as one of the utmost prehistoric animals living on earth. They have lived over 400 million years [32]. This old evolutionary changes attribute mainly to develop weapon of efficient venom that will support their requirement to prey and their defense. In the whole globe they have wide distribution and they have about 1500 species [33]. The venomous glands of the scorpions comprises of large number of biologically active molecules such as lipids, nucleotides, biogenic amines, enzymes and other molecules that are unknown [34, 35]. Beside these it also comprises of numerous peptides having

multiple activities biologically. These peptides are considered to be the main component of an innate immune system that give protection to the scorpion against various pathogens [36, 37]. A less abundant group of peptides called non-disulfide bridged peptides, having no disulfide bridges, have currently achieved great interest. They have many biological activities such as anticancer activity, hemolytic activity, activity against inflammation and immune-modulatory effect. Beside these they also have activity against microbes (38, 39, 40). By keeping in mind the various biological activities of bee venom and scorpion venom reported in literature, we piloted our study to evaluate the antibacterial potential of the scorpion and honey bee venom available commercially against common selected pathogenic bacteria and fungi.

MATERIAL AND METHODS

Commercialized sources of venom were collected in lyophilized form in the department of zoology, Kohat University of science and technology, Khyber Pakhtunkhwa, Pakistan. Sterile condition were strictly adopted during collection of venoms and at 4°C they were centrifuged and after six hours of extraction they were frozen and lyophilized. Packing and storage of the venom was taken in the dark at 20°C. These information were taken from the leaflet provided with the commercial venom. Standard antibiotics and antifungal that includes Chloramphenicol, streptomycin and penicillin, fluconazole and amphotericin B were used as control for antimicrobial activity. MuellerHinton (MH) agar medium was used to check the antimicrobial activity of both bee and scorpion venom. *Staphylococcus aureus*, *Salmonella typhimurium*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Candida albicans*, *Trichophyton mentagrophytes* and *Trichophyton rubrum* were used in our study to determine the antimicrobial activity of the bee and scorpion venom. The bacteria were picked up from the research laboratory in the department of microbiology, Kohat University of science and technology, Khyber Pakhtunkhwa, Pakistan, in which these cultures were preserved at -80°C. The bacteria and fungi with the ATCC number are given in table 1. The Muller Hinton agar prepared and then autoclaved. Then 20ml of it was poured in 90mm plate. 100µg lyophilized crude venoms were dissolved in 1 ml of buffer (Tris-HCl) and then it was filtered with syringe filter having pore size of 0.22µm. After that it was stored for further use at 4°C. Disc-diffusion method was used to determine the antimicrobial activity as done by Bauer et al (41). Sterile cotton swab was used to spread inoculums of the bacteria on the MH agar plates. For about three minutes the medium surface was allowed to dry. On the surface of the MH agar 7mm (Diameter) sterile paper discs were placed. Then 20µl samples of both the venom having concentration of 100µg/ml were added in the discs. Then at 37°C the plate incubation was done for 24 hours. Finally the zone of inhibition was measured by using guidelines of NCCLS (2002) (42). To determine the antifungal activity of bee venom and scorpion venom the fungal ATCC cultures collected from microbiology department, Kohat University of science and technology, Khyber Pakhtunkhwa, Pakistan were first cultured on sabouraud dextrose agar plates, and then it was incubated at 35°C for 48 hours. To make stock solution amphotericin B and fluconazole were dissolved in 2% dimethyl sulfoxide. Then it was diluted for further use. Mueller-Hinton agar glucose methylene blue medium was used for disk diffusion method to determine the antifungal activity of scorpion and bee venom. Sterile cotton swab was used to spread inoculums of the fungi on the MH-GMB agar plates. For about three minutes the medium surface was allowed to dry. On the surface of the MH agar 7mm (Diameter) sterile paper discs were placed. Then 20µl samples of both the venom having concentration of 100µg/ml were added in the discs. Then at 35°C the plate incubation was done for 24 hours. Finally the zone of inhibition was measured. All the data was analyzed statistically.

Table 1: ATCC number of bacteria and fungi used in our study

Serial NO	Bacteria/ Fungi	ATCC No
1.	<i>S. aureus</i>	25923
2.	<i>Pseudomonas aeruginosa</i>	27853
3.	<i>E. coli</i>	25923
4.	<i>Salmonella typhimurium</i>	25923
5.	<i>Candida albicans</i>	10231
6.	<i>T. rubrum</i>	28188
7.	<i>T. mentagrophytes</i>	18748

RESULTS

The antimicrobial activity of honeybee venom and scorpion venom was checked against common pathogenic bacteria including *Staphylococcus aureus* (*S. aureus*), *Salmonella typhimurium*, *Escherichia coli*, and *Pseudomonas aeruginosa*. The antimicrobial activity of these venoms was also checked against pathogenic fungi *C. albicans*, *T. mentagrophytes* and *T. rubrum*. Disc diffusion method was used to evaluate

the antimicrobial activity of these venom. Standard antibiotics and antifungal that includes Chloramphenicol, streptomycin and penicillin, fluconazole and amphotericin B were used as control for antimicrobial activity. Against all selected common pathogenic bacteria and fungi both of the venom of bee and scorpion shows antibacterial and antifungal activity. Both bee venom and scorpion venom was observed to have a substantial antibacterial effect against *E. coli*, *S. aureus*, and *Salmonella typhimurium* and *Pseudomonas aeruginosa*. The bee venom give zone of inhibition 25mm, 18mm, 21mm 15mm against *E. coli*, *S. aureus*, and *Salmonella typhimurium* and *Pseudomonas aeruginosa* respectively (Table 2) while the scorpion venom gives 23mm, 16mm, 21mm and 17mm zone of inhibition against *E. coli*, *S. aureus*, and *Salmonella typhimurium* and *Pseudomonas aeruginosa* respectively. (Table 3)

Figure 1 and figure 2 shows zone of inhibition of scorpion and bee venom against bacteria and fungi. While in case of fungi the zone of inhibition of bee venoms against *C. albicans*, *T. mentagrophytes* and *T. rubrum* was 18mm, 20mm and 17mm. (Table 4) while scorpion venom gives 17mm, 21mm and 15mm zone of inhibition against *C. albicans*, *T. mentagrophytes* and *T. rubrum* respectively. (Table 5) Figure 3 shows comparative zone of inhibition between bee venom scorpion venom against selected pathogens.

Table 2: Bee venom zone of inhibition against selected bacteria

Serial No	Common name	Scientific name	Bacteria /Zone of inhibition			
			<i>Salmonella typhimurium</i>	<i>E.coli</i>	<i>S. Aureus</i>	<i>Pseudomonas aeruginosa</i>
1.	Bee venom	<i>Apis mellifera</i>	21	25	18	15
2.	Antibiotic					
	Chloramphenicol (CHL)	30 µg/ disc	23	20	20	18
	Streptomycin (STR)	10 µg/ disc	25	22	20	18
	Penicillin (P)	10 µg/ disc	20	18	16	16

Table 3: Scorpion venom zone of inhibition against selected bacteria

Serial No	Common name	Scientific name	Bacteria /Zone of inhibition			
			<i>Salmonella typhimurium</i>	<i>E.coli</i>	<i>S. Aureus</i>	<i>Pseudomonas aeruginosa</i>
1.	Scorpion	<i>Buthotus hottentota</i>	21	23	16	17
2.	Antibiotic					
	Chloramphenicol (CHL)	30 µg/ disc	23	20	20	18
	Streptomycin (STR)	10 µg/ disc	25	22	20	18
	Penicillin (P)	10 µg/ disc	20	18	16	16

Table 4: Bee venom zone of inhibition against selected fungi

Serial No	Common name	Scientific name	Fungi /Zone of inhibition		
			<i>C. albicans</i>	<i>T. mentagrophytes</i>	<i>T. rubrum</i>
1.	Bee venom	<i>Apis mellifera</i>	18	20	17
2.	Antibiotic				
	fluconazole	100mg per disc	20	22	20
	amphotericin B	100mg per disc	23	21	22

Table 5: Scorpion venom zone of inhibition against selected fungi

Serial No	Common name	Scientific name	Fungi /Zone of inhibition		
			<i>C. albicans</i>	<i>T. mentagrophytes</i>	<i>T. rubrum</i>
1.	Scorpion	<i>Buthotus hottentota</i>	17	21	15
2.	Antibiotic		20	22	20
	Fluconazole	100mg per disc	23	21	22
	Amphotericin B	100mg per disc	20	22	20

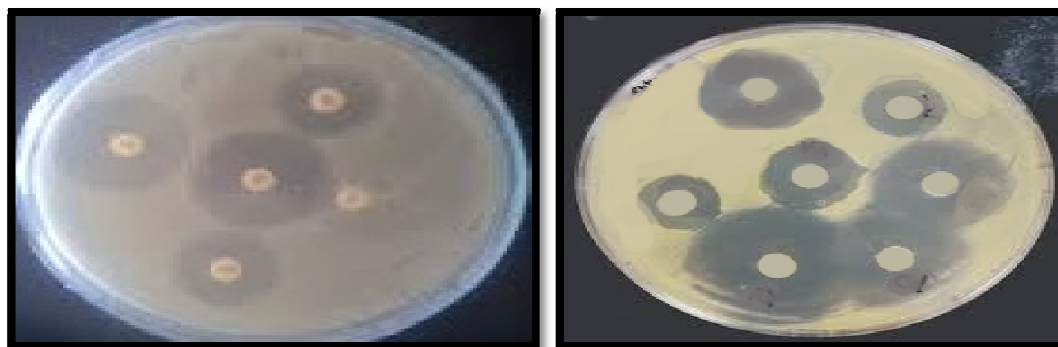


Figure 1: Zone of inhibition against selected pathogen **Figure 2:** Zone of inhibition against selected pathogen

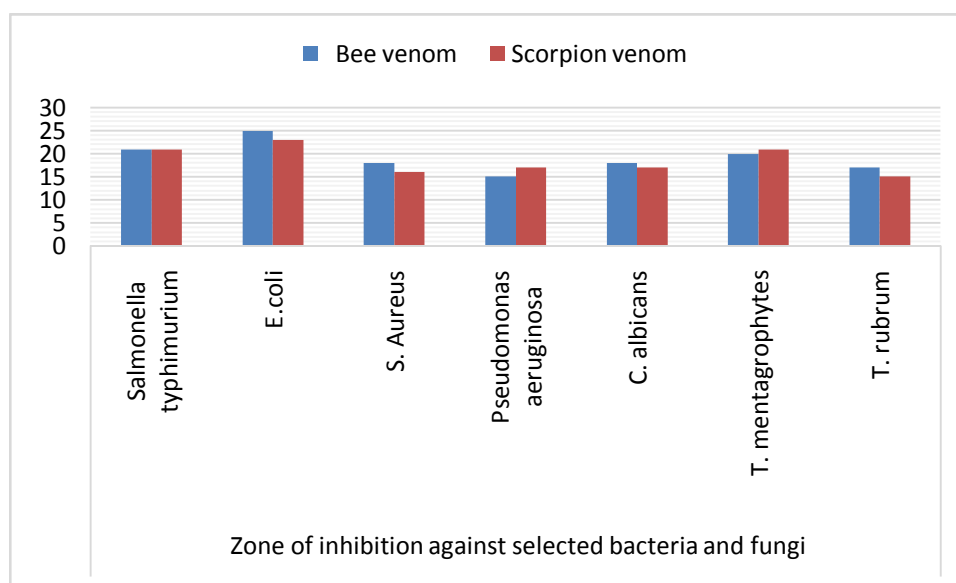


Figure 3: Comparative zone of inhibition of scorpion and bee venom against selected pathogens

DISCUSSION

Multidrug resistant pathogens are reported in high numbers in the world. This cause failure of the currently available antibiotics to treat these multidrug resistant pathogens. This lead to the global concern to discover alternative to antibiotics. Various researchers have conducted many studies about the biological activities of bee venom and scorpion venom. In the literature it is reported that the venom of bee and scorpion have some peptide which have antimicrobial activity. Bee venom and scorpion venom antimicrobial activity was tested against common selected pathogenic bacteria and fungi that includes *Staphylococcus aureus*, *Salmonella typhimurium*, *Escherichiacoli*, *Pseudomonas aeruginosa*, *Candida albicans*, *Trichophyton mentagrophytes* and *Trichophyton rubrum*. Disc diffusion method was used to evaluate the antimicrobial activity of these venom. Standard antibiotics and antifungal that includes Chloramphenicol, streptomycin and penicillin, fluconazole and amphotericin B were used as control for antimicrobial activity. Against all selected common pathogenic bacteria and fungi both of the venom of bee and scorpion shows antibacterial and antifungal activity. The bee venom give zone of inhibition 25mm, 18mm, 21mm 15mm against *E. coli*, *S. aureus*, and *Salmonella typhyimurium* and *Pseudomonas aeruginosa* respectively while the scorpion venom gives 23mm, 16mm, 21mm and 17mm zone of inhibition against *E. coli*, *S. aureus*, and *Salmonella typhyimurium* and *Pseudomonas aeruginosa* respectively. While in case of fungi the zone of inhibition of bee venoms against *C. albicans* *T. mentagrophytes* and *T. rubrum* was 18mm, 20mm and 17mm while scorpion venom gives 17mm, 21mm and 15mm zone of inhibition against *C. albicans* *T. mentagrophytes* and *T. rubrum* respectively.

In numerous studies effects of bee venom have been studied biochemically, anti-microbiologically and pharmacologically [43, 44]. The bee venom antimicrobial activity might be due to existence of numerous peptides like melittin, adolapin, apamin, mast-cell-degranulating peptides, various enzymes, amines that are active biologically and various components that are non-peptide [45]. A previous study done by Cujova *et al* [46] shows that melittin is present in bee venom which has more potential against gram positive bacteria as compared to gram negative bacteria. The sensitivity of the bacteria was measured by

measuring the inhibitory zone. Another study done by Ortel and Markwardt shows that gram positive bacteria have more sensitivity towards lower concentration of bee venom as compared to gram negative bacteria [47]. The bee venom antimicrobial activity might be due to existence of numerous peptides like melittin, adolapin, apamin, mast-cell-degranulating peptides, various enzymes, amines that are active biologically and various components that are non-peptide [48]. These components might have ability to interact with the molecules of some bacteria while other bacterial molecules may not have interaction. The antibacterial activity of Phospholipase A2 have been reported previously [49, 50]. The melittin in the bee venom have reported to have antibacterial activity. Depending on the antibacterial agents, various researchers have reported that against gram positive and gram negative bacteria the antimicrobial activity might be different [51]. Our study results are in accordance with the study done by Kondo and Kanai [52]. A previous study done by Hegazi *et al* is in contrast with our study. They reported that bee venom has less antimicrobial activity against *E. coli* [53]. A study done by Rybak *et al* shows that by mixing bee venom and kanamycin, it gives synergetic effect against *S. aureus* that are resistant to kanamycin [54]. Many infections are caused by fungi in the world like superficial skin infections and allergic problems etc. In addition, the major challenge to the public health is that antifungal agents are toxic and many fungus have been reported to be resistant to the available antifungal agents. On the other hand researchers are thinking about natural products like plant products, marine life, microbes and bee products to be used as antifungal agent because they have less side effect [55]. Currently against many fungal pathogens like *Trichophyton mentagrophytes* and *Trichophyton rubrum*, bee venom has been reported to be effective and can inhibit them [56]. The anti-fungal action of BV on Against 10 clinical isolates of *Candida albican*, bee venom for their antifungal activity has been test and MIC was calculated that ranges from 62.5-125 µgm/ml [55]. Previously another study reported the antifungal activity of melittin against numerous fungus and MIC was also determined that ranges between 30-300 µgm/ml (57). Furthermore, numerous amino peptides in scorpion venom have been reported that includes hadrurin [58], scorpine (59), opistoporins, parabutoporin [60]. These amino peptides shows cell lysis activity and also cause inhibition of various functions of microbes.

CONCLUSION

Multidrug resistant pathogens are reported in high numbers in the world. This cause failure of the currently available antibiotics to treat these multidrug resistant pathogens. This lead to the global concern to discover alternative to antibiotics. Various researchers have conducted many studies about the biological activities of bee venom and scorpion venom. In the literature it is reported that the venom of bee and scorpion have some peptide which have antimicrobial activity. Our study shows that both bee venom and scorpion venom have the ability to inhibit bacterial and fungi which may be used complementary to antibiotics. Therefore, our study might conclude that specific mechanism, which is not well known, is used by bee venom and scorpion venom to inhibit the growth of both bacteria and fungi. Further in vivo and in vitro study based on a chemical, pharmacological, and clinical approach must be conducted to understand the exact mechanism of these venoms.

REFERENCES

1. Palfy R, Gardlik R, Behuliak M, Kadasi L, Turna J & Celec P. (2002). On the physiology and pathophysiology of antimicrobial peptides. *Molecular Medicine*; 15 (1-2), 51-59.
2. Allen HK, Donato J, Wang HH, Cloud-Hansen KA, Davies J, et al. (2010) Call of the wild: antibiotic resistance genes in natural environments. *Nat Rev Microbiol* 8: 251-259.
3. Andersson DI, Hughes D (2010) Antibiotic resistance and its cost: is it possible to reverse resistance? *Nat Rev Microbiol* 8: 260-271.
4. Hebert C, Weber SG (2011) Common approaches to the control of multidrug-resistant organisms other than methicillin-resistant *Staphylococcus aureus* (MRSA). *Infect Dis Clin North Am* 25: 181-200.
5. Ippolito G, Leone S, Lauria FN, Nicastrì E, Wenzel RP (2010) Methicillin-resistant *Staphylococcus aureus*: the superbug. *Int J Infect Dis* 14 Suppl 4: S7- 11.
6. Brogden NK, Brogden KA (2011) Will new generations of modified antimicrobial peptides improve their potential as pharmaceuticals? *Int J Antimicrob Agents* 38: 217-225.
7. Chin YW, Balunas MJ, Chai HB and Kinghorn AD. (2006). Drug discovery from natural sources. *AAPS J* 8: 239-253.
8. Alves RR and Alves HN. 2011. The faunal drugstore: Animal-based remedies used in traditional medicines in Latin America. *J Ethnobiol Ethnomed* 7: 9.
9. Trowell S. (2003). Drugs from bugs: the promise of pharmaceutical entomology. *Futurist* 37: 17-19.
10. Zasloff, M. (2002) Antimicrobial peptides of multicellular organism. *Nature* 415, 389-395.
11. Moerman, L., Bosteels, S., Noppe, W., Willems, J., Clynen, E., Schoofs, L., Thevissen, K., Tytgat, J. et al. (2002) Antibacterial and antifungal properties of α -helical, cationic peptides in the venom of scorpions from the South Africa. *Eur J Biochem* 269, 4799-4810.

12. Corzo, G., Escoubas, P., Villegas, E., Barnham, K.J., He, W., Norton, R.S. and Nakajima, T. (2001) Characterization of unique amphipathic antimicrobial peptides from venom of the scorpion *Pandinus imperator*. *J Biochem* 359, 35–45.
13. Xu, K., Ji, Y. and Qu, X. (1989) Purification and characterization of an antibacterial peptide from venom of *Lycosa singoriensis*. *Acta Zool Sinica* 35, 300–305.
14. Fennel, J.E., Shipman, W.H. and Cole, L.J. (1968) Antibacterial action of melittin, a polypeptide from the venom. *Proc Soc Exp Biol Med* 127, 707–710.
15. Blaylock, R. (2000) Antibacterial properties of KwaZulu natal snake venoms. *Toxicon* 38, 1529–1534.
16. Boutrin MC, Foster HA and Pentreath VW. (2008). The effects of bee (*Apis mellifera*) venom phospholipase A2 on *Trypanosoma brucei* and enterobacteria. *Exp Parasitol* 119: 246-251.
17. Alia O, Laila M and Antonious A. (2013). Antimicrobial effect of melittin isolated from Syrian honeybee (*Apismellifera*) venom and its wound healing potential. *Int J Pharm Sci Rev Res* 21: 318-324.
18. Varanda EA and Tavares DC. (1998). Radioprotection: mechanisms and radioprotective agents including honeybee venom. *J Venom Anim Toxins* 4: 5-21.
19. Perumal Samy R, Gopalakrishnakone P, Thwin MM, Chow TK, Bow H, Yap EH and Thong TW. (2007). Antibacterial activity of snake, scorpion and bee venoms: a comparison with purified venom phospholipase A2 enzymes. *J Appl Microbiol* 102: 650-659.
20. Lima PR and Brochetto-Braga MR. (2003). Hymenoptera venom review focusing on *Apis mellifera*. *J Venom Anim Toxins Incl Trop Dis* 9: 149-162.
21. Samel M, Vija H, Kurvet I, Künnis-Beres K, Trummal K, Subbi J, Kahru A And Siigur J. (2013). Interactions of PLA2-s from *Vipera lebetina*, *Vipera berus berus* and *Naja naja oxiana* venom with platelets, bacterial and cancer cells. *Toxins* 24: 203-223.
22. Schaloske RH and Dennis EA. (2006). The phospholipase A2 superfamily and its group numbering system. *Biochim Biophys Acta* 1761: 1246-1259.
23. Burke JE and Dennis EA. (2009). Phospholipase A2 structure/ function, mechanism, and signaling. *J Lipid Res* 59:S237-S242.
24. Van Deenen LLM and DE Haas GH. (1963). The substrate specificity of phospholipase A. *Biochim Biophys Acta* 70: 538-553.
25. Nevalainen TJ, Graham GG and Scott KF. (2008). Antibacterial actions of secreted phospholipases A2. *Biochim Biophys Acta* 1781: 1-9.
26. Pandey BK, Ahmad A, Asthana N, Azmi S, Srivastava RM, Srivastava S, Verma R, Vishwakarma AL and Ghosh JK. (2010). Cell-selective lysis by novel analogues of melittin against human red blood cells and *Escherichia coli*. *Biochemistry* 49: 7920-7929.
27. Zhu WL, Nan YH, Hahm KS and Shin SY. (2007). Cell selectivity of an antimicrobial peptide melittin diastereomer with D-amino acid in the leucine zipper sequence. *J Biochem Mol Biol* 40: 1090-1094.
28. Carvalho LAC and Machini MT. (2013). Hemocidinas derivadas da hemoglobina: Estruturas, propriedades e perspectivas. *Quim Nova* 7: 1021-1029.
29. Raghuraman H and Chattopadhyay A. (2007). Melittin: a membrane-active peptide with diverse functions. *Biosci Rep* 27: 189-223.
30. Wang C, Chen T, Zhang N, Yang M, Li B, Lü X, Cao X And Ling C. (2009). Melittin, a major component of bee venom, sensitizes human hepatocellular carcinoma cells to tumor necrosis factor-related apoptosis-inducing ligand (trail)-induced apoptosis by activating CaMKIITAK1- JNK/p38 and inhibiting IκappaBα kinase-NF kappaB. *J Chem Biol* 284: 3804-3813.
31. Murakami M, Taketomi Y, Miki Y, Sato H, Hirabayashi T and Yamamoto K. (2011). Recent progress in phospholipase A2 research: From cells to animals to humans. *Progr Lipid Res* 50: 152-192.
32. Almaaytah A, Tarazi S, Mhaidat N, Al-Balas Q, Mukattash TL. Mauriporin, (2013). A novel cationic α-helical peptide with selective cytotoxic activity against prostate cancer cell lines from the venom of the scorpion *Androctonus mauritanicus*. *Int J Peptide Res Ther*;19, 291–293.
33. Almaaytah A, Tarazi S, Alsheyab F, Al-Balas Q, Mukattash T. (2014). Antimicrobial and Antibiofilm Activity of Mauriporin, a Multifunctional Scorpion Venom Peptide. *Int J PepResTher* ;20 (4), 397-408.
34. Goudet C, Chi CW, Tytgat J. (2002). An overview of toxins and genes from the venom of the Asian scorpion *Buthus martensii* Karsch. *Toxicon*;40, 1239–1258.
35. Almaaytah A, Zhou M, Wang L, Chen T, Walker B, Shaw C. (2012). Antimicrobial/cytolytic peptides from the venom of the North African scorpion, *Androctonus amoreuxi*: biochemical and functional characterization of natural peptides and a single sitesubstituted analog. *Peptides* ; 35, 291–299
36. Jenssen, H.; Hamill, P.; Hancock, R.E.W. Peptide antimicrobial agents. *Clin. Microbiol. Rev.* (2006), 19, 491–511
37. Almaaytah A, Tarazi S, Abu-Alhajja, A., Altall Y, Alshar' I N et al. Enhanced Antimicrobial Activity of AamAP1-Lysine, a Novel Synthetic Peptide Analog Derived from the Scorpion Venom Peptide AamAP1. *Pharmaceuticals* 2014; 7(5), 502-516
38. Zeng X, Corzo G, Hahin R. (2005). Scorpion venom peptides without disulfide bridges. *IUBMB Life*;57 (1), 13–21.
39. Zeng XC, Li WX, Peng F, Zhu ZH. (2000). Cloning and characterization of a novel cDNA sequence encoding the precursor of a novel venom peptide (BmKbpp) related to a bradykinin-potentiating peptide from Chinese scorpion *Buthus martensii* Karsch. *IUBMB Life* ; 49, 207–210.
40. Rodriguez de la Vega RC, Schwartz EF, Possani LD. (2010). Mining on scorpion venom biodiversity. *Toxicon* ;56:1155–61.

41. Bauer, A.W., Kirby, W.M., Sherris, J.C. and Turck, M. (1966) Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Path* 45, 493–496.
42. NCCLS (2002). Performance Standards for Antimicrobial Susceptibility Testing, Twelfth Informational Supplement. NCCLS Document no. M100-S12. Villanova, PA: National Committee for Clinical Laboratory Standards.
43. Park JW, Jeon JH, Yoon J, Jung TY, Kwon KR, Cho CK, *et al.* (2012). Effects of sweet bee venom pharmacopuncture treatment for chemotherapy-induced peripheral neuropathy: a case series. *Integr Cancer Ther.* 11(2):166-71.
44. Kim KS, Choi US, Lee SD, Kim KH, Chung KH, Chang YC, *et al.* (2005). Effect of bee venom on aromatase expression and activity in leukaemic FLG 29.1 and primary osteoblastic cells. *J Ethnopharmacol.* 99(2):245-52.
45. Leandro LF, Mendes CA, Casemiro LA, Vinholis AH, Cunha WR, de Almeida R, *et al.* (2015). Antimicrobial activity of apitoxin, melittin and phospholipase A₂ of honey bee (*Apis mellifera*) venom against oral pathogens. *An Acad Bras Cienc.* 87(1):147-55.
46. Cujova S, Bednarova L, Slaninova J, Straka J, Cerovsky V.(2014). Interaction of a novel antimicrobial peptide isolated from the venom of solitary bee *colletes daviesanus* with phospholipid vesicles and *Escherichia coli* cells. *J Pept Sci.*;20(11):885-95.
47. Ortel S, Markwardt F. (1955). Studies on the antibacterial properties of bee venom. *Pharmazie.* 10:743-6.
48. Riviere WR, Melzack R. The bee venom test: a new tonic-pain test. *Pain.* 1996;66(2-3):271 7.
49. Nunez V, Arce V, Gutierrez JM, Lomonte B. Structure and functional characterization of myotoxin I, a LYS49 phospholipase A2 homologous from the snake *Bothrops atrox*. *Toxicon.* 2004;44(1):91-101
50. Permul SR, Gopalakrishnakone P, Thwin MM, Chow TK, Bow H, Yap EH, *et al.* (2007). Antibacterial activity of snake, scorpion and bee venoms: a comparison with purified venom phospholipase A2 enzymes. *J Appl Microbiol.* 2007;102(3):650-9.
51. Monk JD, Beuchat LR, Hathcox AK. (1996). Inhibitory effects of sucrose monolaurate, alone and in combination with organic acids, on *Listeria monocytogenes* and *Staphylococcus aureus*. *J Appl Bacteriol.* ;81(1):7-18.
52. Kondo E, Kanai K. (1986). Bactericidal activity of the membrane fraction isolated from phagocytes of mice and its stimulation by melittin. *Japan J Med Sci & Biol.* 39:9-20.
53. Hegazi AG, Moharm NZ, Allah FA, Nour MS, Khair AM. (2002). Antibacterial activity of different Egyptian honeys in relation to some bee products. *Egypt J Vet Sci.* 36:31- 42.
54. Rybak CH, Szczesna T, Rybak M, Pidek A. (1994). Some properties of honey bee venom. *Pszczelnicze Zeszyty Naukowe.* 1994;38:85-90.
55. Lee, S.-B. (2016). Antifungal activity of bee venom and sweet bee venom against clinically isolated *Candida albicans*. *J. Pharmacopunct.* 19, 45–50.
56. Yu, A.R.; Kim, J.J.; Park, G.S.; Oh, S.M.; Han, C.S.; Lee, M.Y. (2012). The antifungal activity of bee venom against dermatophytes. *J. Appl. Biol. Chem.* 55, 7–11.
57. AL-Ani, I.; Zimmermann, S.; Reichling, J.; Wink, M. (2000). Pharmacological synergism of bee venom and melittin with antibiotics and plant secondary metabolites against multi-drug resistant microbial pathogens. 165–168.
58. Conde R, Zamudio FZ, Rodriguez MH, Possani LD (2000) Scorpine, an antimalarial and anti-bacterial agent purified from scorpion venom. *FEBS Lett* 471: 165–168.
59. Moerman L, Bosteels S, Noppe W, Willems J, Clynen E, *et al.* (2002) Antibacterial and antifungal properties of alpha-helical, cationic peptides in the venom of scorpions from southern Africa. *Eur J Biochem* 269: 4799–4810.
60. Dai L, Yasuda A, Naoki H, Corzo G, Andriantsiferana M, *et al.* (2001) IsCT, a novel cytotoxic linear peptide from scorpion *Opisthacanthus madagascariensis*. *Biochem Biophys Res Commun* 286: 820–825.

CITATION OF THIS ARTICLE

Farmanullah, K Abbasi, H M Azam, N U Mandokhail, F Iqbal, F Khan, M A Rafique, H Saeed, F Amin, J Mughal, M Ali, J Ahmad. Antimicrobial activities of scorpion and honey bee venom against some common selected pathogen. *Bull. Env. Pharmacol. Life Sci.*, Vol 9[10] September 2020 : 87-94