Bulletin of Environment, Pharmacology and Life Sciences Bull. Env. Pharmacol. Life Sci., Vol 8 [2] January 2019 : 93-96 ©2019 Academy for Environment and Life Sciences, India Online ISSN 2277-1808 Journal's URL:http://www.bepls.com CODEN: BEPLAD Global Impact Factor 0.876 Universal Impact Factor 0.9804 NAAS Rating 4.95

ORIGINAL ARTICLE



OPEN ACCESS

Screening of cattle population for Bovine Tuberculosis using **Single Intradermal Tuberculin Test**

*Neelam Rani¹, P. K. Kapoor², Naresh Jindal¹, Rajesh Chhabra³ And Piyush Tomar¹

¹Department of Veterinary Public Health and Epidemiology, LUVAS, Hisar ²Registrar, Bihar Animal Sciences University, Patna ³College Central Laboratory, LUVAS, Hisar *Corresponding author: neelamvet2011@gmail.com

ABSTRACT

Bovine tuberculosis (BTB) is a chronic bacterial disease of cattle, caused by Mycobacterium bovis (M. bovis). The OIE (World Organization for Animal Health) recommended test for screening of bovine tuberculosis is Single Intradermal Tuberculin Test (SIDT). In the current study, a total of 82 randomly selected cattle from four gaushalas of Haryana were screened by SIDT. Out of these, 6/82 (7.31%) animals were found to be doubtful/ inconclusive by SIDT using bovine tuberculin. From this study, it could be concluded that bovine tuberculosis is prevalent in Haryana and need to be further investigated by other modern diagnostic tools like interferon gamma assay (IFN-y assay) and DNA based techniques. Keywords: Economic, Mycobacterium bovis, Screening, Tuberculin, Zoonotic.

Received 21.09.2018

Revised 19.11.2018

Accepted 30.12.2018

INTRODUCTION

Bovine tuberculosis (BTB) is an OIE (World Organization for Animal Health) listed chronic, debilitating, bacterial disease of livestock, wildlife and humans, caused by Mycobacterium bovis (M. bovis) and is economically significant because of trade restriction and its zoonotic perspective [1]. It has been estimated that *M. bovis* might infected over 50 million cattle worldwide with resulting economic losses of approximately \$3 billion [2]. A wide range of animals is susceptible to *M. bovis* infection making its eradication difficult [3].

There are many available diagnostic tests for bovine tuberculosis but tuberculin skin test (TST) is the OIE recommended test for screening against TB [4]. Limitations in the sensitivity and specificity of tuberculin testing result in a failure to detect all *M. bovis* infected animals and contribute significantly to disease persistence [5]. The tuberculin used for in vivo tuberculin test in cattle contains a crude mixture of mycobacterial secreted proteins prepared by precipitation of heat killed cultures. Despite of limitations of tuberculin skin test, it is widely used for screening purpose. Traditional test and slaughter/segregation policies based on tuberculin skin testing have not been fully successful so that additional more sensitive and specific diagnostic tests are required [6]. The current study was performed on randomly selected 82 cattle of four different gaushalas of Haryana to screen for bovine tuberculosis by SIDT.

MATERIAL AND METHODS

The study was approved by the institutional animal ethics committee. The present study was conducted on randomly selected 82 cattle from four different gaushalas of Haryana. These animals were screened by single intradermal tuberculin testing (SIDT) using bovine origin purified protein derivative (PPD-B) (IVRI, Izatnagar, U.P.) using standard protocol [4] with the exception of calves of less than 6 months age, animals of more than 8 months pregnancy and one month post-partum animals [7]. The animals were categorized on the basis of breed, age, gender and location (Table 1). Briefly, before PPD-B injection, the injection site of neck region was shaved with sterile razor and cleaned properly. Skin thickness of the neck region of each cattle was measured prior to tuberculin injection with Vernier caliper. A single dose of 0.1 ml (2000 IU) PPD-B (1 mg/ml) was administered intradermally using tuberculin syringe having a

Rani *et al*

short needle. Skin thickness at the injection site was measured with Vernier Caliper after 72 hrs. of PPD administration. The SIDT was considered positive if the increase in skin thickness at the site of injection was 4 mm or more than 4 mm, inconclusive if the increase in skin thickness was more than 2 mm and less than 4 mm and negative if the increase in skin thickness at site of injection was less than 2 mm.

Variables	No. of cattle screened
Breed	
Cross-bred	39
Hariana	26
Rathi	2
Sahiwal	15
Location	
А	25
В	19
С	19
D	19
Gender	
Male	19
Female	63
Age	
Heifer	0
Adult	82
Total cattle screened by SIDT	82

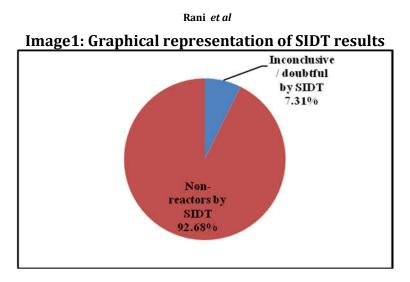
Table 1: Categorizatio	n of animals under study

RESULTS

Out of 82 cattle, 6 (7.31%) were found doubtful/inconclusive as the difference in skin thickness before and after PPD-B administration was more than 2 mm but less than 4 mm. Out of these six cattle, four cattle (4.87%) were from gaushala 'A', one cattle (1.21%) from gaushala 'B' and one cattle (1.21%) from gaushala 'C'. None of the tested cattle from gaushala 'D' was found positive by SIDT. A total of five female cattle (6.09%) and one male (1.21%) animal (cow bull) were found positive by SIDT. Out of these six cattle, five (6.09%) were crossbreds and one (1.21%) was Hariana cattle. All the six doubtful/ inconclusive cattle by SIDT were adult animals (Table 2, Image 1).

Variables	No. of cattle found reactor/doubtful by
	SIDT (%)
Breed	
Cross-bred	5 (6.09%)
Hariana	1 (1.21%)
Rathi	0
Sahiwal	0
Location	·
А	4 (4.87%)
В	1 (1.21%)
С	1 (1.21%)
D	0
Gender	
Male	1 (1.21%)
Female	5 (6.09%)
Age	
Heifer	0
Adult	6 (7.31%)
Total cattle found	6 (7.31%)
inconclusive/doubtful by SIDT	
Total cattle found non-reactor by	76 (92.68%)
SIDT	

Table 2: Categorization of animals in relation to positivity (%) to SIDT



DISCUSSION

Similar to the present study, it was reported that 2.8% animals reacted to the tuberculin test (8) and low prevalence (1.93%) was also reported in cattle from Maharashtra (9). Similarly, in Morogoro, the prevalence was 1.3% by single intra-dermal comparative cervical tuberculin (SICCT) test (10) and in another study, an individual prevalence of 3.7% was estimated using SIDT test in Morogoro (11). The estimated prevalence of TB in animal population of Charsadda (Pakistan) by tuberculin skin testing (TST) found was 4.33% (12).

However, in Tamil Nadu and Karnataka, the prevalence rate was 34.58% and 30-35%, respectively (13). In Himachal Pradesh, BTB prevalence in cattle found was 14.31% by SIDT testing and 20% prevalence by comparative cervical tuberculin (CCT) test (14) and in U. P., 13.12% animals were found positive by SIDT testing (15). In a study of West Bengal, a total of 25.4% animals from organized and 3.2% from backyard farming sector were found positive by tuberculin testing (16).

In the current study, the prevalence of BTB by SIDT testing was very low. The possible explanation for low prevalence of BTB by SIDT test could be early infection, dormant infection, presence of other environmental mycobacteria, immune response of the host, environmental stress factors, poor nutritional status *etc.* It can be speculated that animals with dormant infections fail to respond to PPD stimulation or repeated testing of animals with PPD may increase the number of animals failing to respond (17).

The current findings could also be supported with the findings that the infection with *M. avium* subsp. *paratuberculosis* interferes in the diagnosis of BTB by SIDT test (18). It has been reported that infection with either *Fasciola* spp. or *Stronglus* spp. significantly reduces the skin indurations in response to PPD-B in *M. bovis*-infected heifers (19).

CONCLUSIONS

The complex immune response of cattle to infection with *M. bovis* predisposes difficulties in diagnosis. These can best be dealt with by identifying the herd rather than the individual animal, and using combinational approach *i.e.* gamma interferon gamma assay, nucleic acid based techniques for diagnosis, to quantify the disease burden. The strategic use of IFN- γ assay, as an adjunct to the tuberculin skin test, can facilitate the early removal of the infected animals that are otherwise negative to the tuberculin skin test.

REFERENCES

- 1. WHO, (2011). Global tuberculosis control report. Geneva: World Health Organization. Retrieved from: http://www.who.int/tb/publications/globalreport/2011/gtbr11.
- 2. Steele, J.H. (1995). Regional and country status report, p. 169-172. *In* C. O. Thoen and J. H. Steele (ed.), *Mycobacterium bovis* infection in animals and humans. Iowa State University Press, Ames.
- 3. Delahay, R.J, Smith, G.C, Barlow, A.M., Walker, N. and Harris, A. (2006). Bovine tuberculosis infection in wild mammals in the South-West region of England: A survey of prevalence and a semi-quantitative assessment of the relative risks to cattle. *Vet. J.***173(2)**:287–301.
- 4. OIE (2009). Bovine tuberculosis. In: Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, 6th ed. Office International des Epizooties, Paris (updated 14.8.09), Chapter 2.4.7, pp. 1-16.
- 5. Monaghan, M., Doherty, M., Collins, J., Kazda, J. and Quinn, P. (1994). The tuberculin test. *Vet. Microbiol.***40**: 111–124.
- 6. Pollock, J.M., Welsh, M.D. and McNair, J. (2005). Immune responses in bovine tuberculosis: Towards new strategies for the diagnosis and control of disease. *Vet Immunol.***108**:37-43.

Rani *et al*

- 7. Proano-Perez, F., Benitez-Ortiz, W., Celi-Erazo, M., Ron-Garrido, L., Benitez- Capistros, R., Portaels, F.Rigouts, L. and Linden, A. (2009). Comparative Intradermal Tuberculin Test in Dairy Cattle in the North of Ecuador and Risk Factors Associated with Bovine Tuberculosis. *Am J Trop Med Hyg.***81(6)**:1103–1109.
- 8. Danbirni, S, Okaiyeto, S.O, Bature, C. and Moris, A. (2013). Field Determination of Tuberculosis Prevalence in a Herd of Cattle Using Tuberculin and Quicking® Bovine Tuberculosis Antibody Rapid Tests in Jalingo, Nigeria. *J Vet Adv.***3(1):** 20-23.
- 9. Lall, J.M. (1969). Tuberculosis among animals in India. Vet bull. 39(6):680-685.
- 10. Shirima, G. M., Kazwala, R.R. and Kambarage, D.M. (2003). Prevalence of bovine tuberculosis in cattle in different farming systems in the eastern zone of Tanzania. *Journal of Preventive Vet. Med.***57(3):** 167-172.
- 11. Mwakapuja, R. S., Zachariah, E. M., Joseph, M., Ward, B., Robinson, H. M., Irmgard, M., Rudovick, R. K. and Manfred, T. (2013). Prevalence and significant geospatial clusters of bovine tuberculosis infection at livestock wildlife interface ecosystem in Eastern Tanzania. *Tropical Animal Health Production*.**151(13)**: 1472 -1492.
- 12. Noorrahim, M., Khan, S., Muhammad, S., Alamgir, S., Muzafar, S. and Rafiullah, H. A. (2015). Prevalence of tuberculosis in livestock population of district Charsadda by tuberculin skin test (TST). *JEZS.***3(2)**:15-19.
- 13. Dhinakaran, M, Rao, V.N. and Nedunchellian, S. (1991). Immunological Response in Tuberculin Reactor Cattle. In: Proceedings of the National Symposium on Recent Advances in Control of Diseases of Crossbred and Companion Animals. Mumbai, India: Konkan Krishi Vidyapeeth. Bombay Veterinary College, Department of Medicine.
- 14. Thakur, A., Sharma, M., Katoch, V., Dhar, C. Prasenjit and Katoch, R.C. (2010). A study on the prevalence of Bovine Tuberculosis in farmed dairy cattle in Himachal Pradesh. *Vet World.* **3(9)**:409-414.
- 15. Thakur, M. K., Sinha, D. K. and Singh, B. R. (2016). Evaluation of complementary diagnostic tools for bovine tuberculosis detection in dairy herds from India. *Vet. World.* **9(8)**:862-868.
- 16. Das, R., Dandapat, P., Chakrabarty, A., Nanda, P.K., Bandyopadhyay, S. and Bandyopadhyay, S. (2018). A crosssectional study on prevalence of bovine tuberculosis in Indian and crossbred cattle in Gangetic delta region of West Bengal, India. *Int. J. of One Health.***4**:1-7.
- 17. Thoen, C.O. and Bloom, B.R. (1995). Pathogenesis of *Mycobacterium bovis* infection in animals and humansedited by Charles O. Thoen & James H. Steele. Iowa State University Press, pp.3-14.
- 18. Aranaz, A., De Juan, L., Bezos, J., Alvarez, J., Romero, B. and Lozano, F. (2006). Assessment of diagnostic tools for eradication of bovine tuberculosis in cattle co-infected with *Mycobacterium bovis* and *M. avium* subsp. *paratuberculosis. Vet Res.***37:**593-606.
- 19. Ameni, G., Aseffa, A., Engers, H., Young, D., Hewinson, G. and Vordermeier, M. (2006). Cattle husbandry in Ethiopia is a predominant factor affecting the pathology of bovine tuberculosis and gamma interferon responses to mycobacterial antigens. *Clin. Vaccine Immunol*.**13**:1030-1036.

CITATION OF THIS ARTICLE

Neelam Rani, P. K. Kapoor, Naresh Jindal, Rajesh Chhabra And Piyush Tomar. Screening of cattle population for Bovine Tuberculosis using Single Intradermal Tuberculin Test. Bull. Env. Pharmacol. Life Sci., Vol 8 [2] January 2019: 93-96